







REVIEW

Collecting eco-evolutionary data in the dark: Impediments to subterranean research and how to overcome them

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Abstract

1. Caves and other subterranean habitats fulfill the requirements of experimental model systems to address general questions in ecology and evolution. Yet, the harsh working conditions of these environments and the uniqueness of the subterranean organisms have challenged most attempts to pursue standardized research.
2. Two main obstacles have synergistically hampered previous attempts. First, there is a *habitat impediment* related to the objective difficulties of exploring subterranean habitats and our inability to access the network of fissures that represents the elective habitat for the so-called “cave species.” Second, there is a *biological impediment* illustrated by the rarity of most subterranean species and their low physiological tolerance, often limiting sample size and complicating laboratory experiments.
3. We explore the advantages and disadvantages of four general experimental setups (*in situ*, *quasi in situ*, *ex situ*, and *in silico*) in the light of habitat and biological impediments. We also discuss the potential of indirect approaches to research. Furthermore, using bibliometric data, we provide a quantitative overview of the model organisms that scientists have exploited in the study of subterranean life.
4. Our over-arching goal is to promote caves as model systems where one can perform standardized scientific research. This is important not only to achieve an in-depth understanding of the functioning of subterranean ecosystems but also to fully exploit their long-discussed potential in addressing general scientific questions with implications beyond the boundaries of this discipline.

KEYWORDS

anchialine, *Asellus aquaticus*, *Astyanax*, cave laboratory, computer simulations, experimental design, groundwater, model system, natural laboratory, nonmodel organisms, sampling strategy, stygobite, troglodite

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1 | INTRODUCTION

For a *Homo sapiens*—a clumsy vertebrate inhabiting a primarily lighted world—to enter a cave is enterprising. As the sunlight fades, the air becomes moist, and a maze of passages opens in front of us, our first instinct as humans is to dismiss the subsurface world as one of the most inhospitable environments on Earth. Mentions to this apparent extremeness emerge in most caving stories (MacNeil & Brcic, 2017) insofar as speleology is indeed physically demanding and potentially hazardous (Zagmajster et al., 2010). However, by over-emphasizing this anthropocentric view of caves, we tend to dismiss a different reality: Caves are not so extreme from the perspective of the eyeless and depigmented organisms that have adapted to living in darkness, which in contrast experience the exposure to sunlight and the wide climatic fluctuation of the outside world as harmful threats (Mammola, 2020). Interestingly, this dichotomous interpretation has framed the two main approaches followed by researchers over recent years: Those who have studied subterranean habitats as unique entities versus those who have used them as model to answer general scientific questions beyond the boundaries of subterranean biology (Martínez & Mammola, 2020).

Scientists across several generations have been aware of the potential of subterranean ecosystems (Box 1) as eco-evolutionary models (Poulson & White, 1969), developing innovative methodologies and creative experimental designs to face the challenges associated with subterranean exploration. Thanks to these efforts, we have been able to tackle important subjects in ecology (Mammola, 2019), ethology (Parzefall, 1982), and evolution (Juan et al., 2010), ultimately reaching conclusions relevant to disciplines as diverse as medicine (Riddle et al., 2018; Stockdale et al., 2018; Yoshizawa et al., 2018), engineering (Lepore et al., 2012), and exobiology (Northup et al., 2011). Under this perspective, and despite the numerous obstacles to research, subterranean habitats may well qualify as frontiers for modern scientific research (Mammola et al., 2020).

In this work, we discuss the main impediments that we must address to standardize research in subterranean ecosystems and, subsequently, we illustrate old solutions, recommend best practices, and advance new frontiers to approach subterranean-based studies (Figure 1). By further elaborating on the established model organisms in subterranean biology, we seek to promote caves and other subterranean habitats as experimental arenas for asking general questions in ecology, ethology, evolution, and beyond. In other words, we call for moving caves from the niche of an exotic, understudied environment to the forefront of biological science. The gain

BOX 1 A modern definition of subterranean habitats and implications for research.

The term “subterranean habitat/ecosystem” is often used as a synonym for “cave” (Mammola, 2019; Poulson & White, 1969). However, scientists have become aware that caves represent only a small fraction of the total habitat available to the subterranean fauna. More precisely, subterranean habitats comprise the breadth of underground voids of different sizes, either dry or filled with water, sharing two main ecological features: darkness and buffered climatic conditions (Culver & Pipan, 2019). These voids may open a few centimeters below ground level (Culver & Pipan, 2014) or descend several kilometers toward areas where the environmental conditions exceed the limits of life (Fišer et al., 2014). They are widespread on all continents, having been documented from different geological substrates, including carbonates (limestone and dolomite), sandstones, gypsum, granites, lava fields, iron ores, and even unconsolidated sediments (Keith et al., 2020). In summary, the cavities that we can access and explore by entering them represent just the tip of the iceberg of what lies below our feet (Ficetola et al., 2019; Mammola, Cardoso, et al., 2019).

Even though subterranean habitats are more widespread and diversified than it is usually recognized, subterranean research started with field observations in human-accessible habitats (different types of terrestrial caves, artificial subterranean habitats such as mines and bunkers, lava tubes, cenotes, etc.), later encompassed pumped water (e.g., from drinking water wells), and only then extended to other difficult-to-access voids. Even today, there is still a significant research bias toward human-accessible habitats, which should always be kept in mind. In a nutshell, it implies that we may have to relativize part of the information available to date, that is, being aware that we have mostly documented how animals behave in cave-like environments, rather than in the extended network of fissures.

in doing so lies in the controlled conditions they offer, since they are little exposed to influences that one would have to control for in other environments.

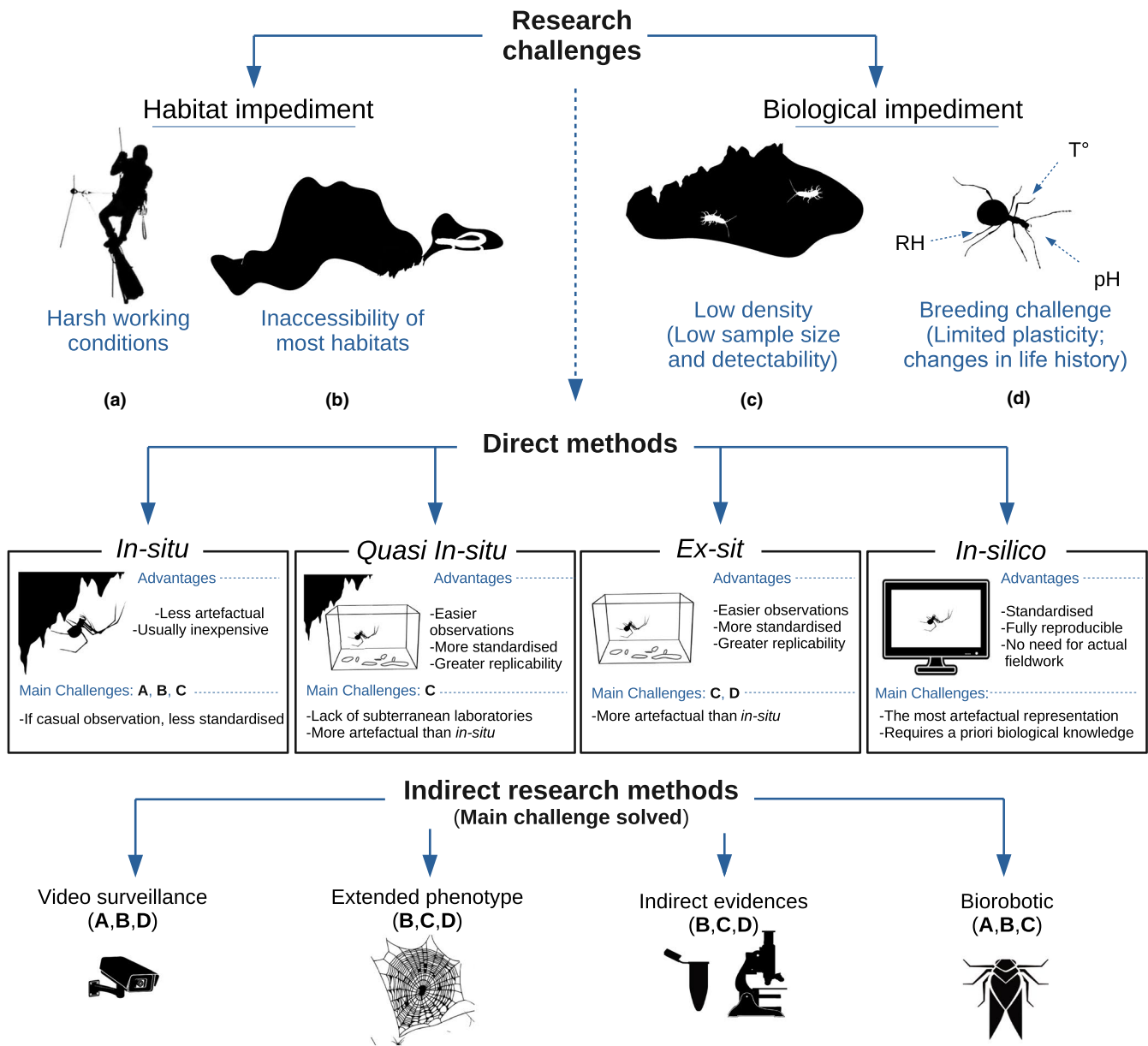


FIGURE 1 Challenges of subterranean research and experimental designs to avoid these. Schematic representation of the main challenges of subterranean research (coded with capital letters), and main experimental approaches that can be adopted to overcome these

2 | RATIONALE FOR THIS WORK AND COMPLEMENTARY REVIEWS

It is impossible to cover all methods in subterranean biology while keeping this review tight and comprehensible. Therefore, we decided to focus on the main challenges related to subterranean research and the philosophy underlying the different experimental designs suited to overcome these: Two aspects only marginally discussed in the recent literature. Readers interested in other aspects of research in subterranean biology are referred to the classic review on biomonitoring (Culver & Sket, 2002) and published syntheses on sampling approaches (Dole-Olivier

et al., 2009; Lunghi, Corti, et al., 2020; Oliveira et al., 2019; Weinstein & Slaney, 1995; Wynne et al., 2018, 2019), species distribution modeling (Mammola & Leroy, 2018), and best practices in experimental trials with subterranean organisms (Di Lorenzo et al., 2019). Sampling techniques in non cave subterranean habitats (Box 1) have also been reviewed elsewhere—for example, boreholes (Hancock & Boulton, 2009), epikarst (Brancelj, 2004), subaquatic caves (Humphreys et al., 1999; Iliffe, 2018; Iliffe & Bowen, 2001), *Milieu Souterrain Superficiel* (Mammola et al., 2016), hyporheic (Fraser & Williams, 1997), and interstitial habitats in coastal marine and lotic environments (Schmidt-Rhaesa, 2020).

3 | CHALLENGES TO SUBTERRANEAN RESEARCH

3.1 | Habitat impediment

Whereas different habitats have been categorized as subterranean (Box 1), most in-field research takes place in caves, mines, and other human-accessible voids (Mammola, 2019). These are always dark, often muddy and humid, and sometimes even very cold, hence not offering favorable conditions to perform extensive, standardized observations (MacNeil & Brcic, 2017). There are often high ceilings, narrow fissures, and other geomorphological features that hamper the task of approaching and observing target animals without them being disturbed by light or by the presence of the researcher (CO_2 , heat, vibrations, or even diver's bubbles in the case of submerged passages). Furthermore, cave exploration requires well-trained researchers mastering the use of speleological equipment (Zagmajster et al., 2010). Even more challenging, in this sense, are those studies set in submerged passages of freshwater and marine caves (Exley, 1983; Iliffe & Bowen, 2001), as testified by the frequent fatalities associated with cave diving (Buzzacott et al., 2009).

Given our human size, we can directly access only a small fraction of the habitats available to the subterranean fauna. As noted by Howarth (1983) (p. 380), this is a significant obstacle to scientific research because, more often than not, we cannot directly inspect the extended network of fissures “[...] where probably the major drama in the cave ecosystem occurs”. In a way, caves and other human-accessible habitats may act as surrogates of the subterranean world in its entirety, windows allowing us to glimpse what usually happens away from human sight (Mammola et al., 2016; Polak, 1997; Uéno, 1987; Wilkens et al., 1986). Yet, in this case the existence of a habitat bias should be clearly acknowledged. For instance, we must be aware that the foraging behavior of a centipede that we have observed in a large chamber of a cave may not replicate in the same way—or may not even take place at all!—when the exact same centipede is dwelling in the millimetric fissures connected with the chamber. Not to mention certain typically benthic aquatic animals that have been spotted in the water column of flooded caves only after the disturbance produced by the divers (Humphreys et al., 1999).

