

## Expanded Croatian 12 X-STR loci database with an overview of aberrant profiles

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## 2 3 ABSTRACT

4 In order to implement X-chromosome short tandem repeat (X-STR) typing into routine forensic  
5 practice, reference database of a given population should be established. Therefore we extended  
6 already published data with additional 397 blood samples from unrelated Croatian citizens, and  
7 analyzed the total of 995 samples (549 male and 446 female) typed by Investigator® Argus X-12 Kit.  
8 To test genetic homogeneity of consecutively processed five historic-cultural regions covering the  
9 entire national territory, we calculated pairwise  $F_{st}$  genetic distances between regions based on  
10 allele and full haplotype frequencies. Since the comparison did not yield any statistically significant  
11 difference, we integrated STR profile information from all regions and used the whole data set to  
12 calculate forensic parameters. The most informative marker is DXS10135 (polymorphism information  
13 content (PIC)=0.929) and the most informative linkage group (LG) is LG1 (PIC=0.996). Cumulative  
14 power of discrimination across 12 analyzed loci amounted to 0.9999999999999996 in females and  
15 0.9999999995 in males. We confirmed linkage disequilibrium for seven marker pairs belonging to  
16 LG2, LG3 and LG4. We also compared Croatia with 13 European populations based on haplotype  
17 frequencies and detected no statistically significant  $F_{st}$  values after Bonferroni correction in any LG.  
18 Multi-dimensional scaling plot revealed tight grouping of four Croatian regions amongst populations  
19 of southern, central and northern Europe, with the exception of northern Croatia. In this study we  
20 gave the first extensive overview of aberrant profiles encountered during Investigator® Argus X-12  
21 typing. We found ten profiles consistent with single locus duplication followed by tetranucleotide  
22 tract length polymorphism. Locus DXS10079 is by far the most frequently affected one, presumably  
23 mutated in eight samples. We also found four profiles consistent with X-chromosome aneuploidy  
24 (three profiles with XXX pattern and one profile with XXY pattern). In conclusion, we established  
25 integral forensic Croatian X-chromosome database, proved forensic pertinence of Investigator®  
26 Argus X-12 Kit for the entire Croatian population and identified locus DXS10079 as mutational  
27 hotspot.

28  
29 **Keywords:** Investigator® Argus X-12; forensic X-STR markers; Croatian X-STR database; aberrant X-  
30 STR profiles; X chromosome duplication; locus DXS10079

## 31 32 33 INTRODUCTION

34 Usefulness of X-chromosome short tandem repeat (X-STR) typing has so far been very well described  
35 and established for both identification and kinship testing in the specific forensic contexts [1].  
36 Implementation of X-STR analysis into routine forensic casework primarily requires general  
37 population database for match probability calculations. As the purpose of this study is to build  
38 comprehensive forensic database based on 12 X-STR loci for the Croatian population, we expanded  
39 data set of the already published 598 profiles [2-4] with additional 397 profiles and consolidated  
40 Investigator® Argus X-12 Kit typing results. The ensuing comprehensive database encompasses 995  
41 individuals, originating from five historic-cultural regions of the northern [2], central [3], southern [4],  
42 eastern and western part of the country (**Figure 1**).

43 The Republic of Croatia is a European country bordering with Slovenia and Hungary in the north,  
44 Serbia and Bosnia and Herzegovina in the east, Montenegro in the south and Italy in the south-west.  
45 It is situated on the Adriatic Sea and positioned between Central and Southeast Europe (**Figure 1**).  
46 Regarding its ethnic composition, Croatia is a homogenous country. The population of 4.3 million  
47 mostly consists of Croats (90.4%), while minorities include Serbian (4.4%) and 21 other ethnicities  
48 (less than 1% each) [5]. Croatia in general went through a dynamic demographic history and  
49 particular geographic regions experienced quite different demographic perturbations over the  
50 centuries [6]. In that respect, we assumed that genetic substructuring at the X-STR loci might have  
51 occurred within the Croatian population. Although genetic diversity on the X-chromosome is

52 presumably lower in comparison to autosomes due to lower mutation rate and reduced effective  
53 population size, the latter feature ascertains faster genetic drift for the X-chromosome than for  
54 autosomes. As a consequence, X-chromosome exhibits more pronounced differentiation value for  
55 intra- and inter-population comparison [7]. In this regard, it is important to evaluate potential  
56 regional substructuring within the population under forensic investigation, in order to establish  
57 separate forensic databases, if needed. We therefore aimed to characterize the genetic diversity of  
58 12 X-STRs in the Croatian population.

