



# **Flavonols in Action: Targeting Oxidative Stress and Neuroinflammation in Major Depressive Disorder**

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Abstract: Major depressive disorder is one of the most common mental illnesses that highly impairs quality of life. Pharmacological interventions are mainly focused on altered monoamine neurotransmission, which is considered the primary event underlying the disease's etiology. However, many other neuropathological mechanisms that contribute to the disease's progression and clinical symptoms have been identified. These include oxidative stress, neuroinflammation, hippocampal atrophy, reduced synaptic plasticity and neurogenesis, the depletion of neurotrophic factors, and the dysfunction of the hypothalamic-pituitary-adrenal (HPA) axis. Current therapeutic options are often unsatisfactory and associated with adverse effects. This review highlights the most relevant findings concerning the role of flavonols, a ubiquitous class of flavonoids in the human diet, as potential antidepressant agents. In general, flavonols are considered to be both an effective and safe therapeutic option in the management of depression, which is largely based on their prominent antioxidative and anti-inflammatory effects. Moreover, preclinical studies have provided evidence that they are capable of restoring the neuroendocrine control of the HPA axis, promoting neurogenesis, and alleviating depressive-like behavior. Although these findings are promising, they are still far from being implemented in clinical practice. Hence, further studies are needed to more comprehensively evaluate the potential of flavonols with respect to the improvement of clinical signs of depression.

Keywords: depression; flavonols; neuroinflammation; oxidative stress; HPA axis

## 1. Introduction

Major depressive disorder is a severe neuropsychiatric disease driven by various hereditary, environmental, and psychological factors [1]. It disables normal functioning and quality of life. Typical symptoms include ongoing sadness, feelings of guilt, fatigue, loss of energy, lack of motivation for engaging in previously rewarding activities, impairment of cognitive functions, and disturbances of sleep, weight, and appetite. Depression is a common mental disorder and is one of the leading causes of disability worldwide. Globally, more than 350 million people suffer from depression, with the prevalence being higher among women [2,3].

The clinical manifestations of depression are accompanied by morphological changes of specific brain regions mainly related to mood regulation, rewards, emotions, and cognitive functions (such as decision making and memory). Neuroimaging studies have revealed significant structural alterations, together with abnormal interregional connectivity and brain activity, in various cortical and limbic brain areas. Affected areas include the frontal lobe (e.g., the orbitofrontal and dorsolateral prefrontal cortexes), the hippocampus, the thalamus, the basal ganglia, the amygdala, and the anterior cingulate cortex as well as cerebellar neurons projecting toward the ventral tegmental area (VTA) [4–9]. However, due to the diversity of demographic and clinical data, methodologies used, and study designs,



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**Copyright:** © 2023 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). the results concerning the identification of the depression-related patterns of brain changes are very heterogeneous and far from generalization.

## 2. Neuropathological Mechanisms Underlying the Development of Depression

## 2.1. Monoamine Hypothesis of Depression

The mechanisms underlying the etiology of depression are still not fully understood. From a clinical perspective, the typical abnormality witnessed is a disruption of neurotransmitter levels, particularly those of serotonin, dopamine, and norepinephrine. Depressed patients have reduced levels of monoamines and their metabolites in the cerebrospinal fluid and the post-mortem brain, although the results on this matter are not completely uniform and conclusive [10-13]. The major mechanisms related to monoamine depletion include the impaired activity of enzymes participating in their degradation, the impaired activity of enzymes involved in tryptophan synthesis, and the impaired activity of monoamine transporters and receptors [14–16]. Moreover, most of the behavioral symptoms of depressed patients can be related to disruptions in monoamine levels [17]. Hence, the monoamine hypothesis contends that monoamine deficiency is the core mechanism underlying the pathophysiological events of depression [18]. This hypothesis is supported by the mechanisms of action of clinically used antidepressants. The main pharmacological approaches are based on improving the monoamine balance by using selective serotonin reuptake inhibitors (SSRIs); tricyclic antidepressants that inhibit the reuptake of serotonin, dopamine, and norepinephrine; monoamine oxidase inhibitors; and selective norepinephrine reuptake inhibitors (NARI) [19,20]. The first-line drugs employed are usually SSPIs such as fluoxetine, sertraline, and citalopram, which slowly increase synaptic serotonin levels. However, despite this repertoire of available drugs, many patients do not respond adequately to such therapy [21], and many unwanted side effects are induced alongside the beneficial ones. Some of the adverse effects reported by patients include dizziness, sedation, anxiety, cardiac problems, gastrointestinal and sexual disfunction, sleep disturbances, appetite changes, forgetfulness, confusion, difficulty finding words, and other cognitive impairments [22].

Besides the limited efficacy and slow onset of the most-prescribed drugs, monoamine depletion neither causes depression in healthy individuals nor worsens symptoms in patients not undergoing therapy, thereby raising questions concerning the exact role of monoamine deficiency in the etiopathology of depression and suggesting the involvement of other pathological mechanisms [18,23]. More recent studies have demonstrated neurochemical, structural, and functional impairments of GABAergic and glutamatergic systems. As will be explained below, these dysfunctions are likely mediated by excitotoxicity and increased levels of pro-inflammatory cytokines and adrenal glucocorticoids in combination with certain environmental factors [24]. Evidence revealing that the impaired function of GABAergic and glutamatergic systems is implicated in the pathophysiology of depression resulted in advancements in therapeutic strategies, particularly with respect to the treatment-resistant depression, leading to the introduction of fast-acting antidepressants targeting N-methyl-D-aspartate (NMDA) and  $\gamma$ -aminobutyric acid type A (GABA<sub>A</sub>) receptors [24–26].

## 2.2. Hippocampus and Depression

Brain imaging has revealed lesions mainly localized in the prefrontal and cingulate cortex, hippocampus, and amygdala [8,27], indicating increased susceptibility to neuronal death in distinct brain areas. Structural and functional alterations of the hippocampus are particularly relevant considering this structure's role in the regulation of stress response, memory-related consolidation processes, and adult neurogenesis. Depressed patients have reduced hippocampal volume due to a decreased number of both neuronal and glial cells, suggesting that hippocampal changes are likely a contributing factor in the pathophysiology of depression [4,28,29]. At the cellular and molecular level, it is believed that the dysregulation of the HPA axis and the consequent increase in the quantity of glucocorticoids are the underlying mechanisms of reduced neurogenesis, excitotoxicity,

inflammation, and depleted levels of neurotrophins, which all contribute to reducing the size of the hippocampus [4].

The severity of hippocampal degeneration positively correlates with the duration of the symptoms and negatively correlates with the efficacy of antidepressant therapies and clinical outcomes [30–32]. Boku et al. proposed two hypotheses for explaining changes in the hippocampal volume [33]. The neuroplasticity hypothesis emphasizes the degenerative changes of mature neurons in the hippocampus (the reduced branching and length of dendrites and the reduced density of dendritic spines), whereas the neurogenesis hypothesis considers the reduced neurogenesis and the role of the neural progenitor cells in the dentate gyrus. These hippocampal deteriorations are considered to be largely responsible for delayed responses to antidepressant therapy. It has been shown that only long-term treatment with antidepressants protects mature neurons and promotes the proliferation of neural progenitors and neurogenesis. At least partially, the protection of mature neurons is stimulated by glucocorticoid-receptor-initiated pathways [35]. Thus, the time needed to observe effects on hippocampal neurons corresponds with the therapeutic window required to observe the improvement of clinical manifestations [28,29].

## Adult Neurogenesis and Depression

The process of neurogenesis is tightly related to neurotrophic factors. Brain-derived neurotrophic factor (BDNF) belongs to the neurotrophin family and is one of the most abundant neurotrophins in the brain. BDNF acts via the tropomyosin receptor kinase B (TrkB) receptor, which is expressed on both neuros and glia. The activation of TrkB receptors triggers the activation of signaling cascades involved in neuronal survival and functioning, including the mitogen-activated protein kinase (MAPK), phosphatidylinositol 3-kinase (PI3K)/Akt, and phospholipase C- $\gamma$  (PLC- $\gamma$ ) pathways [36,37]. BDNF plays an important role in the wide spectrum of neurophysiological processes. Among other functions, it regulates neuronal development and survival, synaptogenesis, plasticity, and learning and memory [38–41]. Different BDNF roles are mediated by distinct transcripts that arise from the nine promotors in the *BDNF* gene, which all encode the same BDNF protein [42]. These transcripts are expressed in a complex spatio-temporal pattern, respond to different stimuli, and trigger the activation of distinct signaling networks [43,44]. Hence, depending on the brain region, the effects of BDNF could be antidepressant or pro-depressant. In the hippocampus and prefrontal cortex, BDNF displays antidepressant activity, and depressed patients usually exhibit reduced serum BDNF levels and levels of BDNF IV transcript in the hippocampus and prefrontal cortex [45–47]. The disruption of murine promotor IV impairs GABAergic transmission and the expression of monoaminergic genes in the hippocampus and prefrontal cortex and results in depression-like behavior [48–50]. Moreover, there are indications that the methylation of promotor IV and plasma BDNF levels can be used as a potential predictor of treatment response [51]. It has been proposed that serum BDNF concentrations may serve as the peripheral biomarker of the disease [45], yet its levels in the serum do not always correlate with the severity of symptoms [52–54]. However, BDNF concentrations increase after antidepressant therapy, and there is a good correlation between BDNF changes and improvements in depression scores [55,56].

## 2.3. HPA Axis and Depression

HPA hyperactivity is commonly observed in patients with chronic stress and depression [57–59]. The hippocampus possesses many receptors for glucocorticoids. Through a negative feedback mechanism, it regulates glucocorticoid release from the adrenal cortex, thus playing a critical role in the tuning of the HPA axis [60,61]. The HPA axis starts with the neurosecretory cells of the hypothalamus that release corticotrophin-releasing hormone (CRH). CRH stimulates the release of the adrenocorticotropic hormone (ACTH) from the anterior pituitary, whereas ACTH further stimulates the adrenal gland so that it both produces and releases glucocorticoids. In turn, through inhibitory feedback, the released glucocorticoids attenuate the production of CRH and ACTH via glucocorticoid and mineralocorticoid receptors. These receptors are particularly abundant in the hippocampus. The excitatory output from hippocampal neurons along with the activity of inhibitory GABAergic cells regulate CRF-releasing neurons in the hypothalamus. Under stressful conditions, the HPA axis escapes this regulatory mechanism and produces large amounts of glucocorticoids [60,62]. A long-term increase in glucocorticoid levels reduces dendritic branching and dendritic length and induces the death of mature neurons and progenitor cells, resulting in reduced hippocampal volume. Thus, the disruption of neuroendocrine regulation ultimately impairs the excitability, functions, and integrity of the hippocampus; reduces plasticity and neurogenesis; and promotes susceptibility to the development of depression [58,63–69].

## 2.4. Oxidative Stress and Depression

Depression is accompanied by increased oxidative stress and reduced concentrations of antioxidants in the plasma [70,71].

Briefly, oxidative stress refers to a condition in which there is an imbalance between the generation of reactive oxygen and nitrogen species (ROS/RNS) and the ability of various enzymatic and non-enzymatic mechanisms of endogenous defense to maintain their levels in a physiological range. In appropriate concentrations, ROS have important functions, acting as signaling molecules in various redox-sensitive signaling pathways. However, if present in excess, they disturb neuronal signaling and react with cellular lipids, proteins, and nucleic acids, thus threatening their structure and proper neuronal functioning [72–75]. The brain is particularly vulnerable to oxidative injury due to its high metabolic activity, high content of redox-active transition metals that act as initiators of ROS generation via Fenton chemistry, high content of poly-unsaturated fatty acids (PUFAs) that are highly prone to lipid peroxidation, and limited mechanisms of antioxidative defense, among other reasons [76].

More importantly, specific molecules released or exposed during oxidative injury may elicit an innate immune response in the brain (acting as danger-associated molecular patterns (DAMPS)) and trigger sterile inflammation [77,78]. Thus, oxidative stress contributes to the inflammatory response and the increased production of proinflammatory cytokines, which inevitably results in the appearance of depressive behavior (Figure 1). Hence, both oxidative stress and neuroinflammation have been recognized as underlying factors in the development of depression [77,79–81].

The presence of oxidative stress markers is a common finding in depressed patients and animal models of depression [82]. 8-hydroxy-2'-deoxyguanosine (8-OHdG) is a typical indicator of oxidative DNA damage and it levels are significantly increased in blood samples from depressed patients [83,84]. The extent of lipid peroxidation in the peripheral blood is also increased and correlates with the severity of depressive symptoms [85–88]. Malondialdehyde (MDA), an end marker of lipid peroxidation, is usually found in increased concentrations in depressed patients [89]. Moreover, it has been suggested that certain oxidative stress indicators may be used as prognostic markers of disease severity and for the evaluation of the efficacy of administered antidepressants [82].

An increase in oxidative stress parameters reduces the content of intracellular mechanisms of antioxidative defense and jeopardizes neuronal protection. Thus, changes in the expression and activity of antioxidative and prooxidative enzymes have been observed; however, the results are not straightforward. Levels of the prooxidative enzyme xanthine oxidase, which generates superoxide anions and hydrogen peroxide, are usually increased in both depressed patients and animal models of depression [90,91]. On the other hand, levels of superoxide dismutase (SOD), an enzyme that decomposes superoxide to oxygen and hydrogen peroxide, were found to be reduced, unchanged, or enhanced in depressed patients [92–95]. Hydrogen peroxide is removed by catalase. Similarly, increased, unchanged, and decreased catalase activity has been reported [89,93,95]. Glutathione peroxidase also reduces hydrogen peroxide and regenerates reduced glutathione (GSH) from its oxidized pool (GSSG), thereby increasing the overall scavenging ability of GSH. The activity of glutathione peroxidase is predominantly found to be reduced in depressed patients, which is negatively correlated with the severity of the symptoms [93,96]. However, some studies did not observe changes in the glutathione peroxidase activity of depressed patients [84,92]. In yet another study, increases in the levels of glutathione peroxidase and SOD were dependent on the sample analyzed (plasma or erythrocytes) and clinical manifestations (depression with or without melancholia), suggesting that various factors affect the extent of changes in the activity of antioxidant enzymes in depressed patients [97]. Of note, the polymorphism of several genes participating in ROS metabolism may represent a risk for disease development and progression. It has been shown that single-nucleotide polymorphisms (SNPs) in genes encoding mitochondrial SOD (SOD2), catalase, and glutathione peroxidase 4 may modulate the risk of the onset of disease [98,99].



Figure 1. The main neuropathological mechanisms involved in the development of depression.

Quantities of non-enzymatic antioxidants are also disturbed. Besides developing oxidative stress that decreases the levels of these antioxidant molecules, depressed people often change their food preferences, which may deplete concentrations of specific dietary products, including vitamins [100]. Levels of vitamins A, C, and E as well as other small antioxidants (such as uric acid, albumin, and coenzyme Q10) are lower in depressed patients and negatively correlate with the disease's severity [86,101–103]. Hence, various dietary approaches, such as supplementation with the bioactive antioxidative ingredients from certain types of food, have been considered as adjuvant strategies for mitigating depressive symptoms [104–106]. Several studies have shown that supplementation with vitamin C may induce antidepressant effects and improve moods [107,108]. However, in combination with citalopram therapy, vitamin C was as effective as a placebo [109], whereas in combination with fluoxetine, it significantly mitigated depressive symptoms in a pediatric population [110]. Similarly, results on supplementation with vitamin E are inconclusive and further studies are needed to clarify the possible benefits of vitamin E with respect to the management of depression [102,111,112]. Administration of N-acetyl cysteine, a GSH precursor, also provided promising evidence regarding its use as an adjuvant therapy for depression. Several studies have indicated its ability to reduce the

severity of depression, improve moods, and increase the efficacy of standard antidepressant therapy, but these findings need further confirmation [113–115].

Metabolic reactions in the mitochondria are the major sources of ROS, but mitochondria are also the major targets of ROS action. Considering the principal role of mitochondria in determining cell death and survival via ATP production and the initiation of apoptosis, the preservation of mitochondrial functions is an important prerequisite for neuronal health [116,117]. In depressed patients, the expression and activity of enzymes participating in the mitochondrial respiratory chain are disrupted, leading to a reduction in ATP production [118,119]. A growing body of evidence is indicating that depression is related to mitochondrial dysfunction, which, consequently, disturbs ROS balance and promotes oxidative stress [120,121]. Depressed patients also have increased levels of circulating cell-free mtDNA (ccf-mtDNA), which likely reflects a fraction of the mitochondrial DNA released under cellular stress conditions [122].

Monoamine oxidase MAO-A and MAO-B, isoenzymes located in the outer mitochondrial membrane, catalyze the oxidative deamination of monoamines and deplete their levels in the brain, while also contributing to ROS production and oxidative stress. The MAO-A isoform is mainly involved in the metabolism of serotonin, dopamine, and norepinephrine, whereas MAO-B predominantly metabolizes dopamine. Increased activity of MAO-A is a common finding in depressed patients. It has been suggested that this upregulation is the predominant mechanism underlying monoamine loss [123–125]. Although MAO inhibitors were the first class of antidepressants developed and were used therapeutically for decades, based on concerns related to safety, possible side effects related to potential drug interactions, and dietary restrictions, they have been replaced with safer and more tolerable options. However, inhibitors of MAO activity are still considered the most valuable pharmacological option, particularly with respect to patients who fail to respond to first-line therapy [126–128].

Oxidative stress is usually accompanied by excitotoxicity, which is induced by the hyperactivation of glutamatergic NMDA receptors. It has been proposed that the impaired clearance and increased release of glutamate by activated glial cells upregulates glutamate levels and disturbs signaling through ionotropic and metabotropic glutamate receptors, thus contributing to neuronal dysfunction and, ultimately, behavioral changes [129]. Excitotoxicity also increases the production of NO, which, together with the superoxide anion, generates extremely cytotoxic peroxynitrite [130,131]. In addition, it has been shown that peroxynitrite inactivates tryptophan hydroxylase, the rate-limiting enzyme in serotonin synthesis [132]. Moreover, NO reacts with amino acid residues in proteins (mainly cysteine, tyrosine, tryptophan, and arginine) and causes protein oxidation, nitration, and nitrosylation, further threatening neuronal functioning [78,103,133]. However, peripheral measurements of nitrosative stress markers have provided variable results. For example, reduced levels of nitrate, without changes in NO and nitrite levels, have been reported in depressed patients [134].

There is yet another connection between depression and the activation of NMDA receptors. In inflammatory conditions, the kynurenine pathway and the production of kynurenine from tryptophan are potentiated, which per se reduce serotonin concentrations. On the other hand, kynurenine is metabolized by microglial enzymes, resulting in the production of various metabolites, some of which are neurotoxic, such as quinolinic acid, an NMDA receptor agonist [135,136].

Finally, the reduced expression of BDNF precipitates higher susceptibility to stressinduced oxidative damage, indicating an interplay between these depression-related parameters [137]. In turn, increased oxidative stress affects BDNF production and contributes to reduced neurogenesis and the dysregulation of hippocampal functioning [82].

## 2.5. Neuroinflammation, Cytokines, and Depression

Animal studies have provided evidence for a neuroinflammatory basis of depression, which entails sterile inflammation, glial activation, and the release of pro-inflammatory

cytokines [138]. Depression is accompanied by a chronic, low-grade inflammatory state; the dysregulation of the innate and adaptive immune system; enhanced production of various cytokines, such as interleukin (IL)-1, IL-2, IL-6, IL-10, IL-12, IL-13, and IL-1 $\beta$  and tumor necrosis factor (TNF)- $\alpha$ ; and decreased concentrations of interferon (INF)- $\gamma$  [139–143]. Treatment with antidepressants reduces the severity of inflammation, while severe inflammation correlates with lower treatment efficacy [144,145].

The excessive production of cytokines, the signaling molecules of the immune system, affects neurocircuitry in the basal ganglia and anterior cingulate cortex, the functioning of the HPA axis, synaptic plasticity, and neurotransmission, leading to behavioral alterations, mood changes, and the impairment of cognitive functions characteristic of depression. The behavioral effects of cytokines are largely mediated through the activation of the inflammatory signaling pathways that regulate the production of monoamines, glutamate, neuropeptides, and BDNF [146–149].