As a corollary, however, it must be noted that a number of organisms primarily belong to human-accessible cavities (Moseley, 2009) and *ipso facto* are more readily studied (Mammola, 2019). Classic examples are vertebrates with a centimetric body size, such as different species of cave-roosting bats and groundwater fishes, but also the parasites and commensals associated with them (Lunghi, Ficetola, et al., 2018) or the scavengers that feed upon their carcasses and feces (Ferreira & Martins, 1999). There are also subterranean invertebrates constrained to human-sized voids by their extended phenotypes; notably, different species of orb spiders needing larger voids for web construction (Mammola & Isaia, 2017) or aquatic suspension feeders adapted to drift in the still water column of anchialine caves (Koenemann et al., 2007; Martínez et al., 2017).

Finally, in specific subterranean systems, there may be health risks related to biological diseases or toxic gases, potentially hampering or complicating explorations and studies. Examples include fungi [*Histoplasma capsulatum* (Eurotiomycetes: Ajellomycetaceae) causing histoplasmosis; Hunt et al., 1984; Diaz, 2018; Staffolani et al., 2018], and viruses, such as Marburg virus associated with fruit bats roosting in caves (Kuzmin et al., 2010) and the potential presence of SARS-CoV-2 coronavirus in touristic caves (Barton, 2020). Furthermore, cave-roosting bats may be vectors of rabies and a number of other emerging diseases (Calisher et al., 2006; Kuzmin et al., 2011)—amidst the COVID-19 pandemic in 2020, these potential zoonotic health risks have unfortunately resulted in persecutions of bats (MacFarlane & Rocha, 2020; Rocha et al., 2020). Another human health threat may come from contaminated air due to the accumulation of hazardous gases including CO, CO_2 , and sulfur exhalations. Also, the concentration of radon (a radioactive gas) may be elevated in the subterranean realm (Cigna, 2005; Gillmore et al., 2000)—in certain caves its levels can exceed recommended doses 20-fold.

3.2 | Biological impediment

In several cases, the biology of subterranean species represents a further impediment to research. In general, food-deprived subterranean environments select for long-lived species with low metabolism and small numbers of offspring. As a consequence, the density of individuals of subterranean adapted species is often low—it is not unusual that such species were observed once at the time of their description, and never recorded thereafter (Delić & Sket, 2015; Manenti et al., 2018; Martínez et al., 2013). Also, specialized subterranean species are often unevenly distributed in space and time, mostly because they aggregate around the scarce and heterogeneously distributed food sources (Culver & Sket, 2002). These difficulties in finding sufficient individuals for experiments or in situ observations may result in studies with a reduced sample size and less robust data. This may explain why the ecology and behavior of many subterranean organisms is documented, at best, anecdotally thanks to casual observations.

Furthermore, many specialized subterranean organisms live in environments showing constant and buffered conditions and, over evolutionary time, have reduced their resilience against environmental fluctuations. For example, some terrestrial subterranean species are threatened by the smallest variations in air moisture content (Howarth, 1983), whereas aquatic animals may perish upon changes in pH driven by the water exposure to the air (Carpenter, 1999). Similarly, many terrestrial and aquatic obligate subterranean species survive only within narrow temperature ranges (Mammola, Piano, et al., 2019; Merrilod-Blondin et al., 2013; Pallarés, Colado, et al., 2020; Pallarés, Sanchez-Hernandez, et al., 2020). This limited physiological plasticity may pose a real challenge when a researcher is aiming to conduct experiments in the unnatural conditions of a typical laboratory. Maintaining living individuals of most of these animals is not a trivial

task: breeding them requires skill and experience, in-depth knowledge of their biology and, often, none negligible doses of luck.

In some cases, an additional impediment may be our lack of knowledge on the taxonomy of subterranean organisms (e.g., Asmyhr & Cooper, 2012; Camacho et al., 2018). This impediment (also known as 'Linnean shortfall'; Hortal et al., 2015) is especially problematic in studies focusing on community structure and functioning. It is particularly severe for tropical areas, whose subterranean fauna were largely unknown until very recently (e.g., Alvarenga et al., 2021; Trajano & Bichuette, 2010; Trajano et al., 2016), as well as for certain small-sized animal groups which have been traditionally neglected despite being relatively abundant in subterranean habitats, such as diplurans (Sendra, Antić, et al., 2020; Sendra, Palero, et al., 2020), proturans (Galli et al., 2021), palpi-grades (Mammola et al., 2021), nematodes (Du Preez et al., 2017), gastrotrichs (Kolicka et al., 2017), and other meiofaunal lineages (Martínez et al., 2019; Sánchez & Martínez, 2019). The lack of taxonomists for many groups surely hampers the conduction of more concise studies on cave communities in many regions; a situation that is further aggravated by the existence of often high cryptic diversity within most subterranean taxa (Delić et al., 2017; Eme et al., 2018; Esposito et al., 2015; Fišer et al., 2018; Gonzalez et al., 2017; Niemiller et al., 2012).

4 | EXPERIMENTAL SETUPS

The most classical and intuitive way to learn about subterranean organisms lies in quantitative observational studies, either in the field (*in situ*), under laboratory conditions (*ex situ*) or, when available, in laboratories set within caves (here termed *quasi in-situ*). An experimental setup entirely based on simulations (*in silico*) could also be adopted. The choice among these setups is not always

straightforward. In general, choosing between alternative options is a trade-off between the biological realism of the observations and either the ease or the extensiveness of the study (Figure 2). More detailed pros and cons of each setup are discussed in the following sections.

4.1 | *In situ*

The *in-situ* approach provides the least artefactual representation of the ecology, physiology, and behavior of the target species. Yet, this approach forces the researcher to comply with both the habitat (harsh working conditions and impossibility of exploring inaccessible habitats) and the biological (low density of most subterranean species) impediments. To minimize these impediments, a careful selection of the study site is critical. If possible, one should favor cavities with a linear development and reduced habitat complexity, thereby facilitating standardized observations (Lunghi, Corti, et al., 2020; Mammola & Isaia, 2018; Smithers, 2005) while maximizing detectability of the animals (Lunghi, 2018). In the same vein, studying aquatic target species in a semi-submerged or shallow passageway not only increases permanence times and minimizes decompression procedures, but also maximizes safety (Iliffe, 2018).

It must be noted that *in-situ* studies can be carried out exclusively in cavities the researcher can enter herself or at least insert instruments into. There are different types of traps and sampling devices that allow us to indirectly collect the fauna in inaccessible and interstitial habitats or even tools for detecting the presence of a species indirectly (see section "Indirect means of research"). Conversely, *in-situ* observations are virtually impossible for porous groundwater, forcing researchers to heavily rely on laboratory studies (e.g., Di Lorenzo et al., 2014).

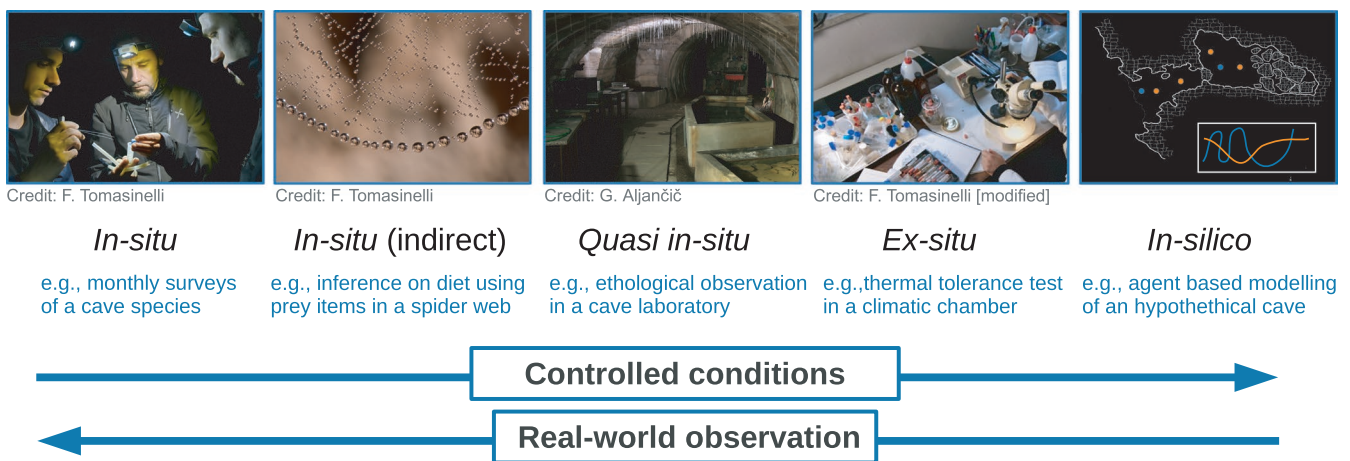


FIGURE 2 A theoretical trade-off between the ease of study and biological realism of the observations in different experimental setups. On the one hand, exploring a cave is physically demanding and requires specific speleological equipment, whereas it is possible to run a simulation sitting at home in front of a computer in a pyjama—and even during a COVID-19 pandemic! Running a simulation or conducting an experiment in the laboratory also allows us to control for a number of confounding factors. On the other hand, the result obtained in the field is often less artefactual, requiring no abstraction or formulation of a priori assumptions. At some point, when studying phenomena in the laboratory or with simulations, one will want to get back to the field to corroborate results using real-world observations

4.2 | Ex-situ

The use of a meso- or microcosmos replicating the species' natural habitat allows us to bypass the habitat impediment in its entirety. In general, obtaining standardized observations in controlled conditions enhance a greater replicability of the results, and potentially allow researchers to perform long-lasting experiments and observations—see Carpenter (2021) for a recent example. Furthermore, *ex-situ* approaches permit to explore the life history of those animals that prefer inaccessible habitat, or that are too small to be observed with the naked eye. Yet, by choosing an *ex-situ* approach, the researcher needs to comply with the biological impediment of maintaining specialized and delicate organisms in the laboratory (Di Lorenzo et al., 2019), as well as with the local conservation policies for endangered species. This can be circumvented by selecting certain model organisms, often not legally protected and relatively easy to keep in the laboratory (see section “Model organisms in subterranean biology”).

As a drawback, laboratory observations may not accurately reflect the natural traits, especially behavioral and physiological, as shown in the cave (Silva et al., 2018). Although laboratory studies are useful, the *ex-situ* conditions rarely resemble those found in the natural habitat (e.g., higher density, different environmental conditions). For well-established model organisms, a prolonged *ex-situ* breeding may even produce unwelcome effects such as artificial selection or adaptation to the laboratory conditions (Ross et al., 2019). This is why observations obtained from studies in the laboratory must be carefully interpreted and preferably confirmed by *in situ* approaches (Blin et al., 2020). For example, by surveying semi-natural replicas of the sheltered reproductive sites of *Hydromantes* salamanders (Amphibia: Plethodontidae) with infrared cameras, Oneto et al. (2010) were able to provide some of the first observational data on their complex reproductive behavior and parental care. Subsequent observations performed under natural conditions (Lunghi, Corti, et al., 2018; Lunghi et al., 2014, 2015) confirmed the validity of these observations.

4.3 | Quasi in-situ

The history of subterranean biology teaches us that a *quasi in-situ* approach—that is, to bring the laboratory into the target species' natural habitat—eases many of the problems associated with experimental studies in the laboratory. Establishing an experimental facility within the cave itself not only spare living animals from long transportation away from the cave, but also facilitates fine-regulation of ambient parameters within a microcosmos. The most famous example is probably the *Laboratoire Souterrain de Moulis* (*Centre national de la recherche scientifique*; CNRS), a cave-based laboratory established in the French Pyrenees by René Jeannel (1879–1965) and Albert Vandel (1894–1980). Since its foundation in 1948, this semi-natural experimental setting has aided generations of subterranean biologists in the challenging task of shedding light on the natural history

and behavior of a wide range of elusive subterranean life forms (Clergue-Gazeau, 1974; Durand, 1970; Juberthie, 1985; Juberthie et al., 1996; Manenti et al., 2020). For aquifers, the equivalent would be to lower sediment, substrate cages, bags into groundwater wells (Schmidt et al., 2004), which, however, would still have to be retrieved every time to study the organisms.

As a corollary, it must be noted that establishing an experimental facility inside a given cave may have a significant local impact in terms of destruction of certain microhabitats and/or alterations of the microclimate. Therefore, the establishment of similar infrastructures should be evaluated on a case-by-case basis and supported by an environmental risk assessment.