59 With the advance of high-throughput genomics techniques, indications emerged that X-chromosome  
60 duplication events might have been underestimated previously [8]. It is therefore possible that yet  
61 undetected minor duplications of X-chromosome fragments, which do not produce obvious clinical  
62 features, appear with increased frequency in the population compared to other chromosomes. A  
63 fraction of duplication events might be detected indirectly during forensic X-STR typing, as already  
64 shown in forensic typing of autosomes [9] and Y chromosome [10, 11]. Since duplications of X-STR  
65 markers might complicate forensic interpretation due to generation of additional electropherogram  
66 peaks (male biallelic and female triallelic profiles), it seems important to report such events  
67 encountered during creation of population X-STR databases. Not only forensic community would  
68 benefit from that kind of information, but also genetic anthropology and molecular medicine.

69 Within the scope of this study, we constructed comprehensive database to achieve final goal of  
70 establishing X-STR typing as a routine forensic practice in Croatian Forensic Science Centre “Ivan  
71 Vučetić”. We computed forensic parameters to test the forensic pertinence of Investigator® Argus X-  
72 12 Kit in Croatian population and tested potential population substructuring to assess the  
73 universality of the comprehensive database. We also put Croatia into wider, inter-population context  
74 by comparing our data with previously published data for 13 European populations. Finally, in order  
75 to help forensic experts in profile evaluation and subsequent interpretation, we gave a detailed  
76 characterization of X-chromosome instability at the population level, manifested as X-STR aberrant  
77 profiles.

## 78 79 MATERIALS AND METHODS

80 The total of 203 samples (103 male and 100 female) from eastern Croatian region and the total of  
81 194 samples (146 male and 48 female) from western Croatian region were analyzed. Sampling was  
82 performed in an attempt to account for all subpopulation variations by choosing unrelated  
83 participants from the entire region covering Požega-Slavonia, Brod-Posavina, Osijek-Baranja and  
84 Vukovar-Srijem Counties of eastern Croatia, and Primorje-Gorski Kotar, Lika-Senj and Istria Counties  
85 of western Croatia. All samples were collected during routine forensic work by the staff of Forensic  
86 Science Centre “Ivan Vučetić”, and their use in the study was approved by the Ethics Committee of  
87 the Institute for Medical Research and Occupational Health, Zagreb, Croatia.

88 Genomic DNA was extracted from Flinders Technology Associates (FTA) cards (Whatman, Maidstone,  
89 Kent, UK) using Chelex-100 method [12]. Concentration of the extracted DNA was determined by  
90 Investigator® Quantiplex Kit (Qiagen GmbH, Hilden, Germany). Normalization of samples was carried  
91 out to approximately 1 ng/μL. Amplification was performed by Investigator® Argus X-12 Kit (Qiagen  
92 GmbH, Hilden, Germany) in multiplex PCR, containing primers for amelogenin (sex determination)  
93 and 12 X-STR markers belonging to four different linkage groups (LGs): LG1 (DXS10148, DXS10135,  
94 DXS8378), LG2 (DXS7132, DXS10079, DXS10074), LG3 (DXS10103, HPRTB, DXS10101), and LG4  
95 (DXS10146, DXS10134, DXS7423). Positive and negative controls were also amplified in each PCR  
96 batch. Amplification products were analyzed on 3500 Genetic Analyzer (Applied Biosystems, Foster  
97 City, CA, USA). Data obtained from capillary electrophoresis were analyzed using GeneMapper ID-X  
98 software (version 1.4, Applied Biosystems). Peak threshold values of 100 RFU and 200 RFU were  
99 applied for heterozygous and homozygous alleles, respectively. All samples containing variant alleles,  
100 female samples with triallelic patterns (**Supplementary material 3 Figures S1-S4**) and male samples  
101 with biallelic pattern (**Supplementary material 3 Figures S5-S10**) were confirmed by re-extraction  
102 followed by amplification and capillary electrophoresis. To exclude the possibility of contamination,

103 four profiles consistent with aneuploidy of X-chromosome (**Supplementary material 3 Figures S11-**  
104 **S14**) were analyzed by AmpFISTR® *SEfiler Plus*™ PCR Amplification Kit (**Supplementary material 4**  
105 **Figures S1-S4**). All procedures and protocols were carried out following manufacturers' instructions.