An increase in glucocorticoid concentrations mediates the effects of cytokines on the activation of the HPA axis and the dysregulation of the inhibitory feedback mechanism. Thus, the inflammatory response stimulates the HPA axis directly (through cytokine action) and indirectly (through glucocorticoid resistance) [150]. In major depressive disorders with melancholic features, a positive correlation has been found between the severity of the disease and concentrations of cortisol and IL-6 [151]. On the other hand, there is evidence that antidepressants normalize levels of proinflammatory cytokines and restore the feedback inhibition of the HPA axis [145,152]. In addition, an increase in proinflammatory cytokine concentrations may induce the tryptophan–kynurenine pathway and the generation of neurotoxic end products, as previously explained [150].

Oxidative stress and uncontrolled production of ROS play important roles in the hyperactivation of the immune pathways and the upregulation of cytokine production [77,153]. Enhanced production of proinflammatory cytokines and oxidative stress are tightly intertwined processes. Excessive production of ROS results in the oxidative damage of various cellular structures and the production of danger signals (DAMPs) that initiate inflammatory responses and microglial activation. Following activation, microglia, the resident cells of the innate immune system in the brain, produce proinflammatory cytokines and ROS. In a vicious loop, the ROS produced induce oxidative stress, supporting the ongoing microglial activation and further increasing ROS concentrations [133,154,155].

DAMPs-initiated signaling cascades transduce the signal to the nucleus and activate transcription factor nuclear factor  $\kappa$ B (NF- $\kappa$ B). The activation of the NF- $\kappa$ B cascade triggers the production of proinflammatory cytokines, reactive oxygen and nitrogen species, and other potentially neurotoxic molecules such as inducible nitric oxide synthase (iNOS). Increased activation of iNOS results in increased production of NO, which, in excess, acts as a powerful oxidant. As mentioned previously, the superoxide anion along with the NO produced by iNOS form extremely dangerous peroxynitrite radicals. Thus, sustained oxidative injury, the neuroinflammatory response, and microglial activation are largely driven by the NF- $\kappa$ B pathway, whereas this pathway's inhibition prevents the induction of the inhibitory effects of chronic stress on neurogenesis and the appearance of depressive behavior [156,157]. In animal models of neuroinflammation and depression, the upregulation of the NF- $\kappa$ B pathway has been demonstrated in the prefrontal cortex and the hippocampus [158,159]. However, in the human population, the results are not so consistent. For example, the activation of the NF- $\kappa$ B pathway was not observed in the peripheral blood mononuclear cells of adolescents and depressed medical students [160,161].

Several enzymes that are downstream targets of NF- $\kappa$ B may affect the course of depression. Such enzymes include cyclooxygenase 2 (COX-2) and NADPH oxidase. COX-2 inhibitors have been observed to have a positive effect on neuronal inflammation and the severity of the depression [162–164], whereas a single-nucleotide polymorphism of the *COX-2* gene may represent a risk for recurrent depressive disorder [165]. NADPH oxidase contributes to oxidative stress and neuroinflammation by overproducing the superoxide

anion. As with COX-2, the pharmacological inhibition of NADPH oxidase presented beneficial antidepressant effects [166].

## 3. Experimental Approaches and Behavioral Tests for Studying Depressive-like Behavior in Animals

Although several animal models of depression are available, none of them address all the aspects of human depression. Chronic-unpredictable-mild-stress (CUMS)-induced depression is considered to be the most reliable model. The corresponding protocol implies the application of various stressors varying in duration and severity according to an unpredictable schedule for 2–3 weeks (typically). Such stressors may include forced swimming in cold or warm water, wet or soiled bedding, food and/or water deprivation, the inversion of the light/dark cycle, overnight illumination, cage tilting, cage shaking, the generation of noise, electric shocks applied to the feet, tail pinching, tail squeezing, 24 h social isolation, etc. CUMS results in the reorganization of cortical and limbic areas, which probably underlies depressive-like behavioral abnormalities [167–169]. There are also some other approaches that induce similar neurochemical, endocrine-related, and behavioral characteristics of human depression, such as olfactory bulbectomy, the social defeat model, the chronic restraint stress model, and the prolonged administration of glucocorticoids (reviewed in [170]).

The antidepressant effects of the pharmacological agents of interest can be further monitored using several behavioral tests (described in detail in [171–173]). The forced swimming test (FST) is one of the most applied tests for estimating the efficacy of a particular drug as an antidepressant agent. Animals are exposed to unescapable stress and forced to swim in a container filled with water. The total immobility period (wherein animals float motionless, keeping their heads above water) indicates the depressive state of the animal. Struggling (active movements to escape) and swimming can also be monitored. Modified versions of the test can also be used.

As false positive results may be obtained with drugs that increase spontaneous locomotor activity, the open field test (OFT) is often used to exclude the possibility of the stimulatory effect of a drug. In this case, animals are placed at the center of an arena whose floor surface is divided into squares. In the OFT, various parameters can be monitored to estimate the effect of a drug on locomotor behavior, including the number of line crossings, the time spent in the center of the field, the number of entries in the peripheral zone, and the time spent in the peripheral zone, together with the number of rearing activities, which is assessed to monitor anxiety [167,174,175].

The tail suspension test (TST) is used to estimate the antidepressant effect of a particular compound. In the TST, animals are suspended above the floor by taping their tails. The parameter that is measured is immobility time as an indicator of behavioral despair [169].

The sucrose preference test (SPT) is used as an indicator of anhedonia. Usually, two bottles of water are placed in different areas of the cage. After adaptation, water in one bottle is replaced with 10% sucrose so that the animal can choose between two drinks. The positions of the bottles are switched on a daily basis to reduce side bias [176].

Additional tests are very frequently used to estimate the cognitive status of the animals. The passive avoidance task evaluates memory acquisition. The apparatus used has light and dark chambers, wherein a wire grid is placed on the floor of the dark compartment that delivers a shock to the feet of the animal that enters. Short-term and long-term memory retention can be estimated using this task. The novel object recognition test is usually performed in a rectangular arena. Animals are placed in the middle of an open field containing two identical objects and allowed to familiarize themselves with the object and arena for several minutes. In the testing phase, the animals are returned to the arena, wherein one object has been replaced with a new one, which should cause the animals to demonstrate greater exploratory activity around the novel object. The parameter that is usually calculated is a discrimination ratio (the time spent exploring the novel object divided by the total exploring time) that estimates recognition memory [177].

As anxiety is often diagnosed alongside depression, in addition to the OFT, the elevated plus maze test is usually employed to assess the antianxiety potential of pharmacological agents. It uses a device—made of two open and two closed arms—that is elevated above the floor. The animals are placed in the central area with their heads directed toward a closed arm. The animals are allowed to explore the open and closed arms, usually for three minutes, and the overall result is represented as the total time spent exploring all arms [178].

## 4. Antidepressant Effects of Flavonols

Current therapeutic strategies for the treatment of depression mainly focus on balancing neurotransmitter levels, often with limited efficacy and many side effects. Hence, searching for novel, multitarget alternatives with more rapidly acting effects and better efficacy and tolerability is of the highest priority in the medical and scientific community. Considering the importance of oxidative and inflammatory mechanisms in the pathophysiology of depression, it has been suggested that compounds able to re-establish redox homeostasis and suppress inflammation might be useful for relieving depressive symptoms. Nowadays, there is a great deal of interest in the use of natural polyphenolic compounds as antidepressant agents, mainly due to their proven powerful antioxidative, anti-inflammatory, and neuroprotective effects [179–182]. These bioactive molecules, of which flavonoids are of particular interest, are abundantly present in fruits and vegetables and also available in the form of commercial dietary supplements.

Based on epidemiological data, diets rich in polyphenols have many health-beneficial effects. From the perspective of depression, besides demonstrating a plethora of antioxidative and anti-inflammatory effects, polyphenols from the diet may regulate the activity of the HPA axis and normalize levels of glucocorticoids, stimulate BDNF production and neurogenesis, inhibit the activity of MAO isoforms, restore neurotransmitter balance, mitigate clinical symptoms, and improve cognitive deficits [175,183,184].

As flavonoids represent a large and diverse family of natural polyphenolic compounds, we focused on flavonols due to their dietary abundance and promising antidepressant activity [185]. Hence, the second part of this review summarizes the mechanisms of action of various flavonols and evaluates their potential for use in adjuvant therapeutic approaches to attenuating symptoms of depression.

## 4.1. Antidepressant Effects of Quercetin

Quercetin (3,3',4',5,7-pentahydroxyflavone), the best-studied compound from this class, is one of the most ubiquitous dietary flavonoids. It can be found in many fruits, vegetables, and beverages, particularly in onions, apples, green tea, and various berries [186,187]. In diverse experimental settings, quercetin displayed anti-oxidative, anti-inflammatory, and neuroprotective effects [188,189]. In general, quercetin has demonstrated powerful antioxidative effects. It acts as a direct ROS scavenger and can increase the levels and activity of the endogenous mechanisms of antioxidative defense (both enzymatic and non-enzymatic) [190,191]. It also provided protection against H<sub>2</sub>O<sub>2</sub>-induced neuronal injury [192–194]. H<sub>2</sub>O<sub>2</sub> is the most abundant endogenous ROS and is generated during oxidative deamination by MAO enzymes. Besides neutralizing the toxic effects of H<sub>2</sub>O<sub>2</sub>, quercetin also directly inhibits MAO-A [184,195]. The inhibitory effect of quercetin on MAO-B activity has also been shown in vitro [196].

Besides acting as an antioxidative agent, quercetin also displays prominent antiinflammatory properties. In cultured microglial cells, it prevented microglial proliferation and its phagocytic activity, ROS production, and the activation of the NF- $\kappa$ B pathway; suppressed the activation of the NOD-, LRR-, and pyrin-domain-containing protein 3 (NLRP3) inflammasomes; promoted mitophagy (to alleviate mitochondrial oxidative stress) and mitochondrial function; and the expression of IL-1 $\beta$  [197]. The same study demonstrated the ability of quercetin to protect neuronal cells against lipopolysaccharide (LPS)-induced microglial activation and neurotoxicity in a mouse model of depression.

Many behavioral tests performed using animal models have demonstrated beneficial effects of quercetin with respect to alleviating depressive-like behaviors (Figure 2, [198,199]). Several studies have shown anti-depressive effects of quercetin against CUMS-induced depression. Quercetin reversed CUMS-induced behavioral changes observed in the modified FST (wherein it increased swimming and escape attempts by climbing and decreased immobility time), TST (wherein it decreased immobility time), and OFT (wherein it increased number of field crossings and rearing behaviors). In the research in question, treatment with orally administered quercetin (25 mg/kg) started 2 weeks from the beginning of the CUMS protocol and lasted for 4 weeks, while the CUMS protocol was applied for 6 weeks. Behavioral improvements were accompanied by increased serotonin levels and improved oxidative stress parameters. Quercetin increased the activity of SOD and catalase, increased GSH content, and reduced glutamate levels, thus indicating the important role of the attenuation of oxidative stress and excitotoxicity in terms of its antidepressant effects. The CUMS-induced enhancement of TNF- $\alpha$  and IL-6 was also reversed by quercetin, further suggesting the contribution of anti-inflammatory mechanisms in the observed antidepressant effects of quercetin [169]. In a similar study (CUMS for 21 days and quercetin applied at a dose of 40 mg/kg, the antidepressant effect of quercetin was confirmed through improved results on the SPT and FST. At least partially, the stimulation of the hippocampal nuclear-factor-E2-related-factor-2 (Nrf2)-related signaling and inhibition of iNOS activity constituted the underlying mechanisms of this action [200]. Yet another study has shown that quercetin prevents behavioral abnormalities in animals exposed to chronic, unpredictable stress for 21 days. Quercetin, administered orally at a dose of 30 mg/kg during these 21 days, reduced anxiety (estimated using an elevated plus maze) and depressive-like behavior (SPT), improved memory retention (evaluated via a passive avoidance step-through task), and normalized locomotor activity (OFT) in stressed animals. Quercetin also attenuated oxidative stress in the hippocampus. It preserved total thiol content; reduced levels of thiobarbituric-acid-reactive substances (TBARS), which constitute a common measure of the severity of lipid peroxidation; reduced NO production; and improved catalase activity. Furthermore, at the gene level, it attenuated the expression of the proinflammatory markers IL-6, TNF- $\alpha$ , IL-1 $\beta$ , and COX-2 in the hippocampus. These antioxidative and anti-inflammatory effects had neuroprotective effects on hippocampal neurons, largely preventing their morphological changes and chronic stress-induced damage [167]. Besides regulating levels of serotonin and glutamate, it has been suggested that the inhibitory effects of quercetin on cholinergic nerves may contribute to this flavonol's antidepressant activity [199].

Quercetin also suppressed an acute stress response induced by water immersion and restraint (WIR) stress in Wistar rats. Quercetin administration (50 mg/kg) reduced plasma levels of WIR-induced stress hormones ACTH and corticosterone and attenuated the expression of CRH mRNA in the hypothalamus. In addition, quercetin regulated the DNAbinding abilities of glucocorticoid receptors and cyclic adenosine 3',5'-monophosphate (cAMP) response-element-binding protein (CREB). These transcription factors act as critical negative and positive regulators of transcriptional CRH expression, respectively. Kinases ERK1/2 are upstream of CREB, and quercetin suppressed the WIR-stress-induced phosphorylation of ERK1/2 as well. Altogether, these results imply that the role of quercetin in stress response control is mediated through the regulation of glucocorticoid receptors, CREB, and ERK1/2, resulting in the transcriptional suppression of CRH mRNA, which is the primary step in the activation of the HPA hormonal cascade [201]. Other studies also suggested that the antidepressant effects of quercetin are mediated through the suppression of CRF mRNA expression [202] and the ability to regulate the HPA axis [203]. However, in diabetic mice, the antidepressant effect of quercetin was determined to be independent of the HPA axis, as quercetin failed to modify levels of ACTH and corticosterone [204]. In another study performed using LPS-challenged rats, quercetin (40 mg/kg) increased the saccharin preference index and the immobility time in the FST, together with the normalization of synapsin-1, Copine 6, and BDNF levels, which might indicate that the antidepressant



effect was mediated through the restoration of the animals' BDNF levels and expression of synaptic plasticity-related proteins [205].

Figure 2. The main mechanisms contributing to antidepressant effects of quercetin and other flavonols.

Mild depression-like behavior can also be induced by chemotherapeutic agents such as adriamycin. In animals exposed to adriamycin, quercetin (60 mg/kg), which was applied 72 h later, reversed adriamycin-induced behavioral changes in the FST (it reduced immobility, increased swimming, and prolonged struggling behavior in comparison with the adriamycin group), restored corticosterone levels, normalized the number of immune cells and improved brain oxidative status as evidenced by the increased GSH content, decreased the activity of glutathione-S-transferase, and reduced MDA levels [175]. The antidepressant effect of quercetin was further demonstrated in an olfactory bulbectomy model. The ablation of olfactory bulbs in rodents induces hyperactivity in the OFT and increases immobility time in the FST. In addition, it results in the hyperactivation of the HPA axis, an increase in corticosterone levels, the impairment of hippocampal neurogenesis, the promotion of oxidative and nitrosative stress, the upregulation of markers of neuroinflammation, and the stimulation of neuronal apoptosis in cerebral cortex and hippocampus [206]. Quercetin (40 and 80 mg/kg) applied together with the microglial inhibitor minocycline demonstrated protective effects and attenuated behavioral, molecular, and histopathological abnormalities induced by olfactory bulbectomy. It reduced MDA levels and nitrite accumulation (as an indicator of NO production), improved GSH content and SOD and catalase activity, prevented caspase-3 activation, restored serum corticosterone levels, and reduced the secretion of IL-6 and TNF- $\alpha$ . These antioxidative and anti-inflammatory effects of quercetin preserved the normal histology of the cerebral cortex and hippocampus, attenuated the proliferation of microglial cells, and shortened the immobility time of the bulbectomized animals in the FST, thereby demonstrating its antidepressant effects [206].

The effects of quercetin isolated from plants were also studied. Accordingly, it was observed that quercetin and quercetin 4'-O-glucoside isolated from the outer scale of an onion (*Allium cepa*) demonstrated antidepressant-like effects in mice when applied at doses of 10 and 20 mg/kg. The effects of quercetin and quercetin 4'-O-glucoside were studied using the FST and OFT. Both quercetin and quercetin 4'-O-glucoside significantly

decreased the immobility time without displaying changes in the OFT, indicating their anti-depressive effects. However, as only the effects of quercetin 4'-O-glucoside were similar to those of fluoxetine, its effects were investigated further against CUMS-induced depressive-like behavior. Quercetin 4'-O-glucoside reversed CUMS-induced behavioral changes in locomotor activity and prevented a decrease in sucrose preference (a measure of anhedonia), thus improving the animals' capacity to experience pleasure. Together with these behavioral changes, quercetin 4'-O-glucoside reduced MAO-A activity and increased serotonin levels. Furthermore, it restored GSH levels and the accumulation of TBARS, altogether suggesting that the antidepressant effects of quercetin 4'-O-glucoside were mediated by its antioxidative abilities and MAO-inhibitory property that restored serotonin levels [207].

Likewise, the consumption of food rich in quercetin also presented antidepressant-like effects. The antidepressant potential of onion powder was evaluated in a rat model of depression in the FST, but, in contrast to some previous findings, the effect was independent of the HPA axis's regulation [208]. Hence, the levels of quercetin metabolites and range of dietary intake amounts needed for the attenuation of the HPA cascade should be addressed in future studies. The extract of *Ginkgo biloba* leaves, which is rich in quercetin glycosides, also reduced the immobility time in the FST and TST [209]. In addition, food enriched with quercetin (2 g/kg) was efficient in alleviating depressive behaviors in mice exposed to chronic social defeat stress. This was at least partially due to the reduced reactivation of astrocytes in the prefrontal cortex and hippocampus [210].

Hence, based on all these findings, the administration of quercetin can be considered a rational therapeutic approach to combat depression. It targets all of the main mechanisms contributing to the development of pathological processes, namely, oxidative stress, neuroinflammation, the dysfunction of the HPA axis, neurotransmitter levels, neuroplasticity, and neurogenesis. However, despite the convincing in vitro and preclinical evidence that suggests its remarkable potential with respect to alleviating depressive symptoms, there is still no adequate evidence from clinical trials to support its efficacy toward depressed patients. These studies are urgently needed to elucidate the exact capacity of quercetin as an antidepressant agent, to determine the optimal dose and the exact time frame for therapy, and to recognize eventual undesirable side effects when applied alone or in combination with standard therapy.

## 4.2. Antidepressant Effects of Myricetin and Myricitrin

Myricetin (3,3',4'5,5',7-hexahydroxylflavone) is a flavonol with a highly hydroxylated polyphenolic backbone. It is present in various vegetables, fruits, nuts, berries, and beverages [211]. As with quercetin, several studies have confirmed the neuroprotective effects of myricetin, which have largely been assigned to its antioxidative and anti-inflammatory properties [212]. For example, myricetin ameliorated cerebral-ischemia-induced-brain damage. Applied at a dose of 20 mg/kg, it reduced ROS levels and lipid peroxidation and promoted the activity of antioxidative enzymes SOD and catalase. These affordances improved mitochondrial function and stimulated the Nrf2 signaling pathway, resulting in reduced oxidative stress and the prevention of apoptotic death [213]. It has also been shown that myricetin may protect hippocampal neurons from damage [214]. However, caution is required, as in certain environmental conditions, such as increased concentrations of transient metal ions, myricetin may potentiate the production of ROS and exacerbate neuronal damage [215]. In this regard, the results of the meta-analysis have shown that depressed patients have increased blood levels of copper, which may exacerbate oxidative stress and systemic inflammation [216,217].

In mice exposed to repeated restraint stress for 21 days, myricetin (50 mg/kg) reversed the increase in the immobility time in the FST and TST without affecting locomotor activity in the OFT. Myricetin also reduced plasma ACTH and corticosterone levels, increased BDNF production, and partially restored the activity of glutathione peroxidase in hippocampal neurons [218,219]. In male rats exposed to a single prolonged stress (a model for the

induction of post-traumatic stress disorder, which is also accompanied by depression), myricetin was applied for 14 days following stress induction. At a dose of 20 mg/kg, it reduced depression-like behavior in the FST and SPT, reduced the increase in plasma ACTH and corticosterone levels, normalized levels of serotonin and norepinephrine in the hippocampus and prefrontal cortex, and induced the hippocampal production of BDNF and its receptor, tropomyosin-related kinase B (TrkB), by activating the ERK signaling pathway [220]. In SAMP8 mice displaying a phenotype of accelerated aging, the intake of myricetin also induced the production of BDNF and nerve growth factor (NGF) by increasing CREB phosphorylation [221].