4.4 | In-silico

As a consequence of the habitat and biological impediments, studies in subterranean habitats often rely on data that is far from ideal. In a complex subterranean setting, we may lack information on environmental seasonal fluctuations, species abundances across space or time, their physiological rates and life-history traits, or the species they interact with. Not to mention the dependency between observations and the correlation among traits (body size and trophic guild, fecundity with longevity, etc.), which often confounds with putative drivers for the process that we aim to disentangle. In those scenarios, simulations, such as agent-based models and cellular automata, are increasingly used to explore the dynamics of natural ecosystems and trigger novel ideas for further exploration in real-world settings (DeAngelis & Grimm, 2014). These mechanistic models rely on the so-called “first principles,” such as energy budgets, physiology, or fitness seeking (Grimm & Berger, 2016), which define the initial conditions of the simulation so that behavior and interactions emerge rather than being imposed by the modeler. Given robust enough assumptions, simulations are thus able to realistically replicate sets of empirical patterns without restricting them to a single deterministic scenario (Grimm et al., 2005). For example, the use of eco-evolutionary agent-based models, which include heritable traits and the use of genetic algorithms, provides insights on the evolution of certain morphological, physiological, and behavioral traits (Ayllón et al., 2018).

Surprisingly, however, simulations have rarely been applied in subterranean biology. Applications to subsurface systems so far have been restricted to porous groundwater, with the focus being mainly on contaminant degradation (Beniouge et al., 2015, 2017; Schmidt et al., 2018; Tang et al., 2013), and to soils (e.g., Banitz et al., 2013; Borer et al., 2019; Kim & Or, 2016). It is easy to see how the simulation of a virtual cave would be an interesting aid to research. Caves may represent ideal model systems for *in-silico* studies due to their constant environmental conditions, which can be easily and predictably simulated, and their simple community structure with few species and limited interactions. For example, these models would allow us to achieve a mechanistic understanding of the processes behind interactions between species within a typical subterranean community, to explore pathways of

subterranean evolution, and even to elucidate the impact of climate change on subterranean biodiversity.

The applicability of these theoretical models to the real biological world, however, still depends on the quality and availability of data. Parametrization of simulations might be relatively simple for broad questions in spatial or temporal scope, but quite complex for very specific systems, often implying the need for possessing detailed information. Thus, and this is true for other methods as well, the necessity to parametrize theoretical models with the real-world biological observations may require combining simulation approaches with actual fieldwork. Importantly, models may single out those parameters that warrant the most attention and may thus steer experiments toward focussing on sensitive and critical parameters. A complementary avenue is combining qualitative observations, for example, that state changes are confined within a certain interval, for parameterization. Even if a single observation does not contain much information, a combination of several qualitative observations can be as distinctive as a single high-precision observation. This inverse, “pattern-oriented” parameterization (Grimm et al., 2005; Wiegand et al., 2004), has been proven to be a powerful approach and overlaps with the more formal approximate Bayesian computing approach (Hartig et al., 2011).

4.5 | Indirect means of research

A plethora of indirect methodologies can be used to overcome both the habitat and the biological impediments (Figure 1). These approaches are mostly species- and system-specific, and it is impossible to provide widely general recommendations. Therefore, we here discuss examples chosen to illustrate the concept of “indirect research.”

Information about the ecology and behavior of large-sized animals can be acquired via infrared video surveillance. This represents a low-cost and low-personnel effort methodology, which has a long tradition in ethological research and biomonitoring (Swann et al., 2004). In caves, thermal-infrared imaging and laser scanning have been extensively applied to study the swarming and roosting behaviors of bats (Azmy et al., 2012; Elliott et al., 2005), but could potentially be used for other vertebrates, such as cave salamanders (Lunghi, Manenti, et al., 2020). In at least one case, camera trapping has even been used to quantifying wildlife use of cave entrances (Baker, 2015).

Recently, there has also been a great deal of discussion on the use of molecular tools to obtain indirect evidence of the presence and behavior of species, especially in difficult-to-access habitats, as well as to overcome prevalent taxonomic biases (Malard et al., 2020). For example, environmental DNA was successfully used to detect the presence of focal subterranean species, such as amphibians (Gorički et al., 2017) and crustaceans (Boyd et al., 2020; DiStefano et al., 2020; Niemiller et al., 2018). The analysis of gut or stomach content of species inhabiting both human-accessible and interstitial environments provides information on dietary requirements and trophic behaviors taking place in both these compartments

(Lunghi, Cianferoni, et al., 2018; Lunghi, Manenti, et al., 2020), but also trophic web studies with aquatic subterranean species (Saccò et al., 2019). These analyses can be done visually, but also through massive sequencing techniques, allowing the identification of the gut content using DNA (Rastorgueff et al., 2015). Similarly, stable isotopes proved useful to understand species interactions and niche partitioning (Chávez-Solís et al., 2020), as well as identifying potential carbon sources through space (Brankovits et al., 2017) and time (Saccò et al., 2020).

In some circumstances, the species' extended phenotype also informs indirectly on specific behaviors and ecological needs. The web in web-building spiders, for example, can be viewed as an extended phenotype that enlarges the sensory world of its builder in interaction with the environment (Blamires, 2010). The web also provides a record frozen in time of the spider's foraging behavior, as spiders modify their webs in response to a large range of biotic and abiotic stimuli, including previous prey experiences, climatic variables, and the structural complexity of the habitat (Hesselberg, 2015; Vollrath & Selden, 2007). The easily quantifiable two-dimensional orb-web, in particular, is highly suitable for behavioral studies, as orb spiders can easily be maintained in the laboratory (Zschokke & Herberstein, 2005) or their webs measured in the field (Hesselberg, 2010). The ubiquity of orb-web spiders near the entrance of temperate caves makes this approach especially promising (Hesselberg et al., 2019). Likewise, the calcified tubes of several hard-bodied aquatic organisms, such as tube-building polychaetes, bring us information on the evolution of aquatic caves communities and paleoclimate from past geological eras (Moldovan et al., 2011).

The living world has long been used as a source for developing biologically inspired robots using biomimetics design principles to provide innovative technical solutions (Lenau et al., 2018; Pfeifer et al., 2007; Vincent et al., 2006). In recent years, the use of biorobotic models to test and generate biological hypotheses has been gaining ground (Gravish & Lauder, 2018). Following this recent trend, we propose that the use of small, agile biorobots to explore, record, and interact with subterranean animals in their natural habitats might overcome many of the habitat and biological impediments previously discussed (Woodward & Sitti, 2014). For example, the use of a simple biomimetic robot fish has been successfully used to highlight similarities and differences in social behavior between surface and cave-dwelling populations of *Poecilia mexicana* (Actinopterygii: Poeciliidae) (Bierbach et al., 2018).

5 | MODEL ORGANISMS IN SUBTERRANEAN BIOLOGY

Model organisms represent only a small part of Earth's biodiversity and yet have largely contributed to our knowledge on many fields within the biological sciences (Hedges, 2002). The earliest models, such as flies, mice, or roundworms, were selected for the task simply because they were small, proliferative, and easy to culture and manipulate; they were, however, quite limiting in advancing many

aspects in ecology and evolution. Luckily, the growth of modern molecular methods, staining and imaging techniques, and gene editing, have facilitated choosing more appropriate models for the biological question at hand rather than enforcing the ones that can be easily grown and manipulated (Müller & Grossniklaus, 2010). Consequently, the number of model species has diversified along with the number scientific questions, and now includes representatives of many animal phyla (as well as plants and fungi). This exciting transition in contemporary biology is embodied by the term “non-model” organism, which reflects that the diversity of model species has grown nearly parallel with the diversity of problems addressed (Goldstein & King, 2016; Russell et al., 2017; Sullivan, 2015).

The trend of diversification of model systems and research question is evident in cave biology as well. To comprehend it, we have compiled a list of those subterranean animals that can be considered as model organisms (Table S1). We selected models based on two criteria: (a) organisms/groups with accumulated at least 20 papers in the Web of Science (accessed on 25 November 2020); and (b) organisms/groups with at least two independent research laboratories focusing on them. Our list of model organisms includes species with different degrees of subterranean specialization across three phyla, but it is dominated by Teleostei fish and Crustacea (Figure 3). This reflects the traditional research bias in subterranean biology toward these groups, only partially justified by their dominance across subterranean environments. Only a few of these species satisfy the traditional requirement of a model—successful culturing in the lab and keeping long-standing laboratory breeds [e.g., *Astyanax mexicanus* (Actinopterygii: Characidae), *Asellus aquaticus* (Isopoda: Asellidae), and *Poecilia mexicana*]. By far, the most famous and studied among these is the cavefish *Astyanax mexicanus* (Jeffery, 2020; Keene et al., 2016; Torres-Paz et al., 2018), which has been kept in captivity for many generations (Wilkens, 1971) and is increasingly used and recognized as suitable for tackling problems beyond the typical subterranean biology realm (Maher, 2009; McGaugh et al., 2020). Other models thrive in laboratory conditions, but are unable to complete their life cycle therein [e.g., *Gammarus minus* (Amphipoda: Gammaridae), Australian calcrete Dytiscidae insects]. Most models in subterranean biology are lineages with both surface and subterranean populations, or species whose populations exhibit different degrees of subterranean specialization. Among those, *Astyanax mexicanus* and *Asellus aquaticus* are even able to form hybrid offspring between cave and surface morphs in laboratory conditions (Jeffery, 2020; Protas & Jeffery, 2012).

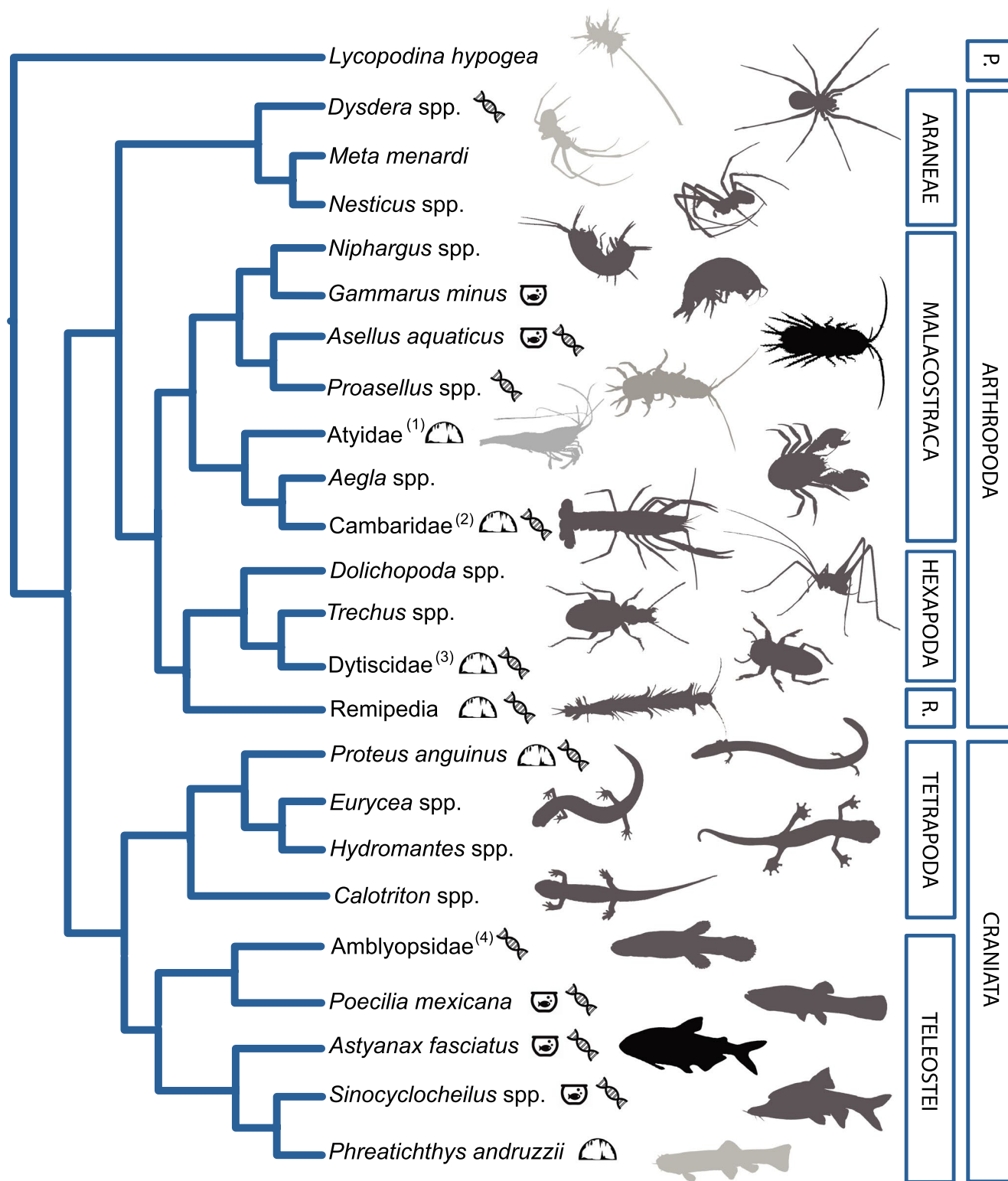
Alongside every other biological discipline, cave biology research has entered the genomics era (Friedrich, 2013; Pérez-Moreno

et al., 2016). Already half of cave models listed in Table S1 have been included in genome (transcriptome) sequencing projects, becoming windows into the molecular basis of adaptation (Barbosa et al., 2017; Berning et al., 2019). With the decreasing prices and the development of more user friendly bioinformatic recourses, so-called -omics tools will soon be at the forefront of cave research and exploited in the remaining model systems. Such tools may enable overcoming traditional restrictions on the use of subterranean species as models and we predict that the peculiar, and even bizarre, traits of subterranean animals are going to draw attention from an increasingly wider audience, and possibly attract new researchers into the field (Mammola et al., 2020).

In subterranean biology, the concept of model organism has also been applied to supra-specific lineages widely used to investigate evolutionary processes associated with cave colonization or to answer biogeographic and macroecological questions. Similar studies typically rely on comparative methods within explicit phylogenetic frameworks, allowing us to distinguish the role played by ecological adaptations and evolutionary history on the observed ecological and distribution patterns (Juan et al., 2010; Mammola et al., 2020). Some of these models account for lineages including both surface and subterranean species exhibiting different degrees of adaptations and ecological preferences, such as *Asellus* (Verovnik et al., 2004), *Niphargus* (Amphipoda: Niphargidae) (Fišer, 2009), *Trechus* (Coleoptera: Carabidae) (Möst et al., 2020), and *Dysdera* (Araneae: Dysderidae) (Arnedo et al., 2007). Others exclusively consist of subterranean species, such as atyd shrimps (Decapoda: Atyidae) of the genera *Typhlatya*, *Stygiocaris*, *Speleocaris*, and *Troglocaris* (Jurado-Rivera et al., 2017; Zakšek et al., 2009). While lineages in the first group are useful to understand different mechanisms for ecological speciation and habitat shift, subterranean-exclusive lineages allow us to understand the role of historical stochastic processes in subterranean diversity and biogeography (Juan et al., 2010). In addition, subterranean-exclusive lineages have been studied in comparison with distantly surface-dwelling relatives to understand the adaptation processes related to the colonization of the subterranean environment [e.g., *Phreatichthys andruzzii* (Actinopterygii: Cyprinidae)]. Although intuitively less ideal, this approach has yielded some important insights, such as the impacts of life in darkness on the circadian clock or DNA repair mechanisms (Cavallari et al., 2011).

Finally, some subterranean species with unique features have been established as models to investigate scientific questions not necessarily related to the classic subterranean research agenda. This is the case of the carnivorous sponge *Lycopodina hypogea* (Demospongiae: Cladorhizidae), used as a model for early nervous system evolution and developmental biology (Godefroy

FIGURE 3 Diversity of model organisms in subterranean biology across the animal Tree of Life. The branch Cambaridae refers to the genera *Cambarus*, *Orconectes*, *Procambarus*, and *Troglocambarus*. Atyidae refers to the exclusively subterranean genera *Speleocaris*, *Stygiocaris*, *Troglocaris*, and *Typhlatya*. Dytiscidae indicates the Australian diving beetles of the genera *Limbodesus*, *Nirridesus*, *Nirripiriti*, and *Paroster*. Amblyopsidae indicates the North American cave fish in the genera *Amblyopsis*, *Chologaster*, *Forbesichthys*, *Speleoplatyrhinus*, and *Typhichthys*. WoS entries: Number of papers focusing on the species in Web of Science (accessed on 25 November 2020). (1–3): The information refers to the genera (1) *Speleocaris*, *Stygiocaris*, *Troglocaris*, and *Typhlatya*; (2) *Cambarus*, *Orconectes*, *Procambarus*, and *Troglocambarus*; (3) *Paroster*, *Limbodesus*, *Nirridesus*, and *Nirripiriti*



>1000 WoS entries



50-1000 WoS entries



20-50 WoS entries

Transcriptome/Genome
assemble available

Established lab cultures



Exclusively subterranean

et al., 2019); the crustaceans in the class Remipedia, key to understanding the evolution of terrestrial arthropods (Lozano-Fernandez et al., 2016), the evolution of the nervous system (Stemme et al., 2013), and venoms toxins (von Reumont et al., 2014); or the “forever young” aquatic salamander *Proteus anguinus* (Amphibia: Proteidae), whose progenetic origin and long lifespan has triggered fruitful research on the molecular mechanisms of aging (Voituron et al., 2011). Also, *A. mexicanus* have been increasingly used as a model organism in various areas of biomedical research, such as diabetes (Riddle et al., 2018), insomnia (Jaggard et al., 2018), autism (Yoshizawa et al., 2018), and regeneration (Stockdale et al., 2018). While those are not the questions that have inspired most cave-based researchers over the years, they have recently attracted considerable interest and funding, thereby illustrating the general idea of our review here: Caves, in their uniqueness for humans, still hold the secrets for understanding broad scientific questions (Martínez & Mammola, 2020).

6 | CONCLUSIONS

In this work, we discussed best practices and novel ideas for performing standardized research in subterranean ecosystems, by focusing on key impediments, experimental ideas, and model systems. The main take-home messages that emerge from this exercise are as follows:

1. *Be aware of the many options out there.* Insofar as each subterranean system and organism is unique to some extent, and in light of the impediments to subterranean research, scientists must be creative in designing their experiments. Research in subterranean biology often implies combining traditional in situ field observations with standardized studies in a laboratory setting, either within a cave (*quasi in-situ*) or outside the cave (*ex-situ*). It is also important to be aware of the potential of novel tools, especially simulations, artificial intelligence methods, and biorobotics (Figure 1).
2. *Choose the right model.* Many impediments to subterranean research can be overcome by focusing on model organisms, which have been established owing to their specific traits and/or their broad availability in subterranean environments. Whereas model systems in subterranean biology are probably not as developed as in other disciplines, there are options across the animal tree of life offering great potential for tackling specific research questions (Figure 3). Since a major challenge before fully exploiting a given model is to breed it in the laboratory, it would be worthwhile endeavor to run a wider screening among candidate organisms. In this way, a model suitable to answer a given set of questions and able to complete its lifecycle in the laboratory can be identified.
3. *Be aware of the taxonomic bias.* As a corollary of the previous point, it is important to remember that our knowledge of subterranean species is still strongly biased in its taxonomical coverage. Even today, the natural history information on subterranean species

remains largely fragmented, rarely standardized, and often biased toward a few well-studied model organisms and temperate regions. We stress the importance of broadening eco-evolutionary studies to incorporate a larger range of organisms and subterranean habitats, to explore hypotheses about the emergence of convergent traits and behaviors across distant taxa while accounting for phylogenetic effects.

4. *Embrace multidisciplinary.* In light of the habitat and biological impediments, combining ecological and behavioral observations with evolutionary approaches, genetic tools, and simulations are a critical premise. In the -omics era, integrative studies are expected to grow, allowing us to understand which molecular adjustments (including epigenetic effects) occur during the surface-subterranean transitions. This is required, for example, to disentangle the role of standing genetic variation and phenotypic plasticity in driving the evolution of subterranean populations (Bilandžija et al., 2020).

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CONFLICTS OF INTEREST

None declared.

AUTHORS CONTRIBUTION

SM wrote the first draft. AM and SIS provided most arguments on aquatic habitats. EL and HB provided most arguments on vertebrates. AM, SM, and HB developed the section on model organisms. PC, SIS, and VG developed the in silico section. TH provided expert opinion on different sections, especially on behavioral topics. AM and SM prepared figures. All authors contributed to the writing with comments and additions.

DATA AVAILABILITY STATEMENT

This review contains no data. Number of papers and molecular sequences included in Table S1 have been derived from Web of Science and GeneBank (accessed on 5 January 2021).