Similar effects, i.e., a reversal of depression-like behavior in the FST and TST, reduced corticosterone levels, and increased activity of hippocampal SOD and glutathione peroxidase in mice exposed to different forms of stress for seven consecutive days, have been observed following an administration of the crude ethanolic extract of *Saraca asoca* flowers, which contains high amounts of myricetin [222].

Myricitrin (myricetin-3-O- $\alpha$ -rhamnoside), the glycosylated form of myricetin, is a natural flavonol present in plants from the genera *Myrica, Eugenia,* and *Pouteria.* Its antioxidant and anti-inflammatory properties have been confirmed in several in vitro and in vivo studies [182,223]. Applied at a dose of 10 mg/kg for 21 days, myricitrin reduced the animals' immobility time in the TST without modifying their general locomotor activity. It also promoted cell proliferation in the subventricular zone and dentate gyrus [224]. In a chronic mild stress model, myricitrin (at a dose of 10 mg/kg for 14 days) reversed depressive-like behavior, which was confirmed by an increased immobility time in the FST and TST, reduced adrenal hypertrophy as an indirect measure of the HPA axis's activation, and attenuated IL-6 levels in the hippocampus [225].

## 4.3. Antidepressant Effects of Rutin

Rutin (quercetin-3-O-rutinoside) is a flavonol with a rutinose (a disaccharide consisting of rhamnose and glucose) attached to the quercetin aglycone. In animal studies, it demonstrated antioxidative, anti-inflammatory, neuroprotective, and antidepressant effects [226–229]. Thus, in a rat model of streptozotocin-induced neurotoxicity related to oxidative damage and inflammation, rutin, which was applied for 3 weeks at a dose of 25 mg/kg, attenuated lipid peroxidation, nitrite accumulation, and the reduction in the amount of GSH and stimulated the activity of glutathione peroxidase, glutathione reductase, and catalase in the hippocampus. Regarding its anti-inflammatory activity, rutin reduced microglial activation and expression of COX-2, iNOS, and NF- $\kappa$ B, altogether preserving hippocampal morphology and cognitive functions [230].

In Swiss albino mice, rutin administered for 16 days (30 mg/kg, 60 mg/kg, and 120 mg/kg) reduced the animals' immobility time in an open-spaced FST and TST, indicating its anti-depressant potential. As the line-crossing in the OFT was increased, the authors suggested that the observed increase in locomotor activity reflected a rutin-induced increase in brain monoamine levels due to the inhibitory effect on MAO-A activity [231]. Similarly, in rats exposed to excess cadmium, rutin (50 mg/kg) induced protective effects by reducing the increase in cadmium-induced MAO activity and by increasing the activities of the antioxidant enzymes SOD and catalase in the hippocampus and cortex [232]. In a study that investigated the neurotoxic effects of silver nanoparticles, rutin reduced oxidative stress severity in the brain (it reduced MDA content, upregulated GSH, and increased activities of SOD, catalase, and glutathione peroxidase), normalized neurotransmitter levels (including the levels of serotonin, dopamine, and norepinephrine), regulated the transcriptional levels of MAO-A and MAO-B, improved histopathological changes induced by silver nanoparticles, and prevented astrogliosis and axonal demyelination [233]. The inhibitory effect of rutin on the MAO-B isoform's activity has also been shown in in vitro studies [196,234]. Among several tested flavonoids, rutin displayed the most potent inhibitory activity. As all the other flavonoids had only one sugar molecule in their structure, it was suggested that sugar portions determine the strength of MAO-B inhibition. In another study, orally

administered rutin decreased the immobility time in the TST at doses of 0.3, 1, and 3 mg/kg, demonstrating a positive effect on behavioral despair. This effect in the TST was prevented by a pretreatment with the inhibitors of serotonin and norepinephrine synthesis, suggesting that rutin increases the synaptic levels of these monoamines. On the other hand, rutin applied at these doses did not affect the number of crossings in the OFT, implying that its antidepressant effects were specific and not attributable to psychostimulatory activity [174]. The effects of rutin were also evaluated in mice exposed to CUMS for 21 days. Rutin improved behavioral deficits and induced antidepressant effects (it increased the preference for sucrose and pleasure), reduced anxiety (it enhanced the number of entries and the time spent in the center), and rescued cognitive functioning and motor coordination by preventing neuronal loss and preserving morphology in the CA3 region of the hippocampus [176]. In rats administered reserpine to induce depression (wherein a prolonged administration of reserpine induces monoamine depletion), rutin at a dose 80 mg/kg increased the number of crossings and rearing in the OFT and increased swimming and climbing behaviors in the FST, thus indicating the increased ability of the animals to cope with unavoidable stress conditions. A lack of helplessness was also confirmed via the TST [235].

Rutin also improved symptoms of depression by regulating the HPA axis. The oral administration of rutin (50 mg/kg) to animals exposed to CUMS improved levels of ACTH and corticosterone and normalized behavioral deteriorations in the OFT and FST. Rutin also increased the animals' intake of sucrose solution in a fluid consumption test [168].

In addition to pure rutin, botanical extracts containing rutin as one of their major constituents also demonstrated antioxidant, anti-inflammatory, and anti-depressive effects; decreased serum corticosterone levels and attenuated the HPS axis; increased the expression of CREB and BDNF; and inhibited the activity of MAO-A [174,236,237].

## 4.4. Antidepressant Effects of Avicularin

Avicularin (quercetin-3- $\alpha$ -L-arabinofuranoside) is a flavonol isolated from several plants, including *Polygonum aviculare* [238]. The effects of fluoxetine and avicularin have been compared in a mouse model of depression induced by chronic exposure to unpredictable mild stressors. Avicularin (1.25, 2.5, or 5.0 mg/kg/d) demonstrated antiinflammatory effects through the inhibition of the ERK/NF- $\kappa$ B pathway and the production of the proinflammatory cytokines IL-1 $\beta$ , IL-6, and TNF- $\alpha$  in the hippocampus. In addition, it inhibited the expression of iNOS, COX-2, and caspase-3. As the upregulated production of proinflammatory mediators plays an important role in the development of depression, the anti-inflammatory activity of avicularin was reflected in the improved results on behavioral tests (SPT, FST, and TST). At the highest dose, avicularin was as effective as fluoxetine in alleviating depressive-like behavior [239]. A prominent anti-inflammatory activity of avicularin mediated through the activation of the ERK signaling pathway was also observed in LPS-induced RAW 264.7 macrophage cells [240].

## 4.5. Antidepressant Effects of Fisetin

Fisetin (3,3',4',7-tetrahydroxyflavone) is a flavonol abundantly present in strawberries. Like other flavonols, fisetin may attenuate oxidative injury and neuroinflammation and mitigate neuronal damage. Many studies have demonstrated the powerful antioxidant ability of fisetin. Fisetin can reduce the severity of oxidative stress and the production of ROS and RNS, stimulate the activity of antioxidant enzymes and increase GSH levels, maintain redox balance and mitochondrial function, and prevent accompanying neuroinflammation. At the molecular level, fisetin modifies the activity of several signaling pathways involved in the determination of cell death or survival, including the PI3K/Akt, Nrf2, NF- $\kappa$ B, protein kinase C (PKC), and MAPK signaling pathways [241–245]. In a murine model of vascular dementia, the administration of fisetin at a dose of 40 mg/kg reduced lipid peroxidation and the death of hippocampal neurons, attenuated the activation of microglia and astrocytes, and promoted BDNF expression. In the same study, it reduced the number of apoptotic cells; upregulated the expression of the antiapoptotic protein Bcl-2

and downregulated expression of proapoptotic Bax; inhibited the inflammasome pathway; and reduced nuclear levels of NF- $\kappa$ B and the release of the pro-inflammatory cytokines IL-1 $\beta$  and IL-18, probably by preventing the ROS-mediated activation of the NF- $\kappa$ B/NLRP3 inflammasome and stimulating the antioxidative Nrf2/HO-1 pathway [246]. Similarly, regarding a brain injury induced by cerebral ischemia-reperfusion, fisetin reduced cell damage and the production of proinflammatory cytokines, inhibited iNOS and COX-2, and improved antioxidant parameters. These effects were mediated through the inhibition of the NF- $\kappa$ B pathway [247]. Likewise, the neuroprotective potential of fisetin against a spinal cord injury in rats was mediated by similar mechanisms. Fisetin (20 and 40 mg/kg) prevented a spinal-cord-injury-induced increase in proinflammatory markers (cytokines TNF- $\alpha$ , IL-1 $\beta$  and IL-6, iNOS, and COX-2) and the activation of the NF- $\kappa$ B/I $\kappa$ B $\alpha$  pathway and demonstrated antiapoptotic effects. It also upregulated the transcription of BDNF mRNA [248]. Hence, although these effects were not shown in animal models of depression, they clearly indicate that fisetin may modulate molecular and cellular processes involved in depression.

In mice, it has been shown that fisetin (10 and 20 mg/kg) may inhibit the immobility time in the TST and FST without affecting locomotor activity. Pre-treatment with p-chlorophenylalanine (PCPA), a selective inhibitor of tryptophane hydroxylase, nullified these beneficial effects. Furthermore, fisetin increased levels of serotonin and norepinephrine in the prefrontal cortex and hippocampus and inhibited MAO-A activity, suggesting that its antidepressant ability relies on its ability to normalize brain monoamine levels [249]. An inhibitory effect on MAO activity has also been demonstrated in astrocytes; this was an important finding, as the attenuation of glial MAO activity may aid neuronal protection [250]. In another study, a depressive-like behavior in mice was induced by LPS exposure. Pretreatment with fisetin (20-80 mg/kg for 7 days) reduced the immobility time in the FSF and TST and reversed an LPS-induced increase in inflammatory mediators, including IL-1 $\beta$ , IL-6, and TNF- $\alpha$ , in the prefrontal cortex and hippocampus. Moreover, at higher doses, fisetin suppressed the expression of iNOS mRNA and nitrite generation by modulating NF-KB, implying that the antidepressant effects of fisetin are largely determined by its anti-inflammatory ability [251]. In mice exposed to spatial restraint stress for 2 weeks, simultaneous treatment with fisetin (applied at a dose of 5 mg/kg) prevented an increase in the immobility time in the TST and FST [252]. In the same study, an antidepressant effect of fisetin was demonstrated in Abelson helper integration site-1 (Ahi1) knockout mice with a depressive phenotype. The effects of fisetin were likely mediated by the activation of the TrkB signaling pathway in the hippocampus [252]. Similarly, in mice exposed to a restraint test for 28 days, fisetin (15 mg/kg) improved behavioral deficits and attenuated the activation of the NF- $\kappa$ B cascade [253].

Finally, in a model of reserpine-induced fibromyalgia, which is characterized by reduced monoamine levels and depressive-like behavior, fisetin (10 and 25 mg/kg) reduced the immobility time in the FST and TST, prevented a monoamine decrease in the prefrontal cortex, and reversed reserpine-induced changes in oxidative and nitrosative stress markers (specifically the levels of ROS, GSH, MDA, SOD, and NO) [254].

## 4.6. Antidepressant Effects of Kaempferol

Kaempferol is a flavonol present in various foods (fruits and vegetables) and beverages and is among the most abundant flavonoids in the human diet [187]. Its neuroprotective effects have been demonstrated in previous studies. These effects were largely determined by its prominent antioxidative and anti-inflammatory action and modulation of distinct signaling cascades, including the NF- $\kappa$ B, p38, and Akt pathways [255]. Kaempferol's action along these pathways also underlies its antidepressant effects. Thus, in mice exposed to chronic social defeat stress for 10 days, kaempferol (administered at doses of 10 and 20 mg/kg for 28 consecutive days after stress) displayed antidepressant effects, which were partly induced by the stimulation of the activation of Akt/ $\beta$ -catenin signaling in the prefrontal cortex [256]. The upregulation of Akt/ $\beta$ -catenin signaling was associated with antioxidant effects (as evidenced by the increased activity of the antioxidant enzymes SOD, catalase, glutathione peroxidase, and glutathione S-transferase; reduced MDA content; and attenuated protein carbonylation) and anti-inflammatory activity (which was demonstrated through the attenuated release of the proinflammatory mediators IL-1 $\beta$  and TNF- $\alpha$  and microglial activation). Ultimately, these kaempferol-induced molecular changes contributed to the improved performance observed in behavioral tests. In particular, kaempferol increased sucrose consumption, time spent engaging in social interactions, and mobility time in the TST [256]. The administration of LY294002, a PI3K inhibitor, prevented improvements in behavioral deficits and the normalization of oxidative-stress- and neuroinflammation-related parameters, suggesting the important contribution of the Akt/ $\beta$ -catenin pathway in the anti-depressive action of kaempferol. In another study, kaempferol was administered at doses of 10, 20, and 40 µg/rat for 4 weeks. Depending on the dose, it improved performance in the TST and FST and suppressed MAO-A activity [257].

Antidepressant effects of kaempferol have also been observed in mice chronically exposed to restraint stress (2 h/day for 14 days). After being restrained, the mice were fed with a diet supplemented with kaempferol (30 mg/kg/day) for 14 days. Kaempferol reduced the immobility time in the TST and FST to the control level. In a rotarod test, the time of performance was also comparable to the control group. Moreover, kaempferol increased the plasma level of  $\beta$  endorphins, which may be relevant since opioid system has been implicated in the effects of certain antidepressants [258].

Yet another interesting study demonstrated that the supplementation with kaempferol-3-*O*-glucoside in female rats on an obesogenic diet during pregnancy and lactation may revert depression-like behavior in the female offspring [259].

A recent study demonstrated that sirtuin 3, the main mitochondrial deacetylase, had a prominent role in mediating the antidepressant and anxiolytic effects of kaempferol in an animal model of menopausal depression. The authors used two different approaches: one in which ovarian failure was induced by the ovotoxin 4-vinylcyclohexene diepoxide, and the other in which aged mice were exposed to CUMS for 8 weeks. Kaempferol (10 mg/kg for 14 days) improved performance in the behavioral tests (FST, OFT, and elevated plus maze) and increased the expression of sirtuin3 and SOD2 deacetylation (note that only the deacetylated form of SOD2 acts as an ROS scavenger), possibly by promoting the nuclear translocation of estrogen receptor  $\alpha$ , which might increase the expression of sirtuin 3. Kaempferol also alleviated mitochondrial dysfunction, increased activity of mitochondrial SOD, and attenuated oxidative stress in the hippocampus. These findings confirmed the important contribution of oxidative stress in menopausal depression and the potential of kaempferol as an antidepressant agent [260].

Similar to quercetin and other flavonols, the lack of clinical data is the major obstacle for evaluating the potential of kaempferol as an antidepressant agent. It has been shown that the antioxidative and anti-inflammatory effects of kaempferol depend on the glycosylation pattern [255]. Hence, clinical studies with kaempferol and its glycosylated derivatives are urgently needed to address their potential as antidepressant agents among depressed patients.

## 4.7. Antidepressant Effects of Morin

Morin (2',3,4',5,7-pentahydroxyflavone) is a flavonol that was originally isolated from plants of the *Moraceae* family, although it is also abundantly present in many fruits, herbs, green tea, and red wine. It possesses prominent antioxidative and anti-inflammatory abilities [261,262], and may prevent acute stress-induced biochemical and behavioral changes [263]. Hence, as with other flavonols, its protective potential against depressive-like behavior has been recognized. In rats exposed to CUMS for 4 weeks, morin administered concomitantly with stressors (15 and 30 mg/kg) induced antidepressant effects. It improved results in the FST (increased swimming score and reduced immobility time), SPT (increased intake of sucrose), and OFT. Furthermore, in the cerebral cortex and hippocampus, it mitigated neurochemical and biochemical changes induced by unpredictable stressors. It increased monoamine levels (serotonin, epinephrine, and norepinephrine) and improved oxidative stress status (evidenced by decreased MDA levels and increased glutathione content). The results also indicated its possible role in inflammasome activation, as morin reduced levels of the inflammasome pathway markers TNF- $\alpha$ , IL-1 $\beta$ , toll-like receptor-4 (TLR-4), NLRP3, and caspase-1, indicating its promising potential in alleviating the inflammatory basis of the disease [264]. Another study performed on mice suggested that the antidepressant effects of morin could be mediated through the L-arginine-NO pathway, as beneficial effects of morin on despair-like behavior were reversed by the NO precursor L-arginine [265].

The neuroprotective effect of morin in a rat model of attention deficit/hyperactivity disorder (ADHD) has also been demonstrated. The study indicated a crucial role of antioxidative and anti-inflammatory mechanisms in attenuating the severity of the disease. Thus, morin improved oxidative stress parameters (by targeting Nrf2/HO-1 pathway), monoamine levels, and inflammatory status (by hindering the TLR4/NLRP3 pathway) in pups with ADHD [266].

## 4.8. Antidepressant Effects of Other Flavonols

Isorhamnetin, a 3'-O-methylated derivative of quercetin, is one of the major bioactive compounds that can be isolated from *Ginkgo biloba* L. leaves. Antidepressant effects of *Ginkgo biloba* L. leaves have been demonstrated in various behavioral tests in rodents [209,267]. As for other flavonols, the antioxidative and anti-inflammatory effects of isorhamnetin have been reported in different pathological conditions and model systems and were mediated through the PI3K/Akt, MAPK, Nrf2, and NF-κB pathways [268,269]. However, further studies are needed, as so far, no antidepressant effects of isorhamnetin have been shown.

Rhamnazin is a methylated derivate of quercetin with antioxidative and anti-inflammatory properties [270]. Chronic administration of rhamnazin (50 mg/kg) improved cognitive dysfunctions induced by chronic restraint in rats (4 h/day for 30 days, without access to food and water), decreased ACTH levels in the plasma, and restored hippocampal BDNF levels [271].

Gossypetin is a flavonol that is abundantly present in *Hibiscus* species. At a dose of 20 mg/kg, gossypetin demonstrated significant antidepressant activity in rats exposed to forced swimming [272].

Galangin is a flavonol whose antidepressant potential has scarcely been investigated. However, it enhanced the antidepressant effect of fluoxetine in a rat model of depression, indicating the possibility of using galangin in combination with SSRIs to improve clinical outcomes [273].

Isoquercetin (quercetin 3'-O-rhamnoside) is a quercetin glycoside with an attached glucose. Like quercetin, it is effective in reducing oxidative stress and levels of proinflammatory cytokines by inhibiting the Nrf2 and NF-κB pathways [274]. For isoquercitrin (quercetin 4'-O-rhamnoside), inhibitory MAO-B activity has been shown [196]. Its administration (at a dose of 0.6 mg/kg/day) may reduce levels of ACTH and corticosterone when given for 2 weeks [275].

Hyperoside (quercetin 3-galactoside; flavonol glycoside) is one of the major bioactive compounds from *Hypericum perforatum* that displays prominent antidepressant effects. Hyperoside demonstrated antidepressant activity in CUMS-induced mice through increased BDNF levels and reduced expression of the NLRP1 inflammasome. Behavioral effects were confirmed via the SPT, TST, and FST [276]. The anti-immobility effect of hyperoside in rats (10 and 20 mg/kg) was prevented by a D2 antagonist, suggesting that the dopaminergic system could be involved in the antidepressant effect of hyperoside [277]. Moreover, treatment with hyperoside (0.6 mg/kg/day) for 2 weeks reduced plasma levels of ACTH and corticosterone [275].