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REFERENCES

- Alvarenga, D., Silva, M., & Ferreira, R. (2021). Troglорегионы: Delimiting subterranean faunistic identities in Afrotropics and Neotropics. *Nature Portfolio*, <https://doi.org/10.21203/rs.3.rs-53597/v1>
- Arnedo, M. A., Oromí, P., Múrria, C., Macías-Hernández, N., & Ribera, C. (2007). The dark side of an island radiation: Systematics and evolution of troglobitic spiders of the genus *Dysdera* Latreille (Araneae:Dysderidae) in the Canary Islands. *Invertebrate Systematics*, 21(6), 623–660. <https://doi.org/10.1071/IS07015>
- Asmyhr, M. G., & Cooper, S. J. B. (2012). Difficulties barcoding in the dark: The case of crustacean stygofauna from eastern Australia. *Invertebrate Systematics*, 26(6), 583–591. <https://doi.org/10.1071/IS12032>
- Ayllón, D., Railsback, S. F., Almodóvar, A., Nicola, G. G., Vincenzi, S., Elvira, B., & Grimm, V. (2018). Eco-evolutionary responses to recreational fishing under different harvest regulations. *Ecology and Evolution*, 8(19), 9600–9613. <https://doi.org/10.1002/ece3.4270>
- Azmy, S. N., Sah, S. A. M., Shafie, N. J., Ariffin, A., Majid, Z., Ismail, M. N. A., & Shamsir, M. S. (2012). Counting in the dark: Non-intrusive laser scanning for population counting and identifying roosting bats. *Scientific Reports*, 2, 524. <https://doi.org/10.1038/srep00524>
- Baker, G. M. (2015). Quantifying wildlife use of cave entrances using remote camera traps. *Journal of Cave and Karst Studies*, 77, 200–210. <https://doi.org/10.4311/2015ES0101>
- Banitz, T., Johst, K., Wick, L. Y., Schamfuß, S., Harms, H., & Frank, K. (2013). Highways versus pipelines: Contributions of two fungal transport mechanisms to efficient bioremediation. *Environmental Microbiology Reports*, 5(2), 211–218. <https://doi.org/10.1111/1758-2229.12002>
- Barbosa, P., Leal, E. V., da Silva, M., de Almeida, M. C., Moreira-Filho, O., & Artoni, R. F. (2017). Variability and evolutionary implications of repetitive DNA dynamics in genome of *Astyanax scabripinnis* (Teleostei, Characidae). *Comparative Cytogenetics*, 11(1), 143–162. <https://doi.org/10.3897/CompCytogen.v11i1.11149>
- Barton, H. A. (2020). Safe and effective disinfection of show cave infrastructure in a time of COVID-19. *International Journal of Speleology*, 49, 137–147. <https://doi.org/10.5038/1827-806X.49.2.232>
- Benioug, M., Golfier, F., Oltéan, C., Buès, M. A., Bahar, T., & Cuny, J. (2017). An immersed boundary-lattice Boltzmann model for biofilm growth in porous media. *Advances in Water Resources*, 107, 65–82. <https://doi.org/10.1016/j.advwatres.2017.06.009>
- Benioug, M., Golfier, F., Tinet, A.-J., Buès, M. A., & Oltéan, C. (2015). Numerical efficiency assessment of IB-LB method for 3D pore-scale modeling of flow and transport. *Transport in Porous Media*, 109(1), 1–23. <https://doi.org/10.1007/s11242-015-0497-6>
- Berning, D., Adams, H., Luc, H., & Gross, J. B. (2019). In-frame indel mutations in the genome of the blind Mexican cavefish, *Astyanax mexicanus*. *Genome Biology and Evolution*, 11(9), 2563–2573. <https://doi.org/10.1093/gbe/evz180>
- Bierbach, D., Lukas, J., Bergmann, A., Elsner, K., Höhne, L., Weber, C., Weimar, N., Arias-Rodríguez, L., Mönck, H. J., Nguyen, H., Romanczuk, P., Landgraf, T., & Krause, J. (2018). Insights into the social behavior of surface and Cave-Dwelling Fish (*Poecilia mexicana*) in light and darkness through the use of a Biomimetic robot. *Frontiers Robotics AI*, 5, 3. <https://doi.org/10.3389/frobt.2018.00003>
- Bilandžija, H., Hollifield, B., Steck, M., Meng, G., Ng, M., Koch, A. D., Gračan, R., Četković, H., Porter, M. L., Renner, K. J., & Jeffery, W. (2020). Phenotypic plasticity as a mechanism of cave colonization and adaptation. *Elife*, 9, e51830. <https://doi.org/10.7554/eLife.51830>
- Blamires, S. J. (2010). Plasticity in extended phenotypes: Orb web architectural responses to variations in prey parameters. *Journal of Experimental Biology*, 213, 3207–3212. <https://doi.org/10.1242/jeb.045583>
- Blin, M., Fumey, J., Lejeune, C., Policarpo, M., Leclercq, J., Père, S., Torres-Paz, J., Pierre, C., Imarazene, B., & Rétaux, S. (2020). Diversity of olfactory responses and skills in *Astyanax mexicanus* cavefish populations inhabiting different caves. *Diversity*, 12, 395. <https://doi.org/10.3390/d12100395>
- Borer, B., Ataman, M., Hatzimanikatis, V., & Or, D. (2019). Modeling metabolic networks of individual bacterial agents in heterogeneous and dynamic soil habitats (IndiMeSH). *PLoS Computational Biology*, 15(6), e1007127. <https://doi.org/10.1371/journal.pcbi.1007127>
- Boyd, S. H., Niemiller, K. D. K., Dooley, K. E., Nix, J., & Niemiller, M. L. (2020). Using environmental DNA methods to survey for rare groundwater fauna: Detection of an endangered endemic cave crayfish in northern Alabama. *PLoS One*, 15(12), e0242741. <https://doi.org/10.1371/journal.pone.0242741>
- Brancelj, A. (2004). Biological sampling methods for epikarst water. In W. K. Jones, D. C. Culver, & J. S. Herman (Eds.), *Epikarst* (pp. 99–103). Karst Waters Inst. Special Publ.
- Brankovits, D., Pohlman, J. W., Niemann, H., Leigh, M. B., Leewis, M. C., Becker, K. W., Iliffe, T. M., Alvarez, F., Lehmann, M. F., & Phillips, B. (2017). Methane-and dissolved organic carbon-fueled microbial loop supports a tropical subterranean estuary ecosystem. *Nature Communications*, 8(1), 1–12. <https://doi.org/10.1038/s41467-017-01776-x>
- Buzzacott, P. L., Zeigler, E., Denoble, P., & Vann, R. (2009). American cave diving fatalities 1969–2007. *International Journal of Aquatic Research and Education*, 3, 162–177. <https://doi.org/10.25035/ijare.03.02.07>
- Calisher, C. H., Childs, J. E., Field, H. E., Holmes, K. V., & Schountz, T. (2006). Bats: Important reservoir hosts of emerging viruses. *Clinical Microbiology Reviews*, 19(3), 531–545. <https://doi.org/10.1128/CMR.00017-06>
- Camacho, A. I., Mas-Peinado, P., Dorda, B. A., Casado, A., Brancelj, A., Knight, L. R. F. D., Hutchins, B., Bou, C., Perina, G., & Rey, I. (2018). Molecular tools unveil an underestimated diversity in a stygofauna family: A preliminary world phylogeny and an updated morphology of Bathynellidae (Crustacea: Bathynellacea). *Zoological Journal of the Linnean Society*, 183(1), 70–96. <https://doi.org/10.1093/zoolinnean/zlx063>
- Carpenter, J. H. (1999). Behavior and ecology of *Speleonectes epilimnium* (Remipedia, Speleonektidae) from surface water of an anchialine cave on San Salvador Island. *Bahamas Crustaceana*, 72(8), 979–991. <https://doi.org/10.1163/156854099503889>
- Carpenter, J. H. (2021). Forty-year natural history study of *Bahalana geracei* Carpenter, 1981, an anchialine cave-dwelling isopod (Crustacea, Isopoda, Cirolanidae) from San Salvador Island, Bahamas: Reproduction, growth, longevity, and population structure. *Subterranean Biology*, 37, 105–156. <https://doi.org/10.3897/subtbiol.37.60653>
- Cavallari, N., Frigato, E., Vallone, D., Fröhlich, N., Lopez-Olmeda, J. F., Foà, A., Berti, R., Sánchez-Vázquez, F. J., Bertolucci, C., & Foulkes, N. S. (2011). A blind circadian clock in cavefish reveals that opsins mediate peripheral clock photoreception. *PLoS Biology*, 9(9), e1001142. <https://doi.org/10.1371/journal.pbio.1001142>
- Chávez-Solis, E. M., Solís, C., Simões, N., & Mascaró, M. (2020). Distribution patterns, carbon sources and niche partitioning in cave shrimps (Atyidae: Typhlatya). *Scientific Reports*, 10(1), 1–16. <https://doi.org/10.1038/s41598-020-69562-2>
- Cigna, A. A. (2005). Radon in caves. *International Journal of Speleology*, 34, 1–18. <https://doi.org/10.5038/1827-806X.34.1.1>

- Clergue-Gazeau, M. (1974). Reproduction des Urodèles: Perturbation du cycle sexuel des Euproctes males en élevage à la Grotte de Moulis. *Memoires De Biospeologie*, 29, 137–141.
- Culver, D. C., & Pipan, T. (2014). *Shallow subterranean habitats: Ecology, evolution, and conservation*. Oxford University Press. <https://doi.org/10.4311/2014br0127>
- Culver, D. C., & Pipan, T. (2019). *The biology of caves and other subterranean habitats* (2nd ed.). Oxford University Press.
- Culver, D. C., & Sket, B. (2002). Biological monitoring in caves. *Acta Carsologica*, 31, 55–64. <https://doi.org/10.3986/ac.v31i1.403>
- da Silva, A. P. B., Oliveira, I. P. M. R., Bastos-Pereira, R., & Ferreira, R. L. (2018). Are laboratory studies on behavior of troglobitic species always trustful? A case study with an isopod from Brazil. *Behavioural Processes*, 153, 55–65. <https://doi.org/10.1016/j.beproc.2018.05.009>
- DeAngelis, D. L., & Grimm, V. (2014). Individual-based models in ecology after four decades. *F1000Prime Reports*, 6, 39. <https://doi.org/10.12703/P6-39>
- Delić, T., & Sket, B. (2015). Found after 60 years: The hows and whys of *Sphaeromides virei montenigrina* (Crustacea: Isopoda: Cirolanidae) rediscovery in Obodska pečina, Montenegro. *Natura Sloveniae*, 17, 59–65.
- Delić, T., Trontelj, P., Rendoš, M., & Fišer, C. (2017). The importance of naming cryptic species and the conservation of endemic subterranean amphipods. *Scientific Reports*, 7(1), 3391. <https://doi.org/10.1038/s41598-017-02938-z>
- Di Lorenzo, T., Di Marzio, W. D., Fiasca, B., Galassi, D. M. P., Korbel, K., Iepure, S., Pereira, J. L., Reboleira, A. S. P. S., Schmidt, S. I., & Hose, G. C. (2019). Recommendations for ecotoxicity testing with stygobiotic species in the framework of groundwater environmental risk assessment. *Science of the Total Environment*, 681, 292–304. <https://doi.org/10.1016/J.SCITOTENV.2019.05.030>
- Di Lorenzo, T., Di Marzio, W. D., Sáenz, M. E., Baratti, M., Dedonno, A. A., Iannucci, A., Cannicci, S., Messana, G., & Galassi, D. M. P. (2014). Sensitivity of hypogean and epigean freshwater copepods to agricultural pollutants. *Environmental Science and Pollution Research*, 21, 4643–4655. <https://doi.org/10.1007/s11356-013-2390-6>
- Diaz, J. H. (2018). Environmental and wilderness-related risk factors for histoplasmosis: More than bats in caves. *Wilderness & Environmental Medicine*, 29(4), 531–540. <https://doi.org/10.1016/j.wem.2018.06.008>
- DiStefano, R. J., Ashley, D., Brewer, S. K., Mouser, J. B., & Niemiller, M. (2020). Preliminary investigation of the critically imperiled Caney Mountain cave crayfish *Orconectes stygocaneyi* (Hobbs III, 2001) (Decapoda: Cambaridae) in Missouri, USA. *Freshwater Crayfish*, 25, 47–57. <https://doi.org/10.5869/fc.2020.v25-1.047>
- Dole-olivier, M.-J., Castellari, F., Coineau, N., Galassi, D. M. P., Martin, P., Mori, N., Valdecasas, A., & Gibert, J. (2009). Towards an optimal sampling strategy to assess groundwater biodiversity: Comparison across six European regions. *Freshwater Biology*, 54, 777–796. <https://doi.org/10.1111/j.1365-2427.2008.02133.x>
- Du Preez, G., Majdi, N., Swart, A., Traunspurger, W., & Fourie, H. (2017). Nematodes in caves: A historical perspective on their occurrence, distribution and ecological relevance. *Nematology*, 19, 627–644. <https://doi.org/10.1163/15685411-00003068>
- Durand, P. J. (1970). Fortpflanzung und Entwicklung von Hydromantes, dem Höhlenmolch. *Aqua Terra*, 7, 42–48.
- Elliott, R. W., Kaufmann, J. E., Samoray, S. T., & Gardner, S. E. (2005). The MDC method: Counting bats with infrared video. In G. T. Rea (Ed.), *Proceedings of the 2005 National Cave and Karst management symposium* (pp. 147–153). The NCKMS Steering Committee.
- Eme, D., Zagmajster, M., Delić, T., Fišer, C., Flot, J.-F., Konecny-Dupré, L., Pálsson, S., Stoch, F., Zakšek, V., Douady, C. J., & Malard, F. (2018). Do cryptic species matter in macroecology? Sequencing European groundwater crustaceans yields smaller ranges but does not challenge biodiversity determinants. *Ecography*, 41(2), 424–436. <https://doi.org/10.1111/ecog.02683>
- Esposito, L. A., Bloom, T., Caicedo-Quiroga, L., Alicea-Serrano, A. M., Sánchez-Ruiz, J. A., May-Collado, L. J., Binford, G. J., & Agnarsson, I. (2015). Islands within islands: Diversification of tailless whip spiders (Amblypygi, Phrynus) in Caribbean caves. *Molecular Phylogenetics and Evolution*, 93, 107–117. <https://doi.org/10.1016/J.YMPEV.2015.07.005>
- Exley, S. (1983). Lanzarote volcanic cave expedition 1983. *Explorers Journal*, 118–123.
- Ferreira, R. L., & Martins, R. P. (1999). Trophic structure and natural history of bat guano invertebrate communities, with special reference to Brazilian caves. *Tropical Zoology*, 12, 231–252. <https://doi.org/10.1080/03946975.1999.10539391>
- Ficetola, G. F., Canedoli, C., & Stoch, F. (2019). The Racovitza impediment and the hidden biodiversity of unexplored environments. *Conservation Biology*, 33(1), 214–216. <https://doi.org/10.1111/cobi.13179>
- Fišer, C. (2009). Niphargus—A model system for evolution and ecology. In D. C. Culver & W. B. White (Eds.), *Encyclopedia of caves* (pp. 746–755). Academic Press.
- Fišer, C., Pipan, T., & Culver, D. C. (2014). The vertical extent of groundwater metazoans: An ecological and evolutionary perspective. *BioScience*, 64(11), 971–979. <https://doi.org/10.1093/biosci/biu148>
- Fišer, C., Robinson, C. T., & Malard, F. (2018). Cryptic species as a window into the paradigm shift of the species concept. *Molecular Ecology*, 27(3), 613–635. <https://doi.org/10.1111/mec.14486>
- Fraser, B. G., & Williams, D. D. (1997). Accuracy and precision in sampling hyporheic fauna. *Canadian Journal of Fisheries and Aquatic Sciences*, 54(5), 1135–1141. <https://doi.org/10.1139/f97-024>
- Friedrich, M. (2013). Biological clocks and visual systems in cave-adapted animals at the dawn of speleogenomics. *Integrative and Comparative Biology*, 53(1), 50–67. <https://doi.org/10.1093/icb/ict058>
- Galli, L., Janžekovič, F., Kozel, P., & Novak, T. (2021). Protura (Arthropoda: Hexapoda) in Slovenian caves. *International Journal of Speleology*, 50, 65–74. <https://doi.org/10.5038/1827-806X.50.1.2380>
- Gillmore, G. K., Sperrin, M., Phillips, P., & Denman, A. (2000). Radon hazards, geology, and exposure of cave users: A case study and some theoretical perspectives. *Ecotoxicology and Environmental Safety*, 46(3), 279–288. <https://doi.org/10.1006/eesa.2000.1922>
- Godefroy, N., Le Goff, E., Martinand-Mari, C., Belkhir, K., Vacelet, J., & Baghdiguan, S. (2019). Sponge digestive system diversity and evolution: Filter feeding to carnivory. *Cell and Tissue Research*, 377(3), 341–351. <https://doi.org/10.1007/s00441-019-03032-8>
- Goldstein, B., & King, N. (2016). The future of cell biology: Emerging model organisms. *Trends in Cell Biology*, 26(11), 818–824. <https://doi.org/10.1016/j.tcb.2016.08.005>
- Gonzalez, B. C., Martínez, A., Borda, E., Iliffe, T. M., Fontaneto, D., & Worsaae, K. (2017). Genetic spatial structure of an anchialine cave annelid indicates connectivity within - But not between - Islands of the Great Bahama Bank. *Molecular Phylogenetics and Evolution*, 109, 259–270. <https://doi.org/10.1016/j.ympev.2017.01.003>
- Gorički, Š., Stanković, D., Snoj, A., Kuntner, M., Jeffery, W. R., Trontelj, P., Pavičević, M., Grizelj, Z., Năpăruș-Aljančič, M., & Aljančič, G. (2017). Environmental DNA in subterranean biology: Range extension and taxonomic implications for Proteus. *Scientific Reports*, 7, 91–93. <https://doi.org/10.1038/srep45054>
- Gravish, N., & Lauder, G. V. (2018). Robotics-inspired biology. *Journal of Experimental Biology*, 221, jeb138438. <https://doi.org/10.1242/jeb.138438>
- Grimm, V., & Berger, U. (2016). Structural realism, emergence, and predictions in next-generation ecological modelling: Synthesis from a special issue. *Ecological Modelling*, 326, 177–187. <https://doi.org/10.1016/j.ecolmodel.2016.01.001>