## 5. Flavonols and Gut-Brain Axis

Gut microbiota constitute a wide spectrum of commensal microorganisms (bacteria, fungi, and viruses) that regulate the metabolic, endocrine, and immune functions of a host [278]. A growing body of data suggests that the dysbiosis of gut flora plays a contributory role in the pathogenesis of various neuropsychiatric diseases, including major depressive disorder. Depression affects the composition of gut microbiota [279,280], and various depressants exhibit antimicrobial activity [281,282]. Moreover, rats that received fecal microbiota from depressed patients developed depression-like behavior, indicating the causative role of gut microbiota in neurobehavioral symptoms. Likewise, behavioral alterations were accompanied by reduced levels of hippocampal neurotransmitters and serum cytokines, which also supports the existence of a functional interplay between gut microorganisms and the pathogenesis of depression [283]. Communication between the gut microbiota and the brain, termed the gut-brain axis, is bidirectional. The brain may affect the composition and function of gut microbiota by producing cytokines and antimicrobial peptides, while, through the enteric nervous system and the vagus nerve, bacteria in the gut may transmit signal to the brain and regulate its functioning. Gut microbiota produce various neuroactive chemicals, including neurotransmitters (such as GABA, catecholamines, histamine, and acetylcholine), as well as their precursors which may reach the brain through blood circulation and modify the synthesis and concentrations of neurotransmitters in the brain [284].Besides, some bacterial metabolites may regulate the production of neurotransmitters by enteroendocrine cells [285]. Moreover, various inflammatory molecules originating from the gut microflora (lipopolysaccharides, endotoxins, etc.) may activate the peripheral immune system, reach the brain, activate microglia, and promote the development of depression through cytokine release [284,286,287]. The microbiota-inflammasome hypothesis of major depressive disorder states that the dysbiosis of the gut microbiota deteriorates the integrity of the blood-brain barrier through peripheral inflammation. This results in the inflammasome's activation, the development of depressive symptoms, and further modulation of the gut microbiota [288,289]. On the other hand, intake of specific probiotics could contribute to the restoration of the health-promoting composition of the gut microbiota, prevent infiltration of the peripheral immune cells and microglial activation in the brain, and ultimately exert antidepressant effects [289]. It has been shown that many fecal metabolites and microbial genes distributed differently in patients and healthy individuals are related to amino acid metabolism ( $\gamma$ -aminobutyrate, phenylalanine, and tryptophan) and inflammatory imbalance [290]. Analysis of fecal samples also revealed that depressed patients have increased levels of Bacteroidetes, Proteobacteria, and Actinobacteria but a reduced relative abundance of *Firmicutes* [279]. Another study demonstrated that an abundance of GABA-producing Bacteroides negatively correlates with the functional connectivity in brain areas associated with depression [291]. Reduced abundance of *Bifi*dobacterium and Lactobacillus is also more common in depressed patients [292]. Similarly, in a rat model of ACTH-induced depression, microbial profiling revealed a reduced number of Lactobacillus and Akkermansia together with an increase in Ruminococcus and Klebsiella [280].

Among other contributing factors, the composition of the gut microflora is influenced by diet. In this regard, dietary polyphenols can modulate the intestinal microenvironment and functions of gut microbiota, which may affect the outcome of a therapeutic treatment [286,293,294]. On the other hand, polyphenols, including flavonols, need to be metabolized and activated by microorganisms in the gut. Via microbial metabolism, dietary polyphenols are broken down into small aromatic metabolites, some of which may directly act as neurotransmitters [287,293]. In fact, interactions between the microbiota and polyphenols in the diet are important factors in determining the effects of polyphenols on brain functions and the biological hallmarks of depression. There is increasing evidence that gut microbiota and dietary polyphenols from the diet or herbal medicinal products modulate the composition of gut microbiota by directly promoting the growth of health-beneficial bacterial species, whereas gut microbiota produce numerous metabolites from dietary polyphenols, mainly in the form of bioactive phenolic acids, with antidepressant properties usually superior in comparison with standard therapies [295,296]. Thus, during biotransformation by gut microbiota, polyphenols and their intermediary metabolites are degraded into various phenolic acids and their derivatives (e.g., ferulic acid, caffeic acid, ellagic acid, chlorogenic acid, benzoic acid, and hippuric acid). These bioactive metabolites are produced through the specific enzymatic activity of bacterial species in the gut. As mentioned, these metabolites usually show better absorption and biological effects. Several studies have demonstrated that they may attenuate depressive-like behavior by regulating monoamine levels, the HPA axis, antioxidative defense, and inflammation [295,297].

In light of these findings, attempts have been made to evaluate the antidepressant potential of flavonols from the perspective of the regulation of gut microbiota [287]. Thus, in LPS-challenged laying hens and broiler chickens, quercetin ameliorated intestinal mucosal injury and inflammation, maintained intestinal functions, and modified microbial communities [298,299]. Some of these changes were related to the increased abundance of bacteria capable of producing short-chain fatty acids (SCFAs) (e.g., acetate, propionate, and butyrate) from dietary fibers. SCFAs have an important role in the regulation of neuro-immunoendocrine functions and the improvement of cognitive abilities [298,300]. Moreover, SCFAs may signal entero-chrommafin cells to increase the production of serotonin, which is the key signaling molecule of the gut–brain axis [297]. SCFAs may also increase the production of the hormone leptin, which was able to reverse CUMS-induced hedonic deficits in the SPT and behavioral despair in the FST, and increase neuronal activity in the limbic system, including the hippocampus, all of which may contribute to the antidepressant effects of polyphenolic compounds [297,301]. Treatment with quercetin also increased Lactobacillus counts [299,302]. Of note, intervention with Lactobaccilus casie modified gut microbiota and improved depression-like behavior in rats exposed to CUMS [303]. Quercetin and its derivatives are also highly metabolized by the gut microbiota; however, the biological effects of their major metabolites have yet to be investigated [304].

Thus, although the mentioned findings could be considered to be indicative and promising, further studies are needed to clarify if flavonols can exert antidepressant effects by improving intestinal homeostasis.

## 6. Conclusions

Following COVID-19, there has been a huge outbreak of major depressive disorder worldwide. Unfortunately, the treatment options for depression are still not satisfactory. Apart from side effects and withdrawal symptoms, many patients do not respond to therapy, and therapy yields disappointing improvements far too often. Pharmacological interventions are mainly focused on balancing solely monoaminergic neurotransmission. However, multitarget approaches offer many advantages compared to classical prescriptions. Bioactive compounds originating from plants constitute an option with fewer unwanted side effects and with remarkable multimodal activities. They show prominent antioxidative activity, anti-inflammatory and neuroprotective effects, and the ability to regulate the HPA axis, protect neurons in the hippocampus, and stimulate neurogenesis.

Results from recent studies indicate the targeting of oxidative stress and inflammation as a promising therapeutic option for alleviating symptoms of depression as both are critical events in the cellular and molecular pathogenesis of this disease. Polyphenols from the flavonol class of flavonoids exert prominent antioxidative and anti-inflammatory effects, and in various animal models of depression, they have demonstrated remarkable anti-depressant activity. Although many research efforts have been directed to the most abundant flavonols in the diet, such as quercetin and kaempferol, studies on other flavonols are accumulating, which generally support the notion of flavonols acting as potential antidepressant agents.

However, clinical studies have yet to investigate the full potential of flavonols in depressed patients. These studies should be correlated not only with monoamine levels but also with the markers of oxidative stress and inflammation and levels of the HPA axis hormones. Regarding serotonin rebalancing, it is also necessary to work intensively on better understanding the interplay between flavonols and specific types of serotonin receptors. Monitoring all these parameters will give better insights into flavonols' capacity to alleviate molecular and cellular events in the brain and immune and endocrine systems that underlie the clinical symptoms of depression. As flavonols are relatively abundant in various fruits and vegetables, their incorporation in a daily diet, such as the Mediterranean one, which, among other beneficial effects, may also have a positive effect on the composition of gut microbiota, could be an effective approach for the maintenance of mental health and the prevention of or therapy for major depressive disorder. In general, the use of symbiotics (a combination of probiotics and dietary polyphenols) has been identified as a promising therapeutic approach. Due to the high variability of the composition of gut microbiota, it is likely that the simultaneous delivery of both probiotics and flavonols would be the best strategy for maximizing and standardizing the health-beneficial outcomes of flavonol administration. Symbiotics show great potential by targeting several pathological mechanisms, mainly those related to the attenuation of oxidative stress and neuroinflammation. However, as inter-individual differences related to genetic background, epigenetic modifications, lifestyle, and diet shape the interactions between gut microbiota and flavonols as well as their biotransformation into various metabolites in the lower colon, further studies are needed to optimize symbiotic formulas and better reveal the potential of flavonols against molecular and behavioral signs of depression. Finally, it should be emphasized that polyphenols from various groups, each with distinct biological effects, are simultaneously present in various plants, and their synergistic action potentially could exert even more prominent antidepressant action in comparison to flavonols alone, which needs to be investigated in further studies.

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## References

- Otte, C.; Gold, S.M.; Penninx, B.W.; Pariante, C.M.; Etkin, A.; Fava, M.; Mohr, D.C.; Schatzberg, A.F. Major depressive disorder. Nat. Rev. Dis. Prim. 2016, 2, 16065. [CrossRef]
- Christensen, M.C.; Wong, C.M.J.; Baune, B.T. Symptoms of Major Depressive Disorder and Their Impact on Psychosocial Functioning in the Different Phases of the Disease: Do the Perspectives of Patients and Healthcare Providers Differ? *Front. Psychiatry* 2020, 11, 280. [CrossRef]
- 3. Lim, G.Y.; Tam, W.W.; Lu, Y.; Ho, C.S.; Zhang, M.W.; Ho, R.C. Prevalence of Depression in the Community from 30 Countries between 1994 and 2014. *Sci. Rep.* 2018, *8*, 2861. [CrossRef]
- 4. Campbell, S.; Macqueen, G. The role of the hippocampus in the pathophysiology of major depression. *J. Psychiatry Neurosci.* 2004, 29, 417–426. [PubMed]
- Palmer, S.M.; Crewther, S.G.; Carey, L.M. START Project Team A meta-analysis of changes in brain activity in clinical depression. *Front. Hum. Neurosci.* 2015, *8*, 1045. [CrossRef] [PubMed]
- Zhang, F.F.; Peng, W.; Sweeney, J.A.; Jia, Z.Y.; Gong, Q.Y. Brain structure alterations in depression: Psychoradiological evidence. CNS Neurosci. Ther. 2018, 24, 994–1003. [CrossRef] [PubMed]
- Helm, K.; Viol, K.; Weiger, T.M.; Tass, P.A.; Grefkes, C.; Del Monte, D.; Schiepek, G. Neuronal connectivity in major depressive disorder: A systematic review. *Neuropsychiatr. Dis. Treat.* 2018, 14, 2715–2737. [CrossRef]
- Sindermann, L.; Redlich, R.; Opel, N.; Böhnlein, J.; Dannlowski, U.; Leehr, E.J. Systematic transdiagnostic review of magneticresonance imaging results: Depression, anxiety disorders and their co-occurrence. J. Psychiatr. Res. 2021, 142, 226–239. [CrossRef]
- Baek, S.J.; Park, J.S.; Kim, J.; Yamamoto, Y.; Tanaka-Yamamoto, K. VTA-projecting cerebellar neurons mediate stress-dependent depression-like behaviors. *eLife* 2022, 11, e72981. [CrossRef]

- Vermeiren, Y.; Van Dam, D.; Aerts, T.; Engelborghs, S.; De Deyn, P.P. Monoaminergic neurotransmitter alterations in postmortem brain regions of depressed and aggressive patients with Alzheimer's disease. *Neurobiol. Aging* 2014, 35, 2691–2700. [CrossRef]
- 11. Kunugi, H.; Hori, H.; Ogawa, S. Biochemical markers subtyping major depressive disorder. *Psychiatry Clin. Neurosci.* 2015, 69, 597–608. [CrossRef]
- 12. Ogawa, S.; Tsuchimine, S.; Kunugi, H. Cerebrospinal fluid monoamine metabolite concentrations in depressive disorder: A meta-analysis of historic evidence. *J. Psychiatr. Res.* 2018, 105, 137–146. [CrossRef]
- Silić, A.; Vukojević, J.; Peitl, V.; De Hert, M.; Karlović, D. Major depressive disorder: A possible typisation according to serotonin, inflammation, and metabolic syndrome. *Acta Neuropsychiaty* 2022, 34, 15–23. [CrossRef]
- 14. Shao, X.; Zhu, G. Associations Among Monoamine Neurotransmitter Pathways, Personality Traits, and Major Depressive Disorder. *Front. Psychiatry* **2020**, *11*, 381. [CrossRef]
- 15. Jacobsen, J.P.; Medvedev, I.O.; Caron, M.G. The 5-HT deficiency theory of depression: Perspectives from a naturalistic 5-HT deficiency model, the tryptophan hydroxylase 2Arg439His knockin mouse. *Philos. Trans R. Soc. Lond. B Biol. Sci.* **2012**, 367, 2444–2459. [CrossRef]
- 16. Correia, A.S.; Vale, N. Tryptophan Metabolism in Depression: A Narrative Review with a Focus on Serotonin and Kynurenine Pathways. *Int. J. Mol. Sci.* 2022, 23, 8493. [CrossRef]
- 17. Jiang, Y.; Zou, D.; Li, Y.; Gu, S.; Dong, J.; Ma, X.; Xu, S.; Wang, F.; Huang, J.H. Monoamine Neurotransmitters Control Basic Emotions and Affect Major Depressive Disorders. *Pharmaceuticals* **2022**, *15*, 1203. [CrossRef]
- 18. Delgado, P.L. Depression: The case for a monoamine deficiency. J. Clin. Psychiatry 2000, 61 (Suppl. S6), 7–11.
- Gabriel, F.C.; de Melo, D.O.; Fráguas, R.; Leite-Santos, N.C.; Mantovani da Silva, R.A.; Ribeiro, E. Pharmacological treatment of depression: A systematic review comparing clinical practice guideline recommendations. *PLoS ONE* 2020, 15, e0231700. [CrossRef]
- Elias, E.; Zhang, A.Y.; Manners, M.T. Novel Pharmacological Approaches to the Treatment of Depression. *Life* 2022, 12, 196. [CrossRef]
- Kessler, R.C.; Berglund, P.; Demler, O.; Jin, R.; Koretz, D.; Merikangas, K.R.; Rush, A.J.; Walters, E.E.; Wang, P.S. National Comorbidity Survey Replication The epidemiology of major depressive disorder: Results from the National Comorbidity Survey Replication (NCS-R). *JAMA* 2003, 289, 3095–3105. [CrossRef] [PubMed]
- Braund, T.A.; Tillman, G.; Palmer, D.M.; Gordon, E.; Rush, A.J.; Harris, A.W.F. Antidepressant side effects and their impact on treatment outcome in people with major depressive disorder: An iSPOT-D report. *Transl. Psychiatry* 2021, 11, 417. [CrossRef] [PubMed]
- 23. Li, Y.F. A hypothesis of monoamine (5-HT)—Glutamate/GABA long neural circuit: Aiming for fast-onset antidepressant discovery. *Pharmacol. Ther.* **2020**, *208*, 107494. [CrossRef] [PubMed]
- Duman, R.S.; Sanacora, G.; Krystal, J.H. Altered Connectivity in Depression: GABA and Glutamate Neurotransmitter Deficits and Reversal by Novel Treatments. *Neuron* 2019, 102, 75–90. [CrossRef] [PubMed]
- Taillefer de Laportalière, T.; Yrondi, A.; Jullien, A.; Cestac, P.; Montastruc, F. How to deprescribe esketamine in resistant depression? A point of view after first clinical uses. *Epidemiol. Psychiatr. Sci.* 2022, 31, e4. [CrossRef] [PubMed]
- Kantrowitz, J.T.; Dong, Z.; Milak, M.S.; Rashid, R.; Kegeles, L.S.; Javitt, D.C.; Lieberman, J.A.; John Mann, J. Ventromedial prefrontal cortex/anterior cingulate cortex Glx, glutamate, and GABA levels in medication-free major depressive disorder. *Transl. Psychiatry* 2021, 11, 419. [CrossRef]
- 27. Pandya, M.; Altinay, M.; Malone, D.A., Jr.; Anand, A. Where in the brain is depression? *Curr. Psychiatry Rep.* **2012**, *14*, 634–642. [CrossRef]
- Malberg, J.E.; Schechter, L.E. Increasing hippocampal neurogenesis: A novel mechanism for antidepressant drugs. *Curr. Pharm. Des.* 2005, 11, 145–155. [CrossRef]
- 29. Malykhin, N.V.; Carter, R.; Seres, P.; Coupland, N.J. Structural changes in the hippocampus in major depressive disorder: Contributions of disease and treatment. *J. Psychiatry Neurosci.* **2010**, *35*, 337–343. [CrossRef]
- 30. MacQueen, G.; Frodl, T. The hippocampus in major depression: Evidence for the convergence of the bench and bedside in psychiatric research? *Mol. Psychiatry* 2011, *16*, 252–264. [CrossRef]
- Roddy, D.W.; Farrell, C.; Doolin, K.; Roman, E.; Tozzi, L.; Frodl, T.; O'Keane, V.; O'Hanlon, E. The Hippocampus in Depression: More Than the Sum of Its Parts? Advanced Hippocampal Substructure Segmentation in Depression. *Biol. Psychiatry* 2019, 85, 487–497. [CrossRef]
- Hsieh, M.H.; McQuoid, D.R.; Levy, R.M.; Payne, M.E.; MacFall, J.R.; Steffens, D.C. Hippocampal volume and antidepressant response in geriatric depression. *Int. J. Geriatr. Psychiatry* 2002, 17, 519–525. [CrossRef]
- Boku, S.; Nakagawa, S.; Toda, H.; Hishimoto, A. Neural basis of major depressive disorder: Beyond monoamine hypothesis. Psychiatry Clin. Neurosci. 2018, 72, 3–12. [CrossRef]
- Kobayashi, K.; Ikeda, Y.; Sakai, A.; Yamasaki, N.; Haneda, E.; Miyakawa, T.; Suzuki, H. Reversal of hippocampal neuronal maturation by serotonergic antidepressants. *Proc. Natl. Acad. Sci. USA* 2010, 107, 8434–8439. [CrossRef]
- Anacker, C.; Zunszain, P.A.; Cattaneo, A.; Carvalho, L.A.; Garabedian, M.J.; Thuret, S.; Price, J.; Pariante, C.M. Antidepressants increase human hippocampal neurogenesis by activating the glucocorticoid receptor. *Mol. Psychiatry* 2011, 16, 738–750. [CrossRef]
- 36. Foltran, R.B.; Diaz, S.L. BDNF isoforms: A round trip ticket between neurogenesis and serotonin? *J. Neurochem.* **2016**, *138*, 204–221. [CrossRef]

- 37. Colucci-D'Amato, L.; Speranza, L.; Volpicelli, F. Neurotrophic Factor BDNF, Physiological Functions and Therapeutic Potential in Depression, Neurodegeneration and Brain Cancer. *Int. J. Mol. Sci.* 2020, *21*, 7777. [CrossRef]
- 38. Baydyuk, M.; Xu, B. BDNF signaling and survival of striatal neurons. Front. Cell. Neurosci. 2014, 8, 254. [CrossRef]
- 39. Wang, C.S.; Kavalali, E.T.; Monteggia, L.M. BDNF signaling in context: From synaptic regulation to psychiatric disorders. *Cell* **2022**, *185*, 62–76. [CrossRef]
- Ying, S.W.; Futter, M.; Rosenblum, K.; Webber, M.J.; Hunt, S.P.; Bliss, T.V.; Bramham, C.R. Brain-derived neurotrophic factor induces long-term potentiation in intact adult hippocampus: Requirement for ERK activation coupled to CREB and upregulation of Arc synthesis. J. Neurosci. 2002, 22, 1532–1540. [CrossRef]
- 41. Rex, C.S.; Lin, C.Y.; Kramár, E.A.; Chen, L.Y.; Gall, C.M.; Lynch, G. Brain-derived neurotrophic factor promotes long-term potentiation-related cytoskeletal changes in adult hippocampus. *J. Neurosci.* **2007**, *27*, 3017–3029. [CrossRef] [PubMed]
- Lau, A.G.; Irier, H.A.; Gu, J.; Tian, D.; Ku, L.; Liu, G.; Xia, M.; Fritsch, B.; Zheng, J.Q.; Dingledine, R.; et al. Distinct 3'UTRs differentially regulate activity-dependent translation of brain-derived neurotrophic factor (BDNF). *Proc. Natl. Acad. Sci. USA* 2010, 107, 15945–15950. [CrossRef] [PubMed]
- Pruunsild, P.; Kazantseva, A.; Aid, T.; Palm, K.; Timmusk, T. Dissecting the human BDNF locus: Bidirectional transcription, complex splicing, and multiple promoters. *Genomics* 2007, 90, 397–406. [CrossRef]
- Kowiański, P.; Lietzau, G.; Czuba, E.; Waśkow, M.; Steliga, A.; Moryś, J. BDNF: A Key Factor with Multipotent Impact on Brain Signaling and Synaptic Plasticity. *Cell. Mol. Neurobiol.* 2018, *38*, 579–593. [CrossRef] [PubMed]
- Karlović, D.; Serretti, A.; Jevtović, S.; Vrkić, N.; Serić, V.; Peleš, A.M. Diagnostic accuracy of serum brain derived neurotrophic factor concentration in antidepressant naïve patients with first major depression episode. *J. Psychiatr. Res.* 2013, 47, 162–167. [CrossRef]
- Molendijk, M.L.; Bus, B.A.; Spinhoven, P.; Penninx, B.W.; Kenis, G.; Prickaerts, J.; Voshaar, R.C.; Elzinga, B.M. Serum levels of brain-derived neurotrophic factor in major depressive disorder: State-trait issues, clinical features and pharmacological treatment. *Mol. Psychiatry* 2011, 16, 1088–1095. [CrossRef]
- Miyanishi, H.; Nitta, A. A Role of BDNF in the Depression Pathogenesis and a Potential Target as Antidepressant: The Modulator of Stress Sensitivity "Shati/Nat8l-BDNF System" in the Dorsal Striatum. *Pharmaceuticals* 2021, 14, 889. [CrossRef]
- Sakata, K.; Woo, N.H.; Martinowich, K.; Greene, J.S.; Schloesser, R.J.; Shen, L.; Lu, B. Critical role of promoter IV-driven BDNF transcription in GABAergic transmission and synaptic plasticity in the prefrontal cortex. *Proc. Natl. Acad. Sci. USA* 2009, 106, 5942–5947. [CrossRef]
- 49. Sakata, K.; Duke, S.M. Lack of BDNF expression through promoter IV disturbs expression of monoamine genes in the frontal cortex and hippocampus. *Neuroscience* **2014**, *260*, 265–275. [CrossRef]
- Sakata, K.; Jin, L.; Jha, S. Lack of promoter IV-driven BDNF transcription results in depression-like behavior. *Genes Brain Behav.* 2010, 9, 712–721. [CrossRef]
- Lieb, K.; Dreimüller, N.; Wagner, S.; Schlicht, K.; Falter, T.; Neyazi, A.; Müller-Engling, L.; Bleich, S.; Tadić, A.; Frieling, H. BDNF Plasma Levels and BDNF Exon IV Promoter Methylation as Predictors for Antidepressant Treatment Response. *Front. Psychiatry* 2018, 9, 511. [CrossRef]
- Jevtović, S.; Karlović, D.; Mihaljević-Peleš, A.; Šerić, V.; Vrkić, N.; Jakšić, N. Serum Brain-derived neurotrophic factor (BDNF): The severity and symptomatic dimensions of depression. *Psychiatr. Danub.* 2011, 23, 363–369.
- 53. Kreinin, A.; Lisson, S.; Nesher, E.; Schneider, J.; Bergman, J.; Farhat, K.; Farah, J.; Lejbkowicz, F.; Yadid, G.; Raskin, L.; et al. Blood BDNF level is gender specific in severe depression. *PLoS ONE* **2015**, *10*, e0127643. [CrossRef]
- Caldieraro, M.A.; Vares, E.A.; Souza, L.H.; Spanemberg, L.; Guerra, T.A.; Wollenhaupt-Aguiar, B.; Ferrari, P.; Nierenberg, A.A.; Fleck, M.P. Illness severity and biomarkers in depression: Using a unidimensional rating scale to examine BDNF. *Compr. Psychiatry* 2017, 75, 46–52. [CrossRef]
- Katsuki, A.; Yoshimura, R.; Kishi, T.; Hori, H.; Umene-Nakano, W.; Ikenouchi-Sugita, A.; Hayashi, K.; Atake, K.; Iwata, N.; Nakamura, J. Serum levels of brain-derived neurotrophic factor (BDNF), BDNF gene Val66Met polymorphism, or plasma catecholamine metabolites, and response to mirtazapine in Japanese patients with major depressive disorder (MDD). *CNS Spectr.* 2012, 17, 155–163. [CrossRef]
- 56. Brunoni, A.R.; Lopes, M.; Fregni, F. A systematic review and meta-analysis of clinical studies on major depression and BDNF levels: Implications for the role of neuroplasticity in depression. *Int. J. Neuropsychopharmacol.* **2008**, *11*, 1169–1180. [CrossRef]
- 57. Martinac, M.; Pehar, D.; Karlović, D.; Babić, D.; Marcinko, D.; Jakovljević, M. Metabolic syndrome, activity of the hypothalamicpituitary-adrenal axis and inflammatory mediators in depressive disorder. *Acta Clin. Croat.* **2014**, *53*, 55–71. [CrossRef]
- 58. Varghese, F.P.; Brown, E.S. The Hypothalamic-Pituitary-Adrenal Axis in Major Depressive Disorder: A Brief Primer for Primary Care Physicians. *Prim. Care Companion J. Clin. Psychiatry* **2001**, *3*, 151–155. [CrossRef]
- Iob, E.; Kirschbaum, C.; Steptoe, A. Persistent depressive symptoms, HPA-axis hyperactivity, and inflammation: The role of cognitive-affective and somatic symptoms. *Mol. Psychiatry* 2020, 25, 1130–1140. [CrossRef]
- Zhu, L.J.; Liu, M.Y.; Li, H.; Liu, X.; Chen, C.; Han, Z.; Wu, H.Y.; Jing, X.; Zhou, H.H.; Suh, H.; et al. The different roles of glucocorticoids in the hippocampus and hypothalamus in chronic stress-induced HPA axis hyperactivity. *PLoS ONE* 2014, 9, e97689. [CrossRef]
- Kino, T. Stress, glucocorticoid hormones, and hippocampal neural progenitor cells: Implications to mood disorders. *Front. Physiol.* 2015, 6, 230. [CrossRef] [PubMed]

- Cole, A.B.; Montgomery, K.; Bale, T.L.; Thompson, S.M. What the hippocampus tells the HPA axis: Hippocampal output attenuates acute stress responses via disynaptic inhibition of CRF+ PVN neurons. *Neurobiol. Stress* 2022, 20, 100473. [CrossRef] [PubMed]
- 63. Keller, J.; Gomez, R.; Williams, G.; Lembke, A.; Lazzeroni, L.; Murphy, G.M., Jr.; Schatzberg, A.F. HPA axis in major depression: Cortisol, clinical symptomatology and genetic variation predict cognition. *Mol. Psychiatry* **2017**, *22*, 527–536. [CrossRef] [PubMed]
- 64. Gjerstad, J.K.; Lightman, S.L.; Spiga, F. Role of glucocorticoid negative feedback in the regulation of HPA axis pulsatility. *Stress* **2018**, *21*, 403–416. [CrossRef]
- 65. Menke, A. Is the HPA Axis as Target for Depression Outdated, or Is There a New Hope? Front. Psychiatry 2019, 10, 101. [CrossRef]
- 66. McEwen, B.S. Glucocorticoids, depression, and mood disorders: Structural remodeling in the brain. *Metabolism* **2005**, 54 (Suppl. S1), 20–23. [CrossRef]
- 67. Krugers, H.J.; Lucassen, P.J.; Karst, H.; Joëls, M. Chronic stress effects on hippocampal structure and synaptic function: Relevance for depression and normalization by anti-glucocorticoid treatment. *Front. Synaptic Neurosci.* **2010**, *2*, 24. [CrossRef]
- Liston, C.; Gan, W.B. Glucocorticoids are critical regulators of dendritic spine development and plasticity in vivo. *Proc. Natl. Acad. Sci. USA* 2011, 108, 16074–16079. [CrossRef]
- 69. Tata, D.A.; Anderson, B.J. The effects of chronic glucocorticoid exposure on dendritic length, synapse numbers and glial volume in animal models: Implications for hippocampal volume reductions in depression. *Physiol. Behav.* **2010**, *99*, 186–193. [CrossRef]
- 70. Bhatt, S.; Nagappa, A.N.; Patil, C.R. Role of oxidative stress in depression. Drug Discover. Today 2020, 25, 1270–1276. [CrossRef]
- Juszczyk, G.; Mikulska, J.; Kasperek, K.; Pietrzak, D.; Mrozek, W.; Herbet, M. Chronic Stress and Oxidative Stress as Common Factors of the Pathogenesis of Depression and Alzheimer's Disease: The Role of Antioxidants in Prevention and Treatment. *Antioxidants* 2021, 10, 1439. [CrossRef]
- 72. Salim, S. Oxidative Stress and the Central Nervous System. J. Pharmacol. Exp. Ther. 2017, 360, 201–205. [CrossRef]
- 73. Wang, X.; Michaelis, E.K. Selective neuronal vulnerability to oxidative stress in the brain. *Front. Aging Neurosci.* **2010**, *2*, 12. [CrossRef]
- Singh, A.; Kukreti, R.; Saso, L.; Kukreti, S. Oxidative Stress: A Key Modulator in Neurodegenerative Diseases. *Molecules* 2019, 24, 1583. [CrossRef]
- 75. Lee, K.H.; Cha, M.; Lee, B.H. Neuroprotective Effect of Antioxidants in the Brain. Int. J. Mol. Sci. 2020, 21, 7152. [CrossRef]
- Cobley, J.N.; Fiorello, M.L.; Bailey, D.M. 13 reasons why the brain is susceptible to oxidative stress. *Redox Biol.* 2018, 15, 490–503. [CrossRef]
- 77. Bakunina, N.; Pariante, C.M.; Zunszain, P.A. Immune mechanisms linked to depression via oxidative stress and neuroprogression. *Immunology* **2015**, 144, 365–373. [CrossRef]
- Morris, G.; Berk, M.; Klein, H.; Walder, K.; Galecki, P.; Maes, M. Nitrosative Stress, Hypernitrosylation, and Autoimmune Responses to Nitrosylated Proteins: New Pathways in Neuroprogressive Disorders Including Depression and Chronic Fatigue Syndrome. *Mol. Neurobiol.* 2017, 54, 4271–4291. [CrossRef]
- Lopresti, A.L.; Maker, G.L.; Hood, S.D.; Drummond, P.D. A review of peripheral biomarkers in major depression: The potential of inflammatory and oxidative stress biomarkers. *Prog. Neuropsychopharmacol. Biol. Psychiatry* 2014, 48, 102–111. [CrossRef]
- Correia, A.S.; Cardoso, A.; Vale, N. Oxidative Stress in Depression: The Link with the Stress Response, Neuroinflammation, Serotonin, Neurogenesis and Synaptic Plasticity. *Antioxidants* 2023, 12, 470. [CrossRef]
- Zhong, J.; Li, G.; Xu, H.; Wang, Y.; Shi, M. Baicalin ameliorates chronic mild stress-induced depression-like behaviors in mice and attenuates inflammatory cytokines and oxidative stress. *Braz. J. Med. Biol. Res.* 2019, 52, e8434. [CrossRef] [PubMed]
- Vaváková, M.; Ďuračková, Z.; Trebatická, J. Markers of Oxidative Stress and Neuroprogression in Depression Disorder. Oxid. Med. Cell. Longev. 2015, 2015, 898393. [CrossRef] [PubMed]
- Forlenza, M.J.; Miller, G.E. Increased serum levels of 8-hydroxy-2'-deoxyguanosine in clinical depression. *Psychosom. Med.* 2006, 68, 1–7. [CrossRef] [PubMed]
- Lindqvist, D.; Dhabhar, F.S.; James, S.J.; Hough, C.M.; Jain, F.A.; Bersani, F.S.; Reus, V.I.; Verhoeven, J.E.; Epel, E.S.; Mahan, L.; et al. Oxidative stress, inflammation and treatment response in major depression. *Psychoneuroendocrinology* 2017, 76, 197–205. [CrossRef] [PubMed]
- Mazereeuw, G.; Herrmann, N.; Andreazza, A.C.; Khan, M.M.; Lanctôt, K.L. A meta-analysis of lipid peroxidation markers in major depression. *Neuropsychiatr. Dis. Treat.* 2015, 11, 2479–2491. [CrossRef]
- Liu, T.; Zhong, S.; Liao, X.; Chen, J.; He, T.; Lai, S. A Meta-Analysis of Oxidative Stress Markers in Depression. *PLoS ONE* 2015, 10, e0138904. [CrossRef]
- Islam, M.R.; Islam, M.R.; Ahmed, I.; Moktadir, A.A.; Nahar, Z.; Islam, M.S.; Shahid, S.F.B.; Islam, S.N.; Islam, M.S.; Hasnat, A. Elevated serum levels of malondialdehyde and cortisol are associated with major depressive disorder: A case-control study. SAGE Open Med. 2018, 6, 2050312118773953. [CrossRef]
- 88. Nobis, A.; Zalewski, D.; Waszkiewicz, N. Peripheral Markers of Depression. J. Clin. Med. 2020, 9, 3793. [CrossRef]
- Camkurt, M.A.; Fındıklı, E.; İzci, F.; Kurutaş, E.B.; Tuman, T.C. Evaluation of malondialdehyde, superoxide dismutase and catalase activity and their diagnostic value in drug naïve, first episode, non-smoker major depression patients and healthy controls. *Psychiatry Res.* 2016, 238, 81–85. [CrossRef]
- 90. Michel, T.M.; Camara, S.; Tatschner, T.; Frangou, S.; Sheldrick, A.J.; Riederer, P.; Grünblatt, E. Increased xanthine oxidase in the thalamus and putamen in depression. *World J. Biol. Psychiatry* **2010**, *11 Pt 2*, 314–320. [CrossRef]

- Xu, M.; Tian, P.; Zhu, H.; Zou, R.; Zhao, J.; Zhang, H.; Wang, G.; Chen, W. Lactobacillus paracasei CCFM1229 and Lactobacillus rhamnosus CCFM1228 Alleviated Depression- and Anxiety-Related Symptoms of Chronic Stress-Induced Depression in Mice by Regulating Xanthine Oxidase Activity in the Brain. Nutrients 2022, 14, 1294. [CrossRef]
- Sarandol, A.; Sarandol, E.; Eker, S.S.; Erdinc, S.; Vatansever, E.; Kirli, S. Major depressive disorder is accompanied with oxidative stress: Short-term antidepressant treatment does not alter oxidative-antioxidative systems. *Hum. Psychopharmacol.* 2007, 22, 67–73. [CrossRef]
- 93. Katrenčíková, B.; Vaváková, M.; Paduchová, Z.; Nagyová, Z.; Garaiova, I.; Muchová, J.; Ďuračková, Z.; Trebatická, J. Oxidative Stress Markers and Antioxidant Enzymes in Children and Adolescents with Depressive Disorder and Impact of Omega-3 Fatty Acids in Randomised Clinical Trial. Antioxidants 2021, 10, 1256. [CrossRef]
- 94. Tsai, M.C.; Huang, T.L. Increased activities of both superoxide dismutase and catalase were indicators of acute depressive episodes in patients with major depressive disorder. *Psychiatry Res.* **2016**, *235*, 38–42. [CrossRef]
- 95. Sabade, S.B.; Gagare, D.B.; Vikhe, B.B.; Bhalerao, M.M. Study of serum catalase in depression at Pravara institute of medical sciences. *Indian J. Clin. Anat. Physiol.* **2022**, *9*, 279–282. [CrossRef]
- 96. Maes, M.; Mihaylova, I.; Kubera, M.; Uytterhoeven, M.; Vrydags, N.; Bosmans, E. Lower whole blood glutathione peroxidase (GPX) activity in depression, but not in myalgic encephalomyelitis/chronic fatigue syndrome: Another pathway that may be associated with coronary artery disease and neuroprogression in depression. *Neuro. Endocrinol. Lett.* **2011**, *32*, 133–140.
- Bilici, M.; Efe, H.; Köroğlu, M.A.; Uydu, H.A.; Bekaroğlu, M.; Değer, O. Antioxidative enzyme activities and lipid peroxidation in major depression: Alterations by antidepressant treatments. J. Affect. Disord. 2001, 64, 43–51. [CrossRef]
- 98. da Cruz Jung, I.E.; da Cruz, I.B.M.; Barbisan, F.; Trott, A.; Houenou, L.J.; Osmarin Turra, B.; Duarte, T.; de Souza Praia, R.; Maia-Ribeiro, E.A.; da Costa Escobar Piccoli, J.; et al. Superoxide imbalance triggered by Val16Ala-SOD2 polymorphism increases the risk of depression and self-reported psychological stress in free-living elderly people. *Mol. Gen. Genom. Med.* 2019, *8*, e1080. [CrossRef]
- 99. Wigner, P.; Czarny, P.; Synowiec, E.; Bijak, M.; Białek, K.; Talarowska, M.; Galecki, P.; Szemraj, J.; Sliwinski, T. Variation of genes involved in oxidative and nitrosative stresses in depression. *Eur. Psychiatry* **2018**, *48*, 38–48. [CrossRef]
- Huang, Q.; Liu, H.; Suzuki, K.; Ma, S.; Liu, C. Linking What We Eat to Our Mood: A Review of Diet, Dietary Antioxidants, and Depression. *Antioxidants* 2019, *8*, 376. [CrossRef]
- Maes, M.; De Vos, N.; Pioli, R.; Demedts, P.; Wauters, A.; Neels, H.; Christophe, A. Lower serum vitamin E concentrations in major depression. Another marker of lowered antioxidant defenses in that illness. J. Affect. Disord. 2000, 58, 241–246. [CrossRef] [PubMed]
- Gautam, M.; Agrawal, M.; Gautam, M.; Sharma, P.; Gautam, A.S.; Gautam, S. Role of antioxidants in generalised anxiety disorder and depression. *Indian J. Psychiatry* 2012, 54, 244–247. [CrossRef] [PubMed]
- 103. Moylan, S.; Berk, M.; Dean, O.M.; Samuni, Y.; Williams, L.J.; O'Neil, A.; Hayley, A.C.; Pasco, J.A.; Anderson, G.; Jacka, F.N.; et al. Oxidative & nitrosative stress in depression: Why so much stress? *Neurosci. Biobehav. Rev.* 2014, 45, 46–62. [CrossRef] [PubMed]
- 104. Abshirini, M.; Siassi, F.; Koohdani, F.; Qorbani, M.; Mozaffari, H.; Aslani, Z.; Soleymani, M.; Entezarian, M.; Sotoudeh, G. Dietary total antioxidant capacity is inversely associated with depression, anxiety and some oxidative stress biomarkers in postmenopausal women: A cross-sectional study. *Ann. Gen. Psychiatry* 2019, 18, 3. [CrossRef]
- 105. Wu, S.-X.; Li, J.; Zhou, D.-D.; Xiong, R.-G.; Huang, S.-Y.; Saimaiti, A.; Shang, A.; Li, H.-B. Possible Effects and Mechanisms of Dietary Natural Products and Nutrients on Depression and Anxiety: A Narrative Review. *Antioxidants* 2022, 11, 2132. [CrossRef]
- 106. Winiarska-Mieczan, A.; Kwiecień, M.; Jachimowicz-Rogowska, K.; Donaldson, J.; Tomaszewska, E.; Baranowska-Wójcik, E. Anti-Inflammatory, Antioxidant, and Neuroprotective Effects of Polyphenols—Polyphenols as an Element of Diet Therapy in Depressive Disorders. Int. J. Mol. Sci. 2023, 24, 2258. [CrossRef]
- 107. Moritz, B.; Schmitz, A.E.; Rodrigues, A.L.S.; Dafre, A.L.; Cunha, M.P. The role of vitamin C in stress-related disorders. *J. Nutr. Biochem.* **2020**, *85*, 108459. [CrossRef]
- 108. Yosaee, S.; Keshtkaran, Z.; Abdollahi, S.; Shidfar, F.; Sarris, J.; Soltani, S. The effect of vitamin C supplementation on mood status in adults: A systematic review and meta-analysis of randomized controlled clinical trials. *Gen. Hosp. Psychiatry* 2021, 71, 36–42. [CrossRef]
- 109. Sahraian, A.; Ghanizadeh, A.; Kazemeini, F. Vitamin C as an adjuvant for treating major depressive disorder and suicidal behavior, a randomized placebo-controlled clinical trial. *Trials* **2015**, *16*, 94. [CrossRef]
- Amr, M.; El-Mogy, A.; Shams, T.; Vieira, K.; Lakhan, S.E. Efficacy of vitamin C as an adjunct to fluoxetine therapy in pediatric major depressive disorder: A randomized, double-blind, placebo-controlled pilot study. *Nutr. J.* 2013, 12, 31. [CrossRef]
- 111. Manosso, L.M.; Camargo, A.; Dafre, A.L.; Rodrigues, A.L.S. Vitamin E for the management of major depressive disorder: Possible role of the anti-inflammatory and antioxidant systems. *Nutr. Neurosci.* 2020, 25, 1310–1324. [CrossRef]
- 112. Lee, A.R.Y.B.; Tariq, A.; Lau, G.; Tok, N.W.K.; Tam, W.W.S.; Ho, C.S.H. Vitamin E, Alpha-Tocopherol, and Its Effects on Depression and Anxiety: A Systematic Review and Meta-Analysis. *Nutrients* **2022**, *14*, 656. [CrossRef]
- 113. Berk, M.; Dean, O.M.; Cotton, S.M.; Jeavons, S.; Tanious, M.; Kohlmann, K.; Hewitt, K.; Moss, K.; Allwang, C.; Schapkaitz, I.; et al. The efficacy of adjunctive N-acetylcysteine in major depressive disorder: A double-blind, randomized, placebo-controlled trial. *J. Clin. Psychiatry* 2014, 75, 628–636. [CrossRef]
- 114. Fernandes, B.S.; Dean, O.M.; Dodd, S.; Malhi, G.S.; Berk, M. N-Acetylcysteine in depressive symptoms and functionality: A systematic review and meta-analysis. *J. Clin. Psychiatry* **2016**, 77, e457–e466. [CrossRef]