- Grimm, V., Revilla, E., Berger, U., Jeltsch, F., Mooij, W. M., Railsback, S. F., DeAngelis, D. L. (2005). Pattern-oriented modeling of agent-based complex systems: Lessons from ecology. *Science*, 310(5750), 987–991. <https://doi.org/10.1126/science.1116681>
- Hancock, P. J., & Boulton, A. J. (2009). Sampling groundwater fauna: Efficiency of rapid assessment methods tested in bores in eastern Australia. *Freshwater Biology*, 54, 902–914. <https://doi.org/10.1111/j.1365-2427.2007.01878.x>
- Hartig, F., Calabrese, J. M., Reineking, B., Wiegand, T., & Huth, A. (2011). Statistical inference for stochastic simulation models – Theory and application. *Ecology Letters*, 14(8), 816–827. <https://doi.org/10.1111/j.1461-0248.2011.01640.x>
- Hedges, S. B. (2002). The origin and evolution of model organisms. *Nature Reviews Genetics*, 3(11), 838–849. <https://doi.org/10.1038/nrg929>
- Hesselberg, T. (2010). Ontogenetic changes in web design in two orb-web spiders. *Ethology*, 116, 535–545. <https://doi.org/10.1111/j.1439-0310.2010.01760.x>
- Hesselberg, T. (2015). Exploration behaviour and behavioural flexibility in orb-web spiders: A review. *Current Zoology*, 61(2), 313–327. <https://doi.org/10.1093/czoolo/61.2.313>
- Hesselberg, T., Simonsen, D., & Juan, C. (2019). Do cave orb spiders show unique behavioural adaptations to subterranean life? A review of the evidence. *Behaviour*, 156(10), 969–996. <https://doi.org/10.1163/1568539X-00003564>
- Hortal, J., de Bello, F., Diniz-Filho, J. A. F., Lewinsohn, T. M., Lobo, J. M., & Ladle, R. J. (2015). Seven shortfalls that beset large-scale knowledge of biodiversity. *Annual Review of Ecology, Evolution, and Systematics*, 46, 523–549. <https://doi.org/10.1146/annurev-ecolsys-112414-054400>
- Howarth, F. G. (1983). Ecology of cave arthropods. *Annual Review of Entomology*, 28(1), 365–389. <https://doi.org/10.1146/annurev.en.28.010183.002053>
- Humphreys, W. F., Poole, A., Eberhard, S. M., & Warren, D. (1999). Effects of research diving on the physico-chemical profile of Bundera Sinkhole, an anchialine remiped habitat at Cape Range, Western Australia. *Journal of the Royal Society of Western Australia*, 82, 99–108.
- Hunt, P. J., Harden, T. J., Hibbins, M., Pritchard, R. C., Muir, D. B., & Gardner, F. J. (1984). Histoplasma capsulatum. Isolation from an Australian cave environment and from a patient. *The Medical Journal of Australia*, 141(5), 280–283.
- Iliffe, T. M. (2018). Collecting and processing crustaceans from anchialine and marine caves. *Journal of Crustacean Biology*, 38(3), 374–379. <https://doi.org/10.1093/jcabi/ruy011>
- Iliffe, T. M., & Bowen, C. (2001). Scientific cave diving. *Marine Technology Society Journal*, 35(2), 36–41. <https://doi.org/10.4031/002533201788001901>
- Jaggard, J. B., Stahl, B. A., Lloyd, E., Prober, D. A., Duboue, E. R., & Keene, A. C. (2018). Hypocretin underlies the evolution of sleep loss in the Mexican cavefish. *Elife*, 7, e32637. <https://doi.org/10.7554/eLife.32637>
- Jeffery, W. R. (2020). Astyanax surface and cave fish morphs. *EvoDevo*, 11(1), 14. <https://doi.org/10.1186/s13227-020-00159-6>
- Juan, C., Guzik, M. T., Jaume, D., & Cooper, S. J. B. (2010). Evolution in caves: Darwin's 'wrecks of ancient life' in the molecular era. *Molecular Ecology*, 19(18), 3865–3880. <https://doi.org/10.1111/j.1365-294X.2010.04759.x>
- Juberthie, C. (1985). Cycle vital de Telema tenella dans la Grotte-Laboratoire de Moulis et strategies de reproduction chez les Araignees cavernicoles. *Memoires De Biospeologie*, 12, 77–89.
- Juberthie, C., Durand, J., & Dupuy, M. (1996). La reproduction des Protées (Proteus anguinus): Bilan de 35 ans d'élevage dans les grottes-laboratoires de Moulis et d'Aulignac. *Memoires De Biospeologie*, 23, 53–56.
- Jurado-Rivera, J. A., Pons, J., Alvarez, F., Botello, A., Humphreys, W. F., Page, T. J., Iliffe, T. M., Willassen, E., Meland, K., Juan, C., & Jaume, D. (2017). Phylogenetic evidence that both ancient vicariance and dispersal have contributed to the biogeographic patterns of anchialine cave shrimps. *Scientific Reports*, 7(1), 2852. <https://doi.org/10.1038/s41598-017-03107-y>
- Keene, A. C., Yoshizawa, M., & McGaugh, S. E. (2016). *Biology and evolution of the Mexican cavefish*. Academic Press (Elsevier).
- Keith, D. A., Ferrer-Paris, J. R., Nicholson, E., & Kingsford, R. T. (2020). *The IUCN Global Ecosystem Typology 2.0: Descriptive profiles for biomes and ecosystem functional groups*. IUCN. <https://doi.org/10.2305/IUCN.CH.2020.13.en>
- Kim, M., & Or, D. (2016). Individual-based model of microbial life on hydrated rough soil surfaces. *PLoS One*, 11(1), e0147394. <https://doi.org/10.1371/journal.pone.0147394>
- Koenemann, S., Schram, F. R., Iliffe, T. M., Hinderstein, L. M., & Bloechl, A. (2007). Behavior of remipedia in the laboratory, with supporting field observations. *Journal of Crustacean Biology*, 27(4), 534–540. <https://doi.org/10.1651/S-2809A.1>
- Kolicka, M., Gadawski, P., & Dabert, M. (2017). A new species of freshwater Chaetonotidae (Gastrotricha, Chaetonotida) from Obodska Cave (Montenegro) based on morphological and molecular characters. *European Journal of Taxonomy*, 354, 1–30.
- Kuzmin, I. V., Bozick, B., Guagliardo, S. A., Kunkel, R., Shak, J. R., Tong, S., & Rupprecht, C. E. (2011). Bats, emerging infectious diseases, and the rabies paradigm revisited. *Emerging Health Threats Journal*, 4(1), 7159. <https://doi.org/10.3402/ehth.v4i0.7159>
- Kuzmin, I. V., Niezgoda, M., Franka, R., Agwanda, B., Markotter, W., Breiman, R. F., Shieh, W.-J., Zaki, S. R., & Rupprecht, C. E. (2010). Marburg virus in fruit bat, Kenya. *Emerging Infectious Disease Journal*, 16(2), 352. <https://doi.org/10.3201/eid1602.091269>
- Lenau, T. A., Metze, A.-L., & Hesselberg, T. (2018). Paradigms for biologically inspired design. In *Paradigms for biologically inspired design* (p. 1059302). Proceedings Volume 10593, Bioinspiration, Biomimetics, and Bioreplication VIII. <https://doi.org/10.1117/12.2296560>
- Lepore, E., Marchioro, A., Isaia, M., Buehler, M. J., & Pugno, N. M. (2012). Evidence of the most stretchable egg sac silk stalk, of the European spider of the year meta menardi. *PLoS One*, 7(2), e30500. <https://doi.org/10.1371/journal.pone.0030500>
- Lozano-Fernandez, J., Carton, R., Tanner, A. R., Puttick, M. N., Blaxter, M., Vinther, J., Olesen, J., Giribet, G., Edgecombe, G. D., & Pisani, D. (2016). A molecular palaeobiological exploration of arthropod terrestrialization. *Philosophical Transactions of the Royal Society B: Biological Sciences*, 371(1699), 20150133. <https://doi.org/10.1098/rstb.2015.0133>
- Lunghi, E. (2018). Ecology and life history of Meta bourneti (Araneae: Tetragnathidae) from Monte Albo (Sardinia, Italy). *PeerJ*, 6, e6049. <https://doi.org/10.7717/peerj.6049>
- Lunghi, E., Cianferoni, F., Ceccolini, F., Veith, M., Manenti, R., Mancinelli, G., Corti, C., & Ficetola, G. F. (2018). What shapes the trophic niche of European plethodontid salamanders? *PLoS One*, 13, e0205672. <https://doi.org/10.1371/journal.pone.0205672>
- Lunghi, E., Corti, C., Manenti, R., Barzaghi, B., Buschetti, S., Canedoli, C., & Ficetola, G. F. (2018). Comparative reproductive biology of European cave salamanders (genus Hydromantes): Nesting selection and multiple annual breeding. *Salamandra*, 54, 101–108.
- Lunghi, E., Corti, C., Mulargia, M., Zhao, Y., Manenti, R., Ficetola, G. F., & Veith, M. (2020). Cave morphology, microclimate and abundance of five cave predators from the Monte Albo (Sardinia, Italy). *Biodiversity Data Journal*, 8, e48623. <https://doi.org/10.3897/BDJ.8.e48623>
- Lunghi, E., Ficetola, G. F., Mulargia, M., Cogoni, R., Veith, M., Corti, C., & Manenti, R. (2018). Batracobdella leeches, environmental features and Hydromantes salamanders. *International Journal of Parasitology*:

- Parasites and Wildlife*, 7, 48–53. <https://doi.org/10.1016/j.jppaw.2018.01.003>
- Lunghi, E., Manenti, R., Canciani, G., Scari, G., Pennati, R., & Ficetola, G. F. (2016). Thermal equilibrium and temperature differences among body regions in European plethodontid salamanders. *Journal of Thermal Biology*, 60, 79–85. <https://doi.org/10.1016/j.jtherbio.2016.06.010>
- Lunghi, E., Manenti, R., Cianferoni, F., Ceccolini, F., Veith, M., Corti, C., Ficetola, G. F., & Mancinelli, G. (2020). Interspecific and interpopulation variation in individual diet specialization: Do environmental factors have a role? *Ecology*, 101(8), e03088. <https://doi.org/10.1002/ecy.3088>
- Lunghi, E., Manenti, R., Manca, S., Mulargia, M., Pennati, R., & Ficetola, G. F. (2014). Nesting of cave salamanders (*Hydromantes flavus* and *H. italicus*) in natural environments. *Salamandra*, 50, 105–109.
- Lunghi, E., Murgia, R., De Falco, G., Buschetti, S., Mulas, C., Mulargia, M., & Ficetola, G. F. (2015). First data on nesting ecology and behaviour in the imperial cave salamander *Hydromantes imperialis*. *North-Western Journal of Zoology*, 11, 324–330.
- MacFarlane, D., & Rocha, R. (2020). Guidelines for communicating about bats to prevent persecution in the time of COVID-19. *Biological Conservation*, 248, 108650. <https://doi.org/10.1016/j.bioccon.2020.108650>
- MacNeil, R. R., & Brcic, J. (2017). Coping with the subterranean environment: A thematic content analysis of the narratives of cave explorers. *Journal of Human Performance in Extreme Environments*, 13(1), 6. <https://doi.org/10.7771/2327-2937.1089>
- Maher, B. (2009). Evolution: Biology's next top model? *Nature*, 458, 695–699. <https://doi.org/10.1038/458695a>
- Malard, F., Grison, P., Duchemin, L., Konecny-Dupré, L., Lefébure, T., Saclier, N., Eme, D., Martin, C., Callou, C., & Douady, C. J. (2020). GOTT: A laboratory application software for optimizing multi-criteria species-based research. *Methods in Ecology and Evolution*, 11(1), 159–167. <https://doi.org/10.1111/2041-210X.13307>
- Mammola, S. (2019). Finding answers in the dark: Caves as models in ecology fifty years after Poulson and White. *Ecography*, 42(7), 1331–1351. <https://doi.org/10.1111/ecog.03905>
- Mammola, S. (2020). On deepest caves, extreme habitats, and ecological superlatives. *Trends in Ecology & Evolution*, 35(6), 469–472. <https://doi.org/10.1016/j.tree.2020.02.011>
- Mammola, S., Amorim, I. R., Bichuette, M. E., Borges, P. A. V., Cheeptham, N., Cooper, S. J. B., Culver, D. C., Deharveng, L., Eme, D., Ferreira, R. L., Fišer, C., Fišer, Ž., Fong, D. W., Griebler, C., Jeffery, W. R., Jugovic, J., Kowalko, J. E., Lilley, T. M., Malard, F., ... Cardoso, P. (2020). Fundamental research questions in subterranean biology. *Biological Reviews*, 95(6), 1855–1872. <https://doi.org/10.1111/brv.12642>
- Mammola, S., Cardoso, P., Culver, D. C., Deharveng, L., Ferreira, R. L., Fišer, C., Galassi, D. M. P., Griebler, C., Halse, S., Humphreys, W. F., Isaia, M., Malard, F., Martinez, A., Moldovan, O. T., Niemiller, M. L., Pavlek, M., Reboleira, A. S. P. S., Souza-Silva, M., Teeling, E. C., ... Zagmajster, M. (2019). Scientists' warning on the conservation of subterranean ecosystems. *BioScience*, 69(8), 641–650. <https://doi.org/10.1093/biosci/biz064>
- Mammola, S., Giachino, P. M., Piano, E., Jones, A., Barberis, M., Badino, G., & Isaia, M. (2016). Ecology and sampling techniques of an understudied subterranean habitat: The Milieu Souterrain Superficiel (MSS). *The Science of Nature*, 103(11–12), 88. <https://doi.org/10.1007/s00114-016-1413-9>
- Mammola, S., & Isaia, M. (2017). Spiders in caves. *Proceedings of the Royal Society B: Biological Sciences*, 284(1853), 20170193. <https://doi.org/10.1098/rspb.2017.0193>
- Mammola, S., & Isaia, M. (2018). Day-night and seasonal variations of a subterranean invertebrate community in the twilight zone. *Subterranean Biology*, 27, 31–51. <https://doi.org/10.3897/SUBTB IOL.27.28909>
- Mammola, S., & Leroy, B. (2018). Applying species distribution models to caves and other subterranean habitats. *Ecography*, 41(7), 1194–1208. <https://doi.org/10.1111/ecog.03464>
- Mammola, S., Piano, E., Malard, F., Vernon, P., & Isaia, M. (2019). Extending Janzen's hypothesis to temperate regions: A test using subterranean ecosystems. *Functional Ecology*, 33(9), 1638–1650. <https://doi.org/10.1111/1365-2435.13382>
- Mammola, S., Souza, M. F. V. R., Isaia, M., & Ferreira, R. L. (2021). Global distribution of microwhip scorpions (Arachnida: Palpigradi). *Journal of Biogeography*, 00, 1–13. <https://doi.org/10.1111/jbi.14094>
- Manenti, R., Barzaghi, B., Lana, E., Stocchino, G. A., Manconi, R., & Lunghi, E. (2018). The stenoendemic cave-dwelling planarians (Platyhelminthes, Tricladida) of the Italian Alps and Apennines: Conservation issues. *Journal for Nature Conservation*, 45, 90–97. <https://doi.org/10.1016/j.jnc.2018.08.001>
- Manenti, R., Melotto, A., Guillaume, O., Ficetola, G. F., & Lunghi, E. (2020). Switching from mesopredator to apex predator: How do responses vary in amphibians adapted to cave living? *Behavioral Ecology and Sociobiology*, 74, 126. <https://doi.org/10.1007/s00265-020-02909-x>
- Martínez, A., Di Domenico, M., Leasi, F., Curini-Galletti, M., Todaro, M. A., Zotto, M. D., Gobert, S., Artois, T., Norenburg, J., Jörger, K. M., Núñez, J., Fontaneto, D., & Worsaae, K. (2019). Patterns of diversity and endemism of soft-bodied meiofauna in an oceanic island, Lanzarote, Canary Islands. *Marine Biodiversity*, 49, 2033–2055. <https://doi.org/10.1007/s12526-019-01007-0>
- Martínez, A., Di Domenico, M., & Worsaae, K. (2013). Evolution of cave Axiobuina and Speleobregma (Scalibregmatidae, Annelida). *Zoologica Scripta*, 42(6), 623–636. <https://doi.org/10.1111/zsc.12024>
- Martínez, A., Kvindebjerg, K., Iliffe, T. M., & Worsaae, K. (2017). Evolution of cave suspension feeding in Protodrilidae (Annelida). *Zoologica Scripta*, 46(2), 214–226. <https://doi.org/10.1111/zsc.12198>
- Martínez, A., & Mammola, S. (2020). Let research on subterranean habitats resonate!. *Subterranean Biology*, 36, 63–71. <https://doi.org/10.3897/subtbiol.36.59960>
- McGaugh, S. E., Kowalko, J. E., Duboué, E., Lewis, P., Franz-Odenaal, T. A., Rohner, N., Gross, J. B., & Keene, A. C. (2020). Dark world rises: The emergence of cavefish as a model for the study of evolution, development, behavior, and disease. *Journal of Experimental Zoology Part B: Molecular and Developmental Evolution*, 334(7–8), 397–404. <https://doi.org/10.1002/jez.b.22978>
- Mermillod-Blondin, F., Lefour, C., Lalouette, L., Renault, D., Malard, F., Simon, L., & Douady, C. J. (2013). Thermal tolerance breadths among groundwater crustaceans living in a thermally constant environment. *The Journal of Experimental Biology*, 216(9), 1683–1694. <https://doi.org/10.1242/jeb.081232>
- Moldovan, O. T., Mihevc, A., Miko, L., Constantin, S., Meleg, I. N., Petculescu, A., & Bosák, P. (2011). Invertebrate fossils from cave sediments: A new proxy for pre-Quaternary paleoenvironments. *Biogeosciences*, 8(7), 1825–1837. <https://doi.org/10.5194/bg-8-1825-2011>
- Moseley, M. (2009). Size matters: Scalar phenomena and a proposal for an ecological definition of 'cave'. *Cave and Karst Science*, 35, 89–94.
- Möst, M. H., Donabauer, M., Arthofer, W., Schlick-Steiner, B. C., & Steiner, F. M. (2020). Towards an evolutionary history of European-Alpine Trechus ground beetles: Species groups and wing reduction. *Molecular Phylogenetics and Evolution*, 149, 106822. <https://doi.org/10.1016/j.ympev.2020.106822>
- Müller, B., & Grossniklaus, U. (2010). Model organisms – A historical perspective. *Journal of Proteomics*, 73(11), 2054–2063. <https://doi.org/10.1016/j.jprot.2010.08.002>
- Niemiller, M. L., Near, T. J., & Fitzpatrick, B. M. (2012). Delimiting species using multilocus data: Diagnosing cryptic diversity in the southern cavefish, *Typhlichthys subterraneus* (Teleostei: Amblyopsidae). *Evolution*, 66(3), 846–866. <https://doi.org/10.1111/j.1558-5646.2011.01480.x>