- 115. Ooi, S.L.; Green, R.; Pak, S.C. N-Acetylcysteine for the Treatment of Psychiatric Disorders: A Review of Current Evidence. *Biomed. Res. Int.* **2018**, 2469486. [CrossRef]
- 116. Woo, J.; Cho, H.; Seol, Y.; Kim, S.H.; Park, C.; Yousefian-Jazi, A.; Hyeon, S.J.; Lee, J.; Ryu, H. Power Failure of Mitochondria and Oxidative Stress in Neurodegeneration and Its Computational Models. *Antioxidants* 2021, 10, 229. [CrossRef]
- 117. Tsang, Y.-L.; Kao, C.-L.; Lin, S.-C.A.; Li, C.-J. Mitochondrial Dysfunction and Oxidative Stress in Aging and Disease. *Biomedicines* **2022**, *10*, 2872. [CrossRef]
- Wang, Q.; Dwivedi, Y. Transcriptional profiling of mitochondria associated genes in prefrontal cortex of subjects with major depressive disorder. World J. Biol. Psychiatry 2017, 18, 592–603. [CrossRef]
- 119. Allen, J.; Romay-Tallon, R.; Brymer, K.J.; Caruncho, H.J.; Kalynchuk, L.E. Mitochondria and Mood: Mitochondrial Dysfunction as a Key Player in the Manifestation of Depression. *Front. Neurosci.* **2018**, *12*, 386. [CrossRef]
- 120. Tobe, E.H. Mitochondrial dysfunction, oxidative stress, and major depressive disorder. *Neuropsychiatr. Dis. Treat.* **2013**, *9*, 567–573. [CrossRef]
- 121. Bansal, Y.; Kuhad, A. Mitochondrial Dysfunction in Depression. Curr. Neuropharmacol. 2016, 14, 610–618. [CrossRef]
- 122. Lindqvist, D.; Wolkowitz, O.M.; Picard, M.; Ohlsson, L.; Bersani, F.S.; Fernström, J.; Westrin, A.; Hough, C.M.; Lin, J.; Reus, V.I.; et al. Circulating cell-free mitochondrial DNA, but not leukocyte mitochondrial DNA copy number, is elevated in major depressive disorder. *Neuropsychopharmacology* 2018, 43, 1557–1564. [CrossRef] [PubMed]
- 123. Meyer, J.H.; Ginovart, N.; Boovariwala, A.; Sagrati, S.; Hussey, D.; Garcia, A.; Young, T.; Praschak-Rieder, N.; Wilson, A.A.; Houle, S. Elevated monoamine oxidase a levels in the brain: An explanation for the monoamine imbalance of major depression. *Arch. Gen. Psychiatry* 2006, *63*, 1209–1216. [CrossRef] [PubMed]
- 124. Chiuccariello, L.; Houle, S.; Miler, L.; Cooke, R.G.; Rusjan, P.M.; Rajkowska, G.; Levitan, R.D.; Kish, S.J.; Kolla, N.J.; Ou, X.; et al. Elevated monoamine oxidase a binding during major depressive episodes is associated with greater severity and reversed neurovegetative symptoms. *Neuropsychopharmacology* **2014**, *39*, 973–980. [CrossRef] [PubMed]
- 125. Naoi, M.; Maruyama, W.; Shamoto-Nagai, M. Type A monoamine oxidase and serotonin are coordinately involved in depressive disorders: From neurotransmitter imbalance to impaired neurogenesis. J. Neural. Transm. 2018, 125, 53–66. [CrossRef] [PubMed]
- Stahl, S.M.; Felker, A. Monoamine oxidase inhibitors: A modern guide to an unrequited class of antidepressants. CNS Spectr. 2008, 13, 855–870. [CrossRef]
- 127. Flockhart, D.A. Dietary restrictions and drug interactions with monoamine oxidase inhibitors: An update. *J. Clin. Psychiatry* **2012**, 73 (Suppl. S1), 17–24. [CrossRef]
- 128. Shulman, K.I.; Herrmann, N.; Walker, S.E. Current place of monoamine oxidase inhibitors in the treatment of depression. *CNS Drugs* **2013**, *27*, 789–797. [CrossRef]
- 129. Haroon, E.; Miller, A.H.; Sanacora, G. Inflammation, Glutamate, and Glia: A Trio of Trouble in Mood Disorders. *Neuropsychopharmacology* **2017**, *42*, 193–215. [CrossRef]
- Jazvinšćak Jembrek, M.; Radovanović, V.; Vlainić, J.; Vuković, L.; Hanžić, N. Neuroprotective effect of zolpidem against glutamate-induced toxicity is mediated via the PI3K/Akt pathway and inhibited by PK11195. *Toxicology* 2018, 406–407, 58–69. [CrossRef]
- 131. Olloquequi, J.; Cornejo-Córdova, E.; Verdaguer, E.; Soriano, F.X.; Binvignat, O.; Auladell, C.; Camins, A. Excitotoxicity in the pathogenesis of neurological and psychiatric disorders: Therapeutic implications. *J. Psychopharmacol.* **2018**, *32*, 265–275. [CrossRef]
- Kuhn, D.M.; Geddes, T.J. Peroxynitrite inactivates tryptophan hydroxylase via sulfhydryl oxidation. Coincident nitration of enzyme tyrosyl residues has minimal impact on catalytic activity. J. Biol. Chem. 1999, 274, 29726–29732. [CrossRef]
- 133. Wang, H.; He, Y.; Sun, Z.; Ren, S.; Liu, M.; Wang, G.; Yang, J. Microglia in depression: An overview of microglia in the pathogenesis and treatment of depression. *J. Neuroinflamm.* **2022**, *19*, 132. [CrossRef]
- 134. Atagun, M.İ.; Atay, O.C.; Balaban, O.D.; Ipekcioglu, D.; Alpugan, B.; Yalcin, S.; Senat, A.; Karamustafalioglu, N.; Ilnem, M.C.; Erel, O. Serum nitric oxide levels are depleted in depressed patients treated with electroconvulsive therapy. *Indian J. Psychiatry* 2021, 63, 456–461. [CrossRef]
- 135. Savitz, J. Role of Kynurenine Metabolism Pathway Activation in Major Depressive Disorders. *Curr. Top. Behav. Neurrosci.* 2017, 31, 249–267. [CrossRef]
- 136. Marx, W.; McGuinness, A.J.; Rocks, T.; Ruusunen, A.; Cleminson, J.; Walker, A.J.; Gomes-da-Costa, S.; Lane, M.; Sanches, M.; Diaz, A.P.; et al. The kynurenine pathway in major depressive disorder, bipolar disorder, and schizophrenia: A meta-analysis of 101 studies. *Mol. Psychiatry* 2021, 26, 4158–4178. [CrossRef]
- Hacioglu, G.; Senturk, A.; Ince, I.; Alver, A. Assessment of oxidative stress parameters of brain-derived neurotrophic factor heterozygous mice in acute stress model. *Iran. J. Basic Med. Sci.* 2016, 19, 388–393.
- 138. Afridi, R.; Suk, K. Neuroinflammatory Basis of Depression: Learning from Experimental Models. *Front. Cell. Neurosci.* 2021, 15, 691067. [CrossRef]
- Crnković, D.; Buljan, D.; Karlović, D.; Krmek, M. Connection between inflammatory markers, antidepressants and depression. *Acta Clin. Croat.* 2012, 51, 25–33.
- 140. Berk, M.; Williams, L.J.; Jacka, F.N.; O'Neil, A.; Pasco, J.A.; Moylan, S.; Allen, N.B.; Stuart, A.L.; Hayley, A.C.; Byrne, M.L.; et al. So depression is an inflammatory disease, but where does the inflammation come from? *BMC Med.* **2013**, *11*, 200. [CrossRef]
- 141. Himmerich, H.; Patsalos, O.; Lichtblau, N.; Ibrahim, M.A.A.; Dalton, B. Cytokine Research in Depression: Principles, Challenges, and Open Questions. *Front. Psychiatry* **2019**, *10*, 30. [CrossRef] [PubMed]

- 142. Beurel, E.; Toups, M.; Nemeroff, C.B. The Bidirectional Relationship of Depression and Inflammation: Double Trouble. *Neuron* **2020**, *107*, 234–256. [CrossRef] [PubMed]
- 143. Kim, I.-B.; Lee, J.-H.; Park, S.-C. The Relationship between Stress, Inflammation, and Depression. *Biomedicines* **2022**, *10*, 1929. [CrossRef] [PubMed]
- 144. Lee, C.H.; Giuliani, F. The Role of Inflammation in Depression and Fatigue. Front. Immunol. 2019, 10, 1696. [CrossRef]
- 145. Hannestad, J.; DellaGioia, N.; Bloch, M. The effect of antidepressant medication treatment on serum levels of inflammatory cytokines: A meta-analysis. *Neuropsychopharmacology* **2011**, *36*, 2452–2459. [CrossRef]
- 146. Felger, J.C.; Lotrich, F.E. Inflammatory cytokines in depression: Neurobiological mechanisms and therapeutic implications. *Neuroscience* **2013**, 246, 199–229. [CrossRef]
- 147. Capuron, L.; Miller, A.H. Immune system to brain signaling: Neuropsychopharmacological implications. *Pharmacol. Ther.* **2011**, 130, 226–238. [CrossRef]
- 148. Jeenger, J.; Singroha, V.; Sharma, M.; Mathur, D.; Mathur, D.M. C-reactive protein, brain-derived neurotrophic factor, interleukin-2, and stressful life events in drug-naive first-episode and recurrent depression: A cross-sectional study. *Indian J. Psychiatry* 2018, 60, 334–339. [CrossRef]
- 149. Rengasamy, M.; Marsland, A.; Spada, M.; Hsiung, K.; Kovats, T.; Price, R.B. A chicken and egg scenario in psychoneuroimmunology: Bidirectional linking cytokines and depression. *J. Affect. Disord. Rep.* **2021**, *6*, 100177. [CrossRef]
- 150. Zunszain, P.A.; Anacker, C.; Cattaneo, A.; Carvalho, L.A.; Pariante, C.M. Glucocorticoids, cytokines and brain abnormalities in depression. *Prog. Neuro-Psychopharmacol. Biol. Psychiatry* **2011**, *35*, 722–729. [CrossRef]
- 151. Karlović, D.; Serretti, A.; Vrkić, N.; Martinac, M.; Marčinko, D. Serum concentrations of CRP, IL-6, TNF-α and cortisol in major depressive disorder with melancholic or atypical features. *Psychiatry Res.* **2012**, *198*, 74–80. [CrossRef]
- 152. Leonard, B.E. The immune system, depression and the action of antidepressants. *Prog. Neuropsychopharmacol. Biol. Psychiatry* **2001**, 25, 767–780. [CrossRef]
- 153. Zhang, J.; Wang, X.; Vikash, V.; Ye, Q.; Wu, D.; Liu, Y.; Dong, W. ROS and ROS-Mediated Cellular Signaling. *Oxid. Med. Cell.* Longev. 2016, 2016, 4350965. [CrossRef]
- 154. Simpson, D.S.A.; Oliver, P.L. ROS Generation in Microglia: Understanding Oxidative Stress and Inflammation in Neurodegenerative Disease. *Antioxidants* 2020, *9*, 743. [CrossRef]
- 155. Lee, K.H.; Cha, M.; Lee, B.H. Crosstalk between Neuron and Glial Cells in Oxidative Injury and Neuroprotection. *Int. J. Mol. Sci.* **2021**, 22, 13315. [CrossRef]
- 156. Koo, J.W.; Russo, S.J.; Ferguson, D.; Nestler, E.J.; Duman, R.S. Nuclear factor-kappaB is a critical mediator of stress-impaired neurogenesis and depressive behavior. *Proc. Natl. Acad. Sci. USA* 2010, 107, 2669–2674. [CrossRef]
- 157. Li, Y.; Song, W.; Tong, Y.; Zhang, X.; Zhao, J.; Gao, X.; Yong, J.; Wang, H. Isoliquiritin ameliorates depression by suppressing NLRP3-mediated pyroptosis via miRNA-27a/SYK/NF-κB axis. *J. Neuroinflamm.* **2021**, *18*, 1. [CrossRef]
- 158. Wang, Y.; Xu, J.; Liu, Y.; Li, Z.; Li, X. TLR4-NF-κB Signal Involved in Depressive-like Behaviors and Cytokine Expression of Frontal Cortex and Hippocampus in Stressed C57BL/6 and ob/ob Mice. *Neural Plast.* **2018**, 2018, 7254016. [CrossRef]
- 159. Chen, H.; Ma, Y.; Chen, M.; Chen, J.; Chen, J. Safflower extract improves depression in mice by inhibiting the TLR4-NLRP3 inflammation signaling pathway. *Ann. Palliat. Med.* **2021**, *10*, 8015–8023. [CrossRef]
- 160. Nazari, M.; Khodadadi, H.; Fathalizadeh, J.; Hassanshahi, G.; Bidaki, R.; Ayoobi, F.; Hajebrahimi, B.; Bagheri, F.; Arababadi, M.K. Defective NF-kB transcription factor as the mediator of inflammatory responses: A study on depressed Iranian medical students. *Clin. Lab.* 2013, 59, 827–830. [CrossRef]
- Miklowitz, D.J.; Portnoff, L.C.; Armstrong, C.C.; Keenan-Miller, D.; Breen, E.C.; Muscatell, K.A.; Eisenberger, N.I.; Irwin, M.R. Inflammatory cytokines and nuclear factor-kappa B activation in adolescents with bipolar and major depressive disorders. *Psychiatry Res.* 2016, 241, 315–322. [CrossRef] [PubMed]
- Müller, N. COX-2 Inhibitors, Aspirin, and Other Potential Anti-Inflammatory Treatments for Psychiatric Disorders. *Front. Psychiatry* 2019, 10, 375. [CrossRef] [PubMed]
- 163. Song, Q.; Feng, Y.B.; Wang, L.; Shen, J.; Li, Y.; Fan, C.; Wang, P.; Yu, S.Y. COX-2 inhibition rescues depression-like behaviors via suppressing glial activation, oxidative stress and neuronal apoptosis in rats. *Neuropharmacology* 2019, 160, 107779. [CrossRef] [PubMed]
- He, Y.; Han, Y.; Liao, X.; Zou, M.; Wang, Y. Biology of cyclooxygenase-2: An application in depression therapeutics. *Front. Psychiatry* 2022, 13, 1037588. [CrossRef]
- Gałecki, P.; Florkowski, A.; Bieńkiewicz, M.; Szemraj, J. Functional polymorphism of cyclooxygenase-2 gene (G-765C) in depressive patients. *Neuropsychobiology* 2010, 62, 116–120. [CrossRef]
- Seo, J.S.; Park, J.Y.; Choi, J.; Kim, T.K.; Shin, J.H.; Lee, J.K.; Han, P.L. NADPH oxidase mediates depressive behavior induced by chronic stress in mice. J. Neurosci. 2012, 32, 9690–9699. [CrossRef]
- Mehta, V.; Parashar, A.; Udayabanu, M. Quercetin prevents chronic unpredictable stress induced behavioral dysfunction in mice by alleviating hippocampal oxidative and inflammatory stress. *Physiol. Behav.* 2017, 171, 69–78. [CrossRef]
- 168. Quraishi, M.; Mokale, S.N.; Sakle, N.S. Ameliorative effect of quercetin and rutin via modulation of hypothalamic–Pituitary– Adrenal axis and regulation of fasting glucose in chronic stress-induced prediabetes. *Phcog. Mag.* 2018, 14 (Suppl. S1), 65–71. Available online: http://www.phcog.com/text.asp?2018/14/55/65/235260 (accessed on 28 June 2018).

- Khan, K.; Najmi, A.; Akhtar, M. A Natural Phenolic Compound Quercetin Showed the Usefulness by Targeting Inflammatory, Oxidative Stress Markers and Augment 5-HT Levels in One of the Animal Models of Depression in Mice. *Drug Res.* 2019, 69, 392–400. [CrossRef]
- 170. Wang, Q.; Timberlake, M.A., 2nd; Prall, K.; Dwivedi, Y. The recent progress in animal models of depression. *Prog. Neuropsychopharmacol. Biol. Psychiatry* 2017, 77, 99–109. [CrossRef]
- 171. D'Aquila, P.S.; Brain, P.; Willner, P. Effects of chronic mild stress on performance in behavioural tests relevant to anxiety and depression. *Physiol. Behav.* **1994**, *56*, 861–867. [CrossRef] [PubMed]
- 172. Belovicova, K.; Bogi, E.; Csatlosova, K.; Dubovicky, M. Animal tests for anxiety-like and depression-like behavior in rats. *Interdiscip. Toxicol.* **2017**, *10*, 40–43. [CrossRef] [PubMed]
- 173. Powell, T.R.; Fernandes, C.; Schalkwyk, L.C. Depression-Related Behavioral Tests. *Curr. Protoc. Mouse Biol.* 2012, 2, 119–127. [CrossRef] [PubMed]
- 174. Machado, D.G.; Bettio, L.E.; Cunha, M.P.; Santos, A.R.; Pizzolatti, M.G.; Brighente, I.M.; Rodrigues, A.L. Antidepressant-like effect of rutin isolated from the ethanolic extract from *Schinus molle* L. in mice: Evidence for the involvement of the serotonergic and noradrenergic systems. *Eur. J. Pharmacol.* 2008, 587, 163–168. [CrossRef]
- 175. Merzoug, S.; Toumi, M.L.; Tahraoui, A. Quercetin mitigates Adriamycin-induced anxiety- and depression-like behaviors, immune dysfunction, and brain oxidative stress in rats. *Naunyn Schmiedebergs Arch. Pharmacol.* 2014, 387, 921–933. [CrossRef]
- 176. Parashar, A.; Mehta, V.; Udayabanu, M. Rutin alleviates chronic unpredictable stress-induced behavioral alterations and hippocampal damage in mice. *Neurosci. Lett.* **2017**, *656*, 65–71. [CrossRef]
- Antunes, M.; Biala, G. The novel object recognition memory: Neurobiology, test procedure, and its modifications. *Cogn. Process.* 2012, 13, 93–110. [CrossRef]
- 178. Pellow, S.; Chopin, P.; File, S.E.; Briley, M. Validation of open:closed arm entries in an elevated plus-maze as a measure of anxiety in the rat. *J. Neurosci. Methods* **1985**, *14*, 149–167. [CrossRef]
- 179. Moore, A.; Beidler, J.; Hong, M.Y. Resveratrol and Depression in Animal Models: A Systematic Review of the Biological Mechanisms. *Molecules* **2018**, *23*, 2197. [CrossRef]
- Singh, A.; Yau, Y.F.; Leung, K.S.; El-Nezami, H.; Lee, J.C.-Y. Interaction of Polyphenols as Antioxidant and Anti-Inflammatory Compounds in Brain–Liver–Gut Axis. *Antioxidants* 2020, 9, 669. [CrossRef]
- 181. Mucha, P.; Skoczyńska, A.; Małecka, M.; Hikisz, P.; Budzisz, E. Overview of the Antioxidant and Anti-Inflammatory Activities of Selected Plant Compounds and Their Metal Ions Complexes. *Molecules* **2021**, *26*, 4886. [CrossRef]
- 182. Jazvinšćak Jembrek, M.; Oršolić, N.; Mandić, L.; Sadžak, A.; Šegota, S. Anti-Oxidative, Anti-Inflammatory and Anti-Apoptotic Effects of Flavonols: Targeting Nrf2, NF-κB and p53 Pathways in Neurodegeneration. *Antioxidants* 2021, 10, 1628. [CrossRef]
- Herraiz, T.; Guillén, H. Monoamine Oxidase-A Inhibition and Associated Antioxidant Activity in Plant Extracts with Potential Antidepressant Actions. *Biomed. Res. Int.* 2018, 2018, 4810394. [CrossRef]
- Dhiman, P.; Malik, N.; Sobarzo-Sánchez, E.; Uriarte, E.; Khatkar, A. Quercetin and Related Chromenone Derivatives as Monoamine Oxidase Inhibitors: Targeting Neurological and Mental Disorders. *Molecules* 2019, 24, 418. [CrossRef]
- Kim, K.; Vance, T.M.; Chun, O.K. Estimated intake and major food sources of flavonoids among US adults: Changes between 1999–2002 and 2007–2010 in NHANES. *Eur. J. Nutr.* 2016, 55, 833–843. [CrossRef]
- Boots, A.W.; Haenen, G.R.; Bast, A. Health effects of quercetin: From antioxidant to nutraceutical. *Eur. J. Pharmacol.* 2008, 585, 325–337. [CrossRef]
- Dabeek, W.M.; Marra, M.V. Dietary Quercetin and Kaempferol: Bioavailability and Potential Cardiovascular-Related Bioactivity in Humans. *Nutrients* 2019, 11, 2288. [CrossRef]
- 188. Zubčić, K.; Radovanović, V.; Vlainić, J.; Hof, P.R.; Oršolić, N.; Šimić, G.; Jazvinšćak Jembrek, M. PI3K/Akt and ERK1/2 Signalling Are Involved in Quercetin-Mediated Neuroprotection against Copper-Induced Injury. Oxid. Med. Cell. Longev. 2020, 2020, 9834742. [CrossRef]
- Islam, M.S.; Quispe, C.; Hossain, R.; Islam, M.T.; Al-Harrasi, A.; Al-Rawahi, A.; Martorell, M.; Mamurova, A.; Seilkhan, A.; Altybaeva, N.; et al. Neuropharmacological Effects of Quercetin: A Literature-Based Review. *Front. Pharmacol.* 2021, 12, 665031. [CrossRef]
- Xu, D.; Hu, M.-J.; Wang, Y.-Q.; Cui, Y.-L. Antioxidant Activities of Quercetin and Its Complexes for Medicinal Application. *Molecules* 2019, 24, 1123. [CrossRef]
- Yang, D.; Wang, T.; Long, M.; Li, P. Quercetin: Its Main Pharmacological Activity and Potential Application in Clinical Medicine. Oxid. Med. Cell. Longev. 2020, 2020, 8825387. [CrossRef] [PubMed]
- 192. Jazvinšćak Jembrek, M.; Vuković, L.; Puhović, J.; Erhardt, J.; Oršolić, N. Neuroprotective effect of quercetin against hydrogen peroxide-induced oxidative injury in P19 neurons. *J. Mol. Neurosci.* **2012**, *47*, 286–299. [CrossRef] [PubMed]
- Suematsu, N.; Hosoda, M.; Fujimori, K. Protective effects of quercetin against hydrogen peroxide-induced apoptosis in human neuronal SH-SY5Y cells. *Neurosci. Lett.* 2011, 504, 223–227. [CrossRef] [PubMed]
- 194. Godoy, J.A.; Lindsay, C.B.; Quintanilla, R.A.; Carvajal, F.J.; Cerpa, W.; Inestrosa, N.C. Quercetin Exerts Differential Neuroprotective Effects Against H<sub>2</sub>O<sub>2</sub> and Aβ Aggregates in Hippocampal Neurons: The Role of Mitochondria. *Mol. Neurobiol.* 2017, 54, 7116–7128. [CrossRef]
- 195. Bandaruk, Y.; Mukai, R.; Terao, J. Cellular uptake of quercetin and luteolin and their effects on monoamine oxidase-A in human neuroblastoma SH-SY5Y cells. *Toxicol. Rep.* 2014, *1*, 639–649. [CrossRef]

- Lee, M.H.; Lin, R.D.; Shen, L.Y.; Yang, L.L.; Yen, K.Y.; Hou, W.C. Monoamine oxidase B and free radical scavenging activities of natural flavonoids in Melastoma candidum D. Don. J. Agric. Food Chem. 2001, 49, 5551–5555. [CrossRef]
- 197. Han, X.; Xu, T.; Fang, Q.; Zhang, H.; Yue, L.; Hu, G.; Sun, L. Quercetin hinders microglial activation to alleviate neurotoxicity via the interplay between NLRP3 inflammasome and mitophagy. *Redox Biol.* 2021, 44, 102010. [CrossRef]
- Silvestro, S.; Bramanti, P.; Mazzon, E. Role of Quercetin in Depressive-Like Behaviors: Findings from Animal Models. *Appl. Sci.* 2021, 11, 7116. [CrossRef]
- 199. Chen, S.; Tang, Y.; Gao, Y.; Nie, K.; Wang, H.; Su, H.; Wang, Z.; Lu, F.; Huang, W.; Dong, H. Antidepressant Potential of Quercetin and its *Glycoside* Derivatives: A Comprehensive Review and Update. *Front. Pharmacol.* **2022**, *13*, 865376. [CrossRef]
- Guan, Y.; Wang, J.; Wu, X.; Song, L.; Wang, Y.; Gong, M.; Li, B. Quercetin reverses chronic unpredictable mild stress-induced depression-like behavior in vivo by involving nuclear factor-E2-related factor 2. *Brain Res.* 2021, 1772, 147661. [CrossRef]
- Kawabata, K.; Kawai, Y.; Terao, J. Suppressive effect of quercetin on acute stress-induced hypothalamic-pituitary-adrenal axis response in Wistar rats. J. Nutr. Biochem. 2010, 21, 374–380. [CrossRef]
- 202. Bhutada, P.; Mundhada, Y.; Bansod, K.; Ubgade, A.; Quazi, M.; Umathe, S.; Mundhada, D. Reversal by quercetin of corticotrophin releasing factor induced anxiety- and depression-like effect in mice. *Prog. Neuropsychopharmacol. Biol. Psychiatry* 2010, 34, 955–960. [CrossRef]
- Kosari-Nasab, M.; Shokouhi, G.; Ghorbanihaghjo, A.; Mesgari-Abbasi, M.; Salari, A.A. Quercetin mitigates anxiety-like behavior and normalizes hypothalamus-pituitary-adrenal axis function in a mouse model of mild traumatic brain injury. *Behav. Pharmacology* 2019, 30, 282–289. [CrossRef]
- Demir, E.A.; Gergerlioglu, H.S.; Oz, M. Antidepressant-like effects of quercetin in diabetic rats are independent of hypothalamicpituitary-adrenal axis. *Acta Neuropsychiatry* 2016, 28, 23–30. [CrossRef]
- 205. Fang, K.; Li, H.R.; Chen, X.X.; Gao, X.R.; Huang, L.L.; Du, A.Q.; Jiang, C.; Li, H.; Ge, J.F. Quercetin Alleviates LPS-Induced Depression-like Behavior in Rats via Regulating BDNF-Related Imbalance of Copine 6 and TREM1/2 in the Hippocampus and PFC. Front. Pharmacol. 2019, 10, 1544. [CrossRef]
- Rinwa, P.; Kumar, A. Quercetin suppress microglial neuroinflammatory response and induce antidepressent-like effect in olfactory bulbectomized rats. *Neuroscience* 2013, 255, 86–98. [CrossRef]
- 207. Singh, V.; Chauhan, G.; Shri, R. Anti-depressant like effects of quercetin 4'-O-glucoside from Allium cepa via regulation of brain oxidative stress and monoamine levels in mice subjected to unpredictable chronic mild stress. *Nutr. Neurosci.* 2021, 24, 35–44. [CrossRef]
- Sakakibara, H.; Yoshino, S.; Kawai, Y.; Terao, J. Antidepressant-like effect of onion (*Allium cepa* L.) powder in a rat behavioral model of depression. *Biosci. Biotechnol. Biochem.* 2008, 72, 94–100. [CrossRef]
- 209. Sakakibara, H.; Ishida, K.; Grundmann, O.; Nakajima, J.; Seo, S.; Butterweck, V.; Minami, Y.; Saito, S.; Kawai, Y.; Nakaya, Y.; et al. Antidepressant effect of extracts from Ginkgo biloba leaves in behavioral models. *Biol. Pharm. Bull.* 2006, 29, 1767–1770. [CrossRef]
- Zhang, J.; Ning, L.; Wang, J. Dietary quercetin attenuates depressive-like behaviors by inhibiting astrocyte reactivation in response to stress. *Biochem. Biophys. Res. Commun.* 2020, 533, 1338–1346. [CrossRef]
- 211. Taheri, Y.; Suleria, H.; Martins, N.; Sytar, O.; Beyatli, A.; Yeskaliyeva, B.; Seitimova, G.; Salehi, B.; Semwal, P.; Painuli, S.; et al. Myricetin bioactive effects: Moving from preclinical evidence to potential clinical applications. *BMC Complement. Med. Ther.* 2020, 20, 241. [CrossRef] [PubMed]
- Semwal, D.K.; Semwal, R.B.; Combrinck, S.; Viljoen, A. Myricetin: A Dietary Molecule with Diverse Biological Activities. *Nutrients* 2016, 8, 90. [CrossRef] [PubMed]
- Wu, S.; Yue, Y.; Peng, A.; Zhang, L.; Xiang, J.; Cao, X.; Ding, H.; Yin, S. Myricetin ameliorates brain injury and neurological deficits via Nrf2 activation after experimental stroke in middle-aged rats. *Food Func.* 2016, 7, 2624–2634. [CrossRef] [PubMed]
- Ramezani, M.; Darbandi, N.; Khodagholi, F.; Hashemi, A. Myricetin protects hippocampal CA3 pyramidal neurons and improves learning and memory impairments in rats with Alzheimer's disease. *Neural Regen Res.* 2016, 11, 1976–1980. [CrossRef] [PubMed]
- Sadžak, A.; Vlašić, I.; Kiralj, Z.; Batarelo, M.; Oršolić, N.; Jazvinšćak Jembrek, M.; Kušen, I.; Šegota, S. Neurotoxic effect of flavonol myricetin in the presence of excess copper. *Molecules* 2021, 26, 845. [CrossRef]
- Ni, M.; You, Y.; Chen, J.; Zhang, L. Copper in depressive disorder: A systematic review and meta-analysis of observational studies. *Psychiatry Res.* 2018, 267, 506–515. [CrossRef]
- Jazvinšćak Jembrek, M.; Vlainić, J.; Radovanović, V.; Erhardt, J.; Oršolić, N. Effects of copper overload in P19 neurons: Impairment of glutathione redox homeostasis and crosstalk between caspase and calpain protease systems in ROS-induced apoptosis. *Biometals* 2014, 27, 1303–1322. [CrossRef]
- Ma, Z.; Wang, G.; Cui, L.; Wang, Q. Myricetin Attenuates Depressant-like Behavior in Mice Subjected to Repeated Restraint Stress. Int. J. Mol. Sci. 2015, 16, 28377–28385. [CrossRef]
- Wang, Q.M.; Wang, G.L.; Ma, Z.G. Protective effects of myricetin on chronic stress-induced cognitive deficits. *Neuroreport* 2016, 27, 652–658. [CrossRef]
- 220. Sur, B.; Lee, B. Myricetin Inhibited Fear and Anxiety-Like Behaviors by HPA Axis Regulation and Activation of the BDNF-ERK Signaling Pathway in Posttraumatic Stress Disorder Rats. Evid. Based Complement. Alternat. Med. 2022, 2022, 8320256. [CrossRef]

- 221. Shimada, Y.; Sato, Y.; Kumazoe, M.; Kitamura, R.; Fujimura, Y.; Tachibana, H. Myricetin improves cognitive function in SAMP8 mice and upregulates brain-derived neurotrophic factor and nerve growth factor. *Biochem. Biophys. Res. Commun.* 2022, 616, 33–40. [CrossRef]
- Shashikumara, S.; Purushotham, K.; Darshan, C.L.; Kalal, B.S. Characterization of antidepressant activity of *Saraca asoca* flower (Roxb.) Wilde in mice subjected to acute restraint stress. *Am. J. Transl. Res.* 2022, *14*, 5014–5023.
- Kim, H.D.; Jeong, K.H.; Jung, U.J.; Kim, S.R. Myricitrin ameliorates 6-hydroxydopamine-induced dopaminergic neuronal loss in the substantia nigra of mouse brain. J. Med. Food 2016, 19, 374–382. [CrossRef]
- 224. Meyer, E.; Mori, M.A.; Campos, A.C.; Andreatini, R.; Guimarães, F.S.; Milani, H.; de Oliveira, R.M. Myricitrin induces antidepressant-like effects and facilitates adult neurogenesis in mice. *Behav. Brain Res.* **2017**, *316*, 59–65. [CrossRef]
- 225. Pereira, M.; Siba, I.P.; Acco, A.; Correia, D.; Lapa, F.R.; Santos, A.R.S.; Ruani, A.P.; Pizzolatti, M.G.; Andreatini, R. Myricitrin exhibits antidepressant-like effects and reduces IL-6 hippocampal levels in the chronic mild stress model. *Behav. Brain Res.* 2022, 429, 113905. [CrossRef]
- Dimpfel, W. Rat electropharmacograms of the flavonoids rutin and quercetin in comparison to those of moclobemide and clinically used reference drugs suggest antidepressive and/or neuroprotective action. *Phytomedicine* 2009, 16, 287–294. [CrossRef]
- 227. Khan, M.M.; Ahmad, A.; Ishrat, T.; Khuwaja, G.; Srivastawa, P.; Khan, M.B.; Raza, S.S.; Javed, H.; Vaibhav, K.; Khan, A.; et al. Rutin protects the neural damage induced by transient focal ischemia in rats. *Brain Res.* **2009**, *1292*, 123–135. [CrossRef]
- Koda, T.; Kuroda, Y.; Imai, H. Rutin supplementation in the diet has protective effects against toxicant-induced hippocampal injury by suppression of microglial activation and pro-inflammatory cytokines: Protective effect of rutin against toxicant-induced hippocampal injury. *Cell. Mol. Neurobiol.* 2009, 29, 523–531. [CrossRef]
- Çelik, H.; Kandemir, F.M.; Caglayan, C.; Özdemir, S.; Çomaklı, S.; Kucukler, S.; Yardım, A. Neuroprotective effect of rutin against colistin-induced oxidative stress, inflammation and apoptosis in rat brain associated with the CREB/BDNF expressions. *Mol. Biol. Rep.* 2020, 47, 2023–2034. [CrossRef]
- 230. Javed, H.; Khan, M.M.; Ahmad, A.; Vaibhav, K.; Ahmad, M.E.; Khan, A.; Ashafaq, M.; Islam, F.; Siddiqui, M.S.; Safhi, M.M.; et al. Rutin prevents cognitive impairments by ameliorating oxidative stress and neuroinflammation in rat model of sporadic dementia of Alzheimer type. *Neuroscience* 2012, 210, 340–352. [CrossRef] [PubMed]
- Yusha'u, Y.; Muhammad, U.A.; Nze, M.; Egwuma, J.M.; Igomu, O.J.; Abdulkadir, M. Modulatory Role of Rutin Supplement on Open Space Forced Swim Test Murine Model of Depression. *Niger. J. Physiol. Sci.* 2017, 32, 201–205. [PubMed]
- Oboh, G.; Adebayo, A.A.; Ademosun, A.O.; Olowokere, O.G. Rutin alleviates cadmium-induced neurotoxicity in Wistar rats: Involvement of modulation of nucleotide-degrading enzymes and monoamine oxidase. *Metab. Brain Dis.* 2019, 34, 1181–1190. [CrossRef] [PubMed]
- Ahmed, M.M.; Hussein, M.M.A. Neurotoxic effects of silver nanoparticles and the protective role of rutin. *Biomed. Pharmacother.* 2017, 90, 731–739. [CrossRef] [PubMed]
- 234. Azam, F.; Abodabos, H.S.; Taban, I.M.; Rfieda, A.R.; Mahmood, D.; Anwar, M.J.; Khan, S.; Sizochenko, N.; Poli, G.; Tuccinardi, T. Rutin as promising drug for the treatment of Parkinson's disease: An assessment of MAO-B inhibitory potential by docking, molecular dynamics and DFT studies. *Mol. Simul.* 2019, 45, 1563–1571. [CrossRef]
- 235. Foudah, A.I.; Alqarni, M.H.; Alam, A.; Devi, S.; Salkini, M.A.; Alam, P. Rutin Improves Anxiety and Reserpine-Induced Depression in Rats. *Molecules* 2022, 27, 7313. [CrossRef]
- 236. Barauna, S.C.; Delwing-Dal Magro, D.; Brueckheimer, M.B.; Maia, T.P.; Sala, G.A.B.N.; Döhler, A.W.; Harger, M.C.; de Melo, D.F.M.; de Gasper, A.L.; Alberton, M.D.; et al. Antioxidant and antidepressant-like effects of Eugenia catharinensis D. Legrand in an animal model of depression induced by corticosterone. *Metab. Brain Dis.* 2018, 33, 1985–1994. [CrossRef]
- 237. Daodee, S.; Monthakantirat, O.; Ruengwinitwong, K.; Gatenakorn, K.; Maneenet, J.; Khamphukdee, C.; Sekeroglu, N.; Chulikhit, Y.; Kijjoa, A. Effects of the Ethanol Extract of Dipterocarpus alatus Leaf on the Unpredictable Chronic Mild Stress-Induced Depression in ICR Mice and Its Possible Mechanism of Action. *Molecules* 2019, 24, 3396. [CrossRef]
- Zhang, X.Q.; Xu, L.S. Determination of avicularin in *Polygonum aviculare* L. by square wave polarography. *Proc. Chin. Acad. Med. Sci. Peking Union Med. Coll.* 1998, 4, 193–195.
- Shen, Z.; Xu, Y.; Jiang, X.; Wang, Z.; Guo, Y.; Pan, W.; Hou, J. Avicularin Relieves Depressive-like Behaviors Induced by Chronic Unpredictable Mild Stress in Mice. *Med. Sci. Monit.* 2019, 25, 2777–2784. [CrossRef]
- Vo, V.A.; Lee, J.W.; Chang, J.E.; Kim, J.Y.; Kim, N.H.; Lee, H.J.; Kim, S.S.; Chun, W.; Kwon, Y.S. Avicularin Inhibits Lipopolysaccharide-Induced Inflammatory Response by Suppressing ERK Phosphorylation in RAW 264.7 Macrophages. *Biomol. Ther.* 2012, 20, 532–537. [CrossRef]
- 241. Maher, P.; Dargusch, R.; Bodai, L.; Gerard, P.E.; Purcell, J.M.; Marsh, J.L. ERK activation by the polyphenols fisetin and resveratrol provides neuroprotection in multiple models of Huntington's disease. *Human Mol. Gen.* **2011**, *20*, 261–270. [CrossRef]
- 242. Chiruta, C.; Schubert, D.; Dargusch, R.; Maher, P. Chemical modification of the multitarget neuroprotective compound fisetin. *J. Med. Chem.* **2012**, *55*, 378–389. [CrossRef]
- Nabavi, S.F.; Braidy, N.; Habtemariam, S.; Sureda, A.; Manayi, A.; Nabavi, S.M. Neuroprotective Effects of Fisetin in Alzheimer's and Parkinson's Diseases: From Chemistry to Medicine. *Curr. Top. Med. Chem.* 2016, 16, 1910–1915. [CrossRef]
- 244. Wang, T.H.; Wang, S.Y.; Wang, X.D.; Jiang, H.Q.; Yang, Y.Q.; Wang, Y.; Cheng, J.L.; Zhang, C.T.; Liang, W.W.; Feng, H.L. Fisetin Exerts Antioxidant and Neuroprotective Effects in Multiple Mutant hSOD1 Models of Amyotrophic Lateral Sclerosis by Activating ERK. *Neuroscience* 2018, 379, 152–166. [CrossRef]