- Niemiller, M. L., Porter, M. L., Keany, J., Gilbert, H., Fong, D. W., Culver, D. C., Hobson, C. S., Kendall, K. D., Davis, M. A., & Taylor, S. J. (2018). Evaluation of eDNA for groundwater invertebrate detection and monitoring: A case study with endangered *Stygobromus* (Amphipoda: Crangonyctidae). *Conservation Genetics Resources*, 10, 247–257. <https://doi.org/10.1007/s12686-017-0785-2>
- Northup, D. E., Melim, L. A., Spilde, M. N., Hathaway, J., Garcia, M. G., Moya, M., Stone, F. D., Boston, P. J., Dapkevicius, M., & Riquelme, C. (2011). Lava cave microbial communities within mats and secondary mineral deposits: Implications for life detection on other planets. *Astrobiology*, 11, 601–618. <https://doi.org/10.1089/ast.2010.0562>
- Oliveira, M. P. A. D., Bastos-Pereira, R., Torres, S. H. S., Pereira, T. S., Batista, F. M., Alves, J. P., Iniesta, L. F. M., Bouzan, R. S., Chagas-Jr, A., Prous, X., Pietrobon, T., & Ferreira, R. L. (2019). Choosing sampling methods for Chilopoda, Diplopoda and Isopoda (Oniscidea): A case study for ferruginous landscapes in Brazilian Amazonia. *Applied Soil Ecology*, 143, 181–191. <https://doi.org/10.1016/j.apsoil.2019.07.012>
- Oneto, F., Ottonello, D., Pastorino, M. V., & Salvidio, S. (2010). Posthatching parental care in salamanders revealed by infrared video surveillance. *Journal of Herpetology*, 44, 649–653. <https://doi.org/10.1670/09-181.1>
- Pallarés, S., Colado, R., Botella-Cruz, M., Montes, A., Balart-García, P., Bilton, D. T., Millán, A., Ribera, I., & Sánchez-Fernández, D. (2020). Loss of heat acclimation capacity could leave subterranean specialists highly sensitive to climate change. *Animal Conservation*, Epub ahead of print. <https://doi.org/10.1111/acv.12654>
- Pallarés, S., Sanchez-Hernandez, J. C., Colado, R., Balart-García, P., Comas, J., & Sánchez-Fernández, D. (2020). Beyond survival experiments: Using biomarkers of oxidative stress and neurotoxicity to assess vulnerability of subterranean fauna to climate change. *Conservation Physiology*, 8(1), <https://doi.org/10.1093/conphys/coaa067>
- Parzefall, J. (1982). Changement of behaviour during the evolution of cave animals. *Memoires De Biospeologie*, 8, 55–62.
- Pérez-Moreno, J. L., Iliffe, T. M., & Bracken-Grissom, H. D. (2016). Life in the Underworld: Anchialine cave biology in the era of speleogenomics. *International Journal of Speleology*, 49, 149–170. <https://doi.org/10.5038/1827-806X.45.2.1954>
- Pfeifer, R., Lungarella, M., & Iida, F. (2007). Self-organization, embodiment, and biologically inspired robotics. *Science*, 318, 1088–1093. <https://doi.org/10.1126/science.1145803>
- Polak, S. (1997). A classification of the subterranean environment and cave fauna. *Acta Carsologica*, 26(2), 351–359.
- Poulson, T. L., & White, W. B. (1969). The cave environment. *Science*, 165(3897), 971–981. <https://doi.org/10.1126/science.165.3897.971>
- Protas, M., & Jeffery, W. R. (2012). Evolution and development in cave animals: From fish to crustaceans. *Wiley Interdisciplinary Reviews. Developmental Biology*, 1(6), 823–845. <https://doi.org/10.1002/wdev.61>
- Rastorgueff, P. A., Rocher, C., Selva, M., & Chevaldonné, P. (2015). Preliminary DNA-based diet assessment of a gutless carnivore, the sponge *Asbestopluma hypogea*. *Journal of Experimental Marine Biology and Ecology*, 467, 108–114. <https://doi.org/10.1016/j.jembe.2015.02.014>
- Riddle, M. R., Aspiras, A. C., Gaudenz, K., Peuß, R., Sung, J. Y., Martineau, B., Peavey, M., Box, A. C., Tabin, J. A., McGaugh, S., Borowsky, R., Tabin, C. J., & Rohner, N. (2018). Insulin resistance in cavefish as an adaptation to a nutrient-limited environment. *Nature*, 555, 647. <https://doi.org/10.1038/nature26136>
- Rocha, R., Aziz, S. A., Brook, C. E., Carvalho, W. D., Cooper-Bohannon, R., Frick, W. F., Huang, J.-C.-C., Kingston, T., López-Baucells, A., Maas, B., Mathews, F., Medellin, R. A., Olival, K. J., Peel, A. J., Plowright, R. K., Razgour, O., Rebelo, H., Rodrigues, L., Rossiter, S. J., ... Webala, P. W. (2020). Bat conservation and zoonotic disease risk: A research agenda to prevent misguided persecution in the aftermath of COVID-19. *Animal Conservation*, Epub ahead of print. <https://doi.org/10.1111/acv.12636>
- Ross, P. A., Endersby-Harshman, N. M., & Hoffmann, A. A. (2019). A comprehensive assessment of inbreeding and laboratory adaptation in *Aedes aegypti* mosquitoes. *Evolutionary Applications*, 12(3), 572–586. <https://doi.org/10.1111/eva.12740>
- Russell, J. J., Theriot, J. A., Sood, P., Marshall, W. F., Landweber, L. F., Fritz-Laylin, L., Polka, J. K., Oliferenko, S., Gerbich, T., Gladfelter, A., Umen, J., Bezanilla, M., Lancaster, M. A., He, S., Gibson, M. C., Goldstein, B., Tanaka, E. M., Hu, C.-K., & Brunet, A. (2017). Non-model model organisms. *BMC Biology*, 15(1), 1–31. <https://doi.org/10.1186/s12915-017-0391-5>
- Saccò, M., Blyth, A., Bateman, P. W., Hua, Q., Mazumder, D., White, N., Humphreys, W. F., Laini, A., Griebler, C., & Grice, K. (2019). New light in the dark – A proposed multidisciplinary framework for studying functional ecology of groundwater fauna. *Science of the Total Environment*, 662, 963–977. <https://doi.org/10.1016/j.scitotenv.2019.01.296>
- Saccò, M., Blyth, A. J., Humphreys, W. F., Karasiewicz, S., Meredith, K. T., Laini, A., Cooper, S. J. B., Bateman, P. W., & Grice, K. (2020). Stygofaunal community trends along varied rainfall conditions: Deciphering ecological niche dynamics of a shallow calcrete in Western Australia. *Ecohydrology*, 13(1), e2150. <https://doi.org/10.1002/eco.2150>
- Sánchez, N., & Martínez, A. (2019). Dungeons and dragons: Two new species and records of Kinorhyncha from anchialine cenotes and marine lava tubes. *Zoologischer Anzeiger*, 282, 161–175. <https://doi.org/10.1016/j.jcz.2019.05.012>
- Schmidt, S. I., Hahn, H. J., Watson, G. D., Woodbury, R. J., & Hatton, T. J. (2004). Sampling fauna in stream sediments as well as groundwater using one net sampler. *Acta Hydrochimica Et Hydrobiologica*, 32, 131–137. <https://doi.org/10.1002/aheh.200300522>
- Schmidt, S. I., Kreft, J.-U., Mackay, R., Picioreanu, C., & Thullner, M. (2018). Elucidating the impact of micro-scale heterogeneous bacterial distribution on biodegradation. *Advances in Water Resources*, 116, 67–76. <https://doi.org/10.1016/j.advwatres.2018.01.013>
- Schmidt-Rhaesa, A. (2020). *Guide to the identification of marine meiofauna*. Verlag Dr Friedrich Pfeil.
- Sendra, A., Antić, D., Barranco, P., Borko, Š., Christian, E., Delić, T., & Reboleira, A. S. P. (2020). Flourishing in subterranean ecosystems: Euro-mediterranean plusiocampinae and tachycampoids (Diplura, Campodeidae). *European Journal of Taxonomy*, 591, 1–138.
- Sendra, A., Palero, F., Jiménez-Valverde, A., & Reboleira, A. S. P. S. (2020). Diplura in caves: Diversity, ecology, evolution and biogeography. *Zoological Journal of the Linnean Society*, 20, 1–15. <https://doi.org/10.1093/zoolinnean/zlaa116>
- Smithers, P. (2005). The early life history and dispersal of the cave spider *Meta menardi* (Latreille, 1804) (Araneae: Tetragnathidae). *Bulletin of the British Arachnological Society*, 13(6), 213–216. <https://doi.org/10.1636/CT-05-2.1>
- Staffolani, S., Buonfrate, D., Angheben, A., Gobbi, F., Giorli, G., Guerriero, M., Bisoffi, Z., & Barchiesi, F. (2018). Acute histoplasmosis in immunocompetent travelers: A systematic review of literature. *BMC Infectious Diseases*, 18(1), 673. <https://doi.org/10.1186/s12879-018-3476-z>
- Stemme, T., Iliffe, T. M., von Reumont, B. M., Koenemann, S., Harzsch, S., & Bicker, G. (2013). Serotonin-immunoreactive neurons in the ventral nerve cord of Remipedia (Crustacea): Support for a sister group relationship of Remipedia and Hexapoda? *BMC Evolutionary Biology*, 13(1), 119. <https://doi.org/10.1186/1471-2148-13-119>
- Stockdale, W. T., Lemieux, M. E., Killen, A. C., Zhao, J., Hu, Z., Riepsaame, J., Hamilton, N., Kudoh, T., Riley, P. R., van Aerle, R., Yamamoto, Y., & Mommersteeg, M. T. M. (2018). Heart regeneration in the Mexican cavefish. *Cell Reports*, 25(8), 1997–2007.e7. <https://doi.org/10.1016/j.celrep.2018.10.072>

- Sullivan, W. (2015). The institute for the study of non-model organisms and other fantasies. *Molecular Biology of the Cell*, 26(3), 387–389. <https://doi.org/10.1091/mbc.E14-03-0814>
- Swann, D. E., Hass, C. C., Dalton, D. C., & Wolf, S. A. (2004). Infrared-triggered cameras for detecting wildlife: An evaluation and review. *Wildlife Society Bulletin*, 32, 357–365. [https://doi.org/10.2193/0091-7648\(2004\)32\[357:icfdwa\]2.0.co;2](https://doi.org/10.2193/0091-7648(2004)32[357:icfdwa]2.0.co;2)
- Tang, Y., Valocchi, A. J., Werth, C. J., & Liu, H. (2013). An improved pore-scale biofilm model and comparison with a microfluidic flow cell experiment. *Water Resources Research*, 49(12), 8370–8382. <https://doi.org/10.1002/2013WR013843>
- Torres-Paz, J., Hyacinthe, C., Pierre, C., & Rétaux, S. (2018). Towards an integrated approach to understand Mexican cavefish evolution. *Biology Letters*, 14(8), 20180101. <https://doi.org/10.1098/rsbl.2018.0101>
- Trajano, E., & Bichuette, M. E. (2010). Diversity of Brazilian subterranean invertebrates, with a list of troglomorphic taxa. *Subterranean Biology*, 7, 1–16.
- Trajano, E., Gallão, J. E., & Bichuette, M. E. (2016). Spots of high diversity of troglobites in Brazil: The challenge of measuring subterranean diversity. *Biodiversity and Conservation*, 25(10), 1805–1828. <https://doi.org/10.1007/s10531-016-1151-5>
- Uéno, S.-I. (1987). The derivation of terrestrial cave animals. *Zoological Science*, 4, 593–606.
- Verovnik, R., Sket, B., & Trontelj, P. (2004). Phylogeography of subterranean and surface populations of water lice *Asellus aquaticus* (Crustacea: Isopoda). *Molecular Ecology*, 13, 1519–1532. <https://doi.org/10.1111/j.1365-294X.2004.02171.x>
- Vincent, J. F. V., Bogatyreva, O. A., Bogatyrev, N. R., Bowyer, A., & Pahl, A.-K. (2006). Biomimetics: Its practice and theory. *Journal of the Royal Society Interface*, 3(9), 471–482. <https://doi.org/10.1098/rsif.2006.0127>
- Voituron, Y., De Fraipont, M., Issartel, J., Guillaume, O., & Clobert, J. (2011). Extreme lifespan of the human fish (*Proteus anguinus*): A challenge for ageing mechanisms. *Biology Letters*, 7, 105–107. <https://doi.org/10.1098/rsbl.2010.0539>
- Vollrath, F., & Selden, P. (2007). The role of behavior in the evolution of spiders, silks, and webs. *Annual Review of Ecology, Evolution, and Systematics*, 38, 819–846. <https://doi.org/10.1146/annurev.ecolsys.37.091305.110221>
- von Reumont, B. M., Blanke, A., Richter, S., Alvarez, F., Bleidorn, C., & Jenner, R. A. (2014). The first venomous crustacean revealed by transcriptomics and functional morphology: Remipede venom glands express a unique toxin cocktail dominated by enzymes and a neurotoxin. *Molecular Biology and Evolution*, 31(1), 48–58. <https://doi.org/10.1093/molbev/mst199>
- Weinstein, P., & Slaney, D. (1995). Invertebrate faunal survey of rope ladder cave, northern Queensland: A comparative study of sampling methods. *Australian Journal of Entomology*, 34(3), 233–236. <https://doi.org/10.1111/j.1440-6055.1995.tb01329.x>
- Wiegand, T., Revilla, E., & Knauer, F. (2004). Dealing with uncertainty in spatially explicit population models. *Biodiversity and Conservation*, 13(1), 53–57. <https://doi.org/10.1023/B:BIOC.0000004313.86836.ab>
- Wilkins, H. (1971). Genetic interpretation of regressive evolutionary processes: Studies on hybrid eyes of two *Astyanax* cave populations (Characidae, Pisces). *Evolution*, 25(3), 530–544. <https://doi.org/10.2307/2407352>
- Wilkins, H., Parzefall, J., & Iliffe, T. M. (1986). Origin and age of the marine Stygofauna of Lanzarote, Canary Islands. *Mitteilungen Aus Den Hamburgischen Zoologischen Museum Und Institut*, 83, 223–230.
- Woodward, M. A., & Sitti, M. (2014). MultiMo-Bat: A biologically inspired integrated jumping-gliding robot. *International Journal of Robotics Research*, 33, 1511–1529. <https://doi.org/10.1177/0278364914541301>
- Wynne, J. J., Howarth, F. G., Sommer, S., & Dickson, B. G. (2019). Fifty years of cave arthropod sampling: Techniques and best practices. *International Journal of Speleology*, 48(1), 33–48. <https://doi.org/10.5038/1827-806X.48.1.2231>
- Wynne, J. J., Sommer, S., Howarth, F. G., Dickson, B. G., & Voyles, K. D. (2018). Capturing arthropod diversity in complex cave systems. *Diversity and Distributions*, 24, 1478–1491. <https://doi.org/10.1111/ddi.12772>
- Yoshizawa, M., Settle, A., Hermosura, M. C., Tuttle, L. J., Cetraro, N., Passow, C. N., & McGaugh, S. E. (2018). The evolution of a series of behavioral traits is associated with autism-risk genes in cavefish. *BMC Evolutionary Biology*, 18(1), 89. <https://doi.org/10.1186/s12862-018-1199-9>
- Zagmajster, M., Culver, D. C., Christman, M. C., & Sket, B. (2010). Evaluating the sampling bias in pattern of subterranean species richness: Combining approaches. *Biodiversity and Conservation*, 19(11), 3035–3048. <https://doi.org/10.1007/s10531-010-9873-2>
- Zakšek, V., Sket, B., Gottstein, S., Franjević, D., & Trontelj, P. (2009). The limits of cryptic diversity in groundwater: Phylogeography of the cave shrimp *Troglocaris anophthalmus* (Crustacea: Decapoda: Atyidae). *Molecular Ecology*, 18(5), 931–946. <https://doi.org/10.1111/j.1365-294X.2008.04061.x>
- Zschokke, S., & Herberstein, M. E. (2005). Laboratory methods for maintaining and studying web-building spiders. *Journal of Arachnology*, 33, 205–213. <https://doi.org/10.1636/CT04-72.1>

SUPPORTING INFORMATION

Additional supporting information may be found online in the Supporting Information section.

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