- 245. Hassan, S.S.U.; Samanta, S.; Dash, R.; Karpiński, T.M.; Habibi, E.; Sadiq, A.; Ahmadi, A.; Bunagu, S. The neuroprotective effects of fisetin, a natural flavonoid in neurodegenerative diseases: Focus on the role of oxidative stress. *Front. Pharmacol.* 2022, 13, 1015835. [CrossRef]
- 246. Cordaro, M.; D'Amico, R.; Fusco, R.; Peritore, A.F.; Genovese, T.; Interdonato, L.; Franco, G.; Arangia, A.; Gugliandolo, E.; Crupi, R.; et al. Discovering the Effects of Fisetin on NF-κB/NLRP-3/NRF-2 Molecular Pathways in a Mouse Model of Vascular Dementia Induced by Repeated Bilateral Carotid Occlusion. *Biomedicines* 2022, 10, 1448. [CrossRef] [PubMed]
- 247. Zhang, P.; Cui, J. Neuroprotective Effect of Fisetin against the Cerebral Ischemia-Reperfusion Damage via Suppression of Oxidative Stress and Inflammatory Parameters. *Inflammation* **2021**, *44*, 1490–1506. [CrossRef]
- 248. Cui, J.; Fan, J.; Li, H.; Zhang, J.; Tong, J. Neuroprotective potential of fisetin in an experimental model of spinal cord injury: Via modulation of NF-κB/IκBα pathway. *Neuroreport* **2021**, *32*, 296–305. [CrossRef]
- Zhen, L.; Zhu, J.; Zhao, X.; Huang, W.; An, Y.; Li, S.; Du, X.; Lin, M.; Wang, Q.; Xu, Y.; et al. The antidepressant-like effect of fisetin involves the serotonergic and noradrenergic system. *Behav. Brain Res.* 2012, 228, 359–366. [CrossRef]
- Mazzio, E.A.; Harris, N.; Soliman, K.F. Food constituents attenuate monoamine oxidase activity and peroxide levels in C6 astrocyte cells. *Planta Med.* 1998, 64, 603–606. [CrossRef]
- 251. Yu, X.; Jiang, X.; Zhang, X.; Chen, Z.; Xu, L.; Chen, L.; Wang, G.; Pan, J. The effects of fisetin on lipopolysaccharide-induced depressive-like behavior in mice. *Metab. Brain Dis.* 2016, *31*, 1011–1021. [CrossRef] [PubMed]
- 252. Wang, Y.; Wang, B.; Lu, J.; Shi, H.; Gong, S.; Wang, Y.; Hamdy, R.C.; Chua, B.H.L.; Yang, L.; Xu, X. Fisetin provides antidepressant effects by activating the tropomyosin receptor kinase B signal pathway in mice. *J. Neurochem.* **2018**, 143, 561–568. [CrossRef]
- 253. Choubey, P.; Kwatra, M.; Pandey, S.N.; Kumar, D.; Dwivedi, D.K.; Rajput, P.; Mishra, A.; Lahkar, M.; Jangra, A. Ameliorative effect of fisetin against lipopolysaccharide and restraint stress-induced behavioral deficits via modulation of NF-κB and IDO-1. *Psychopharmacology* **2018**, 236, 741–752. [CrossRef] [PubMed]
- 254. Yao, X.; Li, L.; Kandhare, A.D.; Mukherjee-Kandhare, A.A.; Bodhankar, S.L. Attenuation of reserpine-induced fibromyalgia via ROS and serotonergic pathway modulation by fisetin, a plant flavonoid polyphenol. *Exp. Ther. Med.* 2020, 19, 1343–1355. [CrossRef]
- 255. Silva Dos Santos, J.; Gonçalves Cirino, J.P.; de Oliveira Carvalho, P.; Ortega, M.M. The Pharmacological Action of Kaempferol in Central Nervous System Diseases: A Review. *Front. Pharmacol.* 2021, 11, 565700. [CrossRef]
- 256. Gao, W.; Wang, W.; Peng, Y.; Deng, Z. Antidepressive effects of kaempferol mediated by reduction of oxidative stress, proinflammatory cytokines and up-regulation of AKT/β-catenin cascade. *Metab. Brain Dis.* 2019, 34, 485–494. [CrossRef]
- Park, S.H.; Sim, Y.B.; Han, P.L.; Lee, J.K.; Suh, H.W. Antidepressant-like Effect of Kaempferol and Quercitirin, Isolated from Opuntia ficus-indica var. saboten. *Exp. Neurobiol.* 2010, 19, 30–38. [CrossRef]
- 258. de la Garza, A.L.; Garza-Cuellar, M.A.; Silva-Hernandez, I.A.; Cardenas-Perez, R.E.; Reyes-Castro, L.A.; Zambrano, E.; Gonzalez-Hernandez, B.; Garza-Ocañas, L.; Fuentes-Mera, L.; Camacho, A. Maternal Flavonoids Intake Reverts Depression-like Behaviour in Rat Female Offspring. *Nutrients* 2019, 11, 572. [CrossRef]
- 259. Li, H.-Y.; Wang, J.; Liang, L.-F.; Shen, S.-Y.; Li, W.; Chen, X.-R.; Li, B.; Zhang, Y.-Q.; Yu, J. Sirtuin 3 Plays a Critical Role in the Antidepressant- and Anxiolytic-like Effects of Kaempferol. *Antioxidants* **2022**, *11*, 1886. [CrossRef]
- Zarei, M.; Mohammadi, S.; Komaki, A.; Golipour Choshali, Z. Antidepressant-like Effects of Intra-cerebroventricular Microinjection of Kaempferol in Male Rats: Involvement of 5-HT2 Receptors. *Avicenna J. Neuro. Psycho. Physiol.* 2022, 9, 23–30. Available online: http://ajnpp.umsha.ac.ir/article-1-379-en.html (accessed on 1 February 2022).
- Zhang, Z.T.; Cao, X.B.; Xiong, N.; Wang, H.C.; Huang, J.S.; Sun, S.G.; Wang, T. Morin exerts neuroprotective actions in Parkinson disease models in vitro and in vivo. *Acta Pharmacol. Sin.* 2010, *31*, 900–906. [CrossRef] [PubMed]
- Alberdi, E.; Sánchez-Gómez, M.V.; Ruiz, A.; Cavaliere, F.; Ortiz-Sanz, C.; Quintela-López, T.; Capetillo-Zarate, E.; Solé-Domènech, S.; Matute, C. Mangiferin and morin attenuate oxidative stress, mitochondrial dysfunction, and neurocytotoxicity, induced by amyloid beta oligomers. Oxid. Med. Cell. Longev. 2018, 2018, 2856063. [CrossRef]
- Olonode, E.T.; Aderibigbe, A.O.; Adeoluwa, O.A.; Ajayi, A.M. Protective Effects of Morin Hydrate on Acute Stress-Induced Behavioral and Biochemical Alterations in Mice. *Basic Clin. Neurosci.* 2018, *9*, 195–208. [CrossRef] [PubMed]
- Hassan, M.M.; Gad, A.M.; Menze, E.T.; Badary, O.A.; El-Naga, R.N. Protective effects of morin against depressive-like behavior prompted by chronic unpredictable mild stress in rats: Possible role of inflammasome-related pathways. *Biochem. Pharmacol.* 2020, 180, 114140. [CrossRef]
- Ben-Azu, B.; Aderibigbe, A.O.; Ajayi, A.M.; Umukoro, S.; Iwalewa, E.O. Involvement of l-arginine-nitric oxide pathway in the antidepressant and memory promoting effects of morin in mice. *Drug Dev. Res.* 2019, 80, 1071–1079. [CrossRef]
- 266. Salem, H.A.; Elsherbiny, N.; Alzahrani, S.; Alshareef, H.M.; Abd Elmageed, Z.Y.; Ajwah, S.M.; Hamdan, A.M.E.; Abdou, Y.S.; Galal, O.O.; El Azazy, M.K.A.; et al. Neuroprotective Effect of Morin Hydrate against Attention-Deficit/Hyperactivity Disorder (ADHD) Induced by MSG and/or Protein Malnutrition in Rat Pups: Effect on Oxidative/Monoamines/Inflammatory Balance and Apoptosis. *Pharmaceuticals* 2022, 15, 1012. [CrossRef]
- Kalkunte, S.S.; Singh, A.P.; Chaves, F.C.; Gianfagna, T.J.; Pundir, V.S.; Jaiswal, A.K.; Vorsa, N.; Sharma, S. Antidepressant and antistress activity of GC-MS characterized lipophilic extracts of Ginkgo biloba leaves. *Phytother. Res.* 2007, 21, 1061–1065. [CrossRef]

- Yang, J.H.; Shin, B.Y.; Han, J.Y.; Kim, M.G.; Wi, J.E.; Kim, Y.W.; Cho, I.J.; Kim, S.C.; Shin, S.M.; Ki, S.H. Isorhamnetin protects against oxidative stress by activating Nrf2 and inducing the expression of its target genes. *Toxicol. Appl. Pharmacol.* 2014, 274, 293–301. [CrossRef]
- Gong, G.; Guan, Y.Y.; Zhang, Z.L.; Rahman, K.; Wang, S.J.; Zhou, S.; Luan, X.; Zhang, H. Isorhamnetin: A review of pharmacological effects. *Biomed. Pharmacother.* 2020, 128, 110301. [CrossRef]
- Pérez-González, A.; Rebollar-Zepeda, A.M.; León-Carmona, J.R.; Galano, A. Reactivity indexes and O-H bond dissociation energies of a large series of polyphenols: Implications for their free radical scavenging activity. *J. Mex. Chem. Soc.* 2012, 56, 241–249. [CrossRef]
- 271. Mohammadi, S.; Golshani, Y. Neuroprotective effects of rhamnazin as a flavonoid on chronic stress-induced cognitive impairment. J. Adv. Neurosci. Res. 2017, 4, 30–37. [CrossRef]
- 272. Gulsheen Kumar, A.; Sharma, A. Antianxiety and antidepressant activity guided isolation and characterization of gossypetin from *Hibiscus sabdariffa* Linn. Calyces. J. Biol. Act. Prod. Nat. 2019, 9, 205–214. [CrossRef]
- 273. García-Durán, L.; Flores-Burgess, A.; Cantero-García, N.; Puigcerver, A.; Narváez, J.Á.; Fuxe, K.; Santín, L.; Millón, C.; Díaz-Cabiale, Z. Galanin(1–15) Potentiates the Antidepressant-like Effects Induced by Escitalopram in a Rat Model of Depression. *Int. J. Mol. Sci.* 2021, 22, 10848. [CrossRef] [PubMed]
- 274. Dai, Y.; Zhang, H.; Zhang, J.; Yan, M. Isoquercetin attenuates oxidative stress and neuronal apoptosis after ischemia/reperfusion injury via Nrf2-mediated inhibition of the NOX4/ROS/NF-κB pathway. *Chem. Biol. Interact.* 2018, 284, 32–40. [CrossRef]
- 275. Butterweck, V.; Hegger, M.; Winterhoff, H. Flavonoids of St. John's Wort reduce HPA axis function in the rat. *Planta Med.* **2004**, 70, 1008–1011. [CrossRef]
- 276. Song, A.; Wu, Z.; Zhao, W.; Shi, W.; Cheng, R.; Jiang, J.; Ni, Z.; Qu, H.; Qiaolongbatu, X.; Fan, G.; et al. The Role and Mechanism of Hyperoside against Depression-like Behavior in Mice via the NLRP1 Inflammasome. *Medicina* **2022**, *58*, 1749. [CrossRef]
- 277. Haas, J.S.; Stolz, E.D.; Betti, A.H.; Stein, A.C.; Schripsema, J.; Poser, G.L.; Rates, S.M. The anti-immobility effect of hyperoside on the forced swimming test in rats is mediated by the D2-like receptors activation. *Planta Med.* **2011**, *77*, 334–339. [CrossRef]
- 278. Portincasa, P.; Bonfrate, L.; Vacca, M.; De Angelis, M.; Farella, I.; Lanza, E.; Khalil, M.; Wang, D.Q.-H.; Sperandio, M.; Di Ciaula, A. Gut Microbiota and Short Chain Fatty Acids: Implications in Glucose Homeostasis. *Int. J. Mol. Sci.* 2022, 23, 1105. [CrossRef]
- 279. Jiang, H.; Ling, Z.; Zhang, Y.; Mao, H.; Ma, Z.; Yin, Y.; Wang, W.; Tang, W.; Tan, Z.; Shi, J.; et al. Altered fecal microbiota composition in patients with major depressive disorder. *Brain Behav. Immun.* **2015**, *48*, 186–194. [CrossRef]
- Song, J.; Ma, W.; Gu, X.; Zhao, L.; Jiang, J.; Xu, Y.; Zhang, L.; Zhou, M.; Yang, L. Metabolomic signatures and microbial community profiling of depressive rat model induced by adrenocorticotrophic hormone. J. Transl. Med. 2019, 17, 224. [CrossRef]
- 281. Ait Chait, Y.; Mottawea, W.; Tompkins, T.A.; Hammami, R. Unravelling the antimicrobial action of antidepressants on gut commensal microbes. *Sci. Rep.* 2020, *10*, 17878. [CrossRef]
- Caldara, M.; Marmiroli, N. Antimicrobial Properties of Antidepressants and Antipsychotics—Possibilities and Implications. *Pharmaceuticals* 2021, 14, 915. [CrossRef]
- 283. Liu, S.; Guo, R.; Liu, F.; Yuan, Q.; Yu, Y.; Ren, F. Gut Microbiota Regulates Depression-like Behavior in Rats through the Neuroendocrine-Immune-Mitochondrial Pathway. *Neuropsychiatr. Dis. Treat.* 2020, *16*, 859–869. [CrossRef]
- Chen, Y.; Xu, J.; Chen, Y. Regulation of Neurotransmitters by the Gut Microbiota and Effects on Cognition in Neurological Disorders. *Nutrients* 2021, 13, 2099. [CrossRef]
- 285. Yano, J.M.; Yu, K.; Donaldson, G.P.; Shastri, G.G.; Ann, P.; Ma, L.; Nagler, C.R.; Ismagilov, R.F.; Mazmanian, S.K.; Hsiao, E.Y. Indigenous bacteria from the gut microbiota regulate host serotonin biosynthesis. *Cell* **2015**, *161*, 264–276. [CrossRef]
- 286. Donati Zeppa, S.; Ferrini, F.; Agostini, D.; Amatori, S.; Barbieri, E.; Piccoli, G.; Sestili, P.; Stocchi, V. Nutraceuticals and Physical Activity as Antidepressants: The Central Role of the Gut Microbiota. *Antioxidants* **2022**, *11*, 236. [CrossRef]
- Wang, X.; Yu, J.; Zhang, X. Dietary Polyphenols as Prospective Natural-Compound Depression Treatment from the Perspective of Intestinal Microbiota Regulation. *Molecules* 2022, 27, 7637. [CrossRef]
- Inserra, A.; Rogers, G.B.; Licinio, J.; Wong, M. The microbiota-inflammasome hypothesis of major depression. *Bioessays* 2018, 40, 1800027. [CrossRef]
- Carlessi, A.S.; Borba, L.A.; Zugno, A.I.; Quevedo, J.; Réus, G.Z. Gut microbiota-brain axis in depression: The role of neuroinflammation. *Eur. J. Neurosci.* 2021, 53, 222–235. [CrossRef]
- 290. Yang, J.; Zheng, P.; Li, Y.; Wu, J.; Tan, X.; Zhou, J.; Sun, Z.; Chen, X.; Zhang, G.; Zhang, H.; et al. Landscapes of bacterial and metabolic signatures and their interaction in major depressive disorders. *Sci. Adv.* **2020**, *6*, eaba8555. [CrossRef]
- 291. Strandwitz, P.; Kim, K.H.; Terekhova, D.; Liu, J.K.; Sharma, A.; Levering, J.; McDonald, D.; Dietrich, D.; Ramadhar, T.R.; Lekbua, A.; et al. GABA-modulating bacteria of the human gut microbiota. *Nat. Microbiol.* **2019**, *4*, 396–403. [CrossRef] [PubMed]
- Aizawa, E.; Tsuji, H.; Asahara, T.; Takahashi, T.; Teraishi, T.; Yoshida, S.; Ota, M.; Koga, N.; Hattori, K.; Kunugi, H. Possible association of Bifidobacterium and Lactobacillus in the gut microbiota of patients with major depressive disorder. *J. Affect. Disord.* 2016, 202, 254–257. [CrossRef] [PubMed]
- Filosa, S.; Di Meo, F.; Crispi, S. Polyphenols-gut microbiota interplay and brain neuromodulation. *Neural Regen. Res.* 2018, 13, 2055–2059. [CrossRef] [PubMed]
- 294. Kumar Singh, A.; Cabral, C.; Kumar, R.; Ganguly, R.; Kumar Rana, H.; Gupta, A.; Rosaria Lauro, M.; Carbone, C.; Reis, F.; Pandey, A.K. Beneficial Effects of Dietary Polyphenols on Gut Microbiota and Strategies to Improve Delivery Efficiency. *Nutrients* 2019, 11, 2216. [CrossRef]

- 295. Espín, J.C.; González-Sarrías, A.; Tomás-Barberán, F.A. The gut microbiota: A key factor in the therapeutic effects of (poly)phenols. *Biochem. Pharmacol.* 2017, 139, 82–93. [CrossRef]
- 296. Westfall, S.; Pasinetti, G.M. The Gut Microbiota Links Dietary Polyphenols with Management of Psychiatric Mood Disorders. *Front. Neurosci.* **2019**, *13*, 1196. [CrossRef]
- 297. Zhou, N.; Gu, X.; Zhuang, T.; Xu, Y.; Yang, L.; Zhou, M. Gut Microbiota: A Pivotal Hub for Polyphenols as Antidepressants. *J. Agric. Food Chem.* **2020**, *68*, 6007–6020. [CrossRef]
- 298. Feng, J.; Li, Z.; Ma, H.; Yue, Y.; Hao, K.; Li, J.; Xiang, Y.; Min, Y. Quercetin alleviates intestinal inflammation and improves intestinal functions via modulating gut microbiota composition in LPS-challenged laying hens. *Poult. Sci.* 2023, 102, 102433. [CrossRef]
- Sun, L.; Guo, L.; Xu, G.; Li, Z.; Appiah, M.O.; Yang, L.; Lu, W. Quercetin Reduces Inflammation and Protects Gut Microbiota in Broilers. *Molecules* 2022, 27, 3269. [CrossRef]
- Silva, Y.P.; Bernardi, A.; Frozza, R.L. The Role of Short-Chain Fatty Acids from Gut Microbiota in Gut-Brain Communication. Front. Endocrinol. 2020, 11, 25. [CrossRef]
- Lu, X.Y.; Kim, C.S.; Frazer, A.; Zhang, W. Leptin: A potential novel antidepressant. Proc. Natl. Acad. Sci. USA 2006, 103, 1593–1598.
  [CrossRef]
- 302. Abdel-Latif, M.A.; Elbestawy, A.R.; El-Far, A.H.; Noreldin, A.E.; Emam, M.; Baty, R.S.; Albadrani, G.M.; Abdel-Daim, M.M.; Abd El-Hamid, H.S. Quercetin Dietary Supplementation Advances Growth Performance, Gut Microbiota, and Intestinal mRNA Expression Genes in Broiler Chickens. *Animals* 2021, 11, 2302. [CrossRef]
- 303. Gu, F.; Wu, Y.; Liu, Y.; Dou, M.; Jiang, Y.; Liang, H. Lactobacillus casei improves depression-like behavior in chronic unpredictable mild stress-induced rats by the BDNF-TrkB signal pathway and the intestinal microbiota. *Food Funct.* 2020, 11, 6148–6157. [CrossRef]
- 304. Tamura, M.; Hoshi, C.; Kobori, M.; Takahashi, S.; Tomita, J.; Nishimura, M.; Nishihira, J. Quercetin metabolism by fecal microbiota from healthy elderly human subjects. *PLoS ONE* **2017**, *12*, e0188271. [CrossRef]

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