106

#### 107 *Statistical analysis*

108 Allele frequencies for all samples and haplotype frequencies for male samples were determined by  
109 counting the number of times each allele/haplotype was observed in all given samples. For biallelic  
110 male and triallelic female samples, allele(s) with highest frequencies were selected for further  
111 calculations. Testing for a departure from Hardy-Weinberg Equilibrium (HWE), including observed  
112 heterozygosity ( $H_o$ ) and expected heterozygosity ( $H_e$ ), was performed only for female samples.  
113 Presence of pairwise linkage disequilibrium (LD) between loci was tested by likelihood-ratio test  
114 using the Expectation-Maximization algorithm for female, and by the exact test using a Markov chain  
115 for male samples. Genetic heterogeneity within population was estimated as gene i.e. haplotype  
116 diversity ( $H$ ) for male haplotype data. For testing of genetic homogeneity among the five Croatian  
117 regions, analysis of molecular variance (AMOVA) and pairwise genetic distances ( $F_{st}$ ) were calculated  
118 based on both allele frequencies (male and female samples) and full haplotypes of male samples.  
119 Pairwise genetic distances were also calculated for inter-population comparison of haplotype  
120 frequencies between pooled Croatian population samples and 13 neighboring European populations:  
121 Slovenia, Italy, Hungary, Albania, Czechia, Germany, Greece, West Mediterranean, Lithuania, Belarus,  
122 Portugal, Denmark and Sweden [13-24]. All aforementioned computations were performed using  
123 Arlequin software v3.5.2.2 [25], while significance level for all statistical tests was set to 0.05 and  
124 corrected for multiple comparisons using Bonferroni adjustment.

125 Forensic parameters encompassing polymorphism information content (PIC), power of exclusion  
126 (PE), power of discrimination (PD) for males and for females, mean exclusion chance (MEC) for  
127 deficiency cases (Krüger's formula), MEC for normal trios consisting of a mother, a daughter and a  
128 putative father (Kishida's formula), and MEC for duos consisting of a daughter and a putative father  
129 (Desmarais' formula), were computed based on allele frequencies data using on-line tool available at  
130 ChrX-STR.org web page [26]. PIC for LGs was calculated using the R script available as supporting  
131 information from Zidkova et al. [16].

132 To additionally examine the relationship with neighboring European populations, pairwise genetic  
133 distances with sample size correction ( $F_{st}^*$ ) [27] were calculated based on allele frequencies using  
134 POPTREE2 software [28]. Inter-population comparison was presented by multi-dimensional scaling  
135 (MDS) plot constructed with IBM SPSS Statistics for Windows, Version 23.0 (IBM Corp., Armonk, NY,  
136 USA).

137

## 138 RESULTS AND DISCUSSION

139

### 140 *Intra-population comparison*

141 In addition to the already published data for northern [2], central [3] and southern Croatia [4], allele  
142 and haplotype frequencies for eastern Croatia (100 females and 103 males) and western Croatia (48  
143 females and 146 males) are given in **Supplementary material 1 (Tables S1-S4)**. In both additional  
144 regions, HWE was confirmed for all loci. All off-ladder alleles found in the sample pool of western  
145 Croatia (DXS7423 9; DXS10074 16.3, 17.2; DXS10079 19.2; DXS10101 27.3; DXS10135 15.1, 20.3;  
146 DXS10146 36.2, 48.2; DXS10148 14, 17, 22, 27.2, 32.1) and eastern Croatia (DXS10079 13, 19.2, 24;  
147 DXS10135 24.1; DXS10146 38.2, 47.2; DXS10148 22) have been reported by others. The most  
148 informative marker in both eastern and western Croatian population is DXS10135 (PIC=0.93 and  
149 0.92, respectively), while the least informative one is DXS8378 (PIC=0.62 and 0.63, respectively).

150 In order to test genetic homogeneity, pairwise  $F_{st}$  genetic distances between five Croatian regions  
151 were calculated based on both allele frequencies (male and female samples) and full haplotypes of  
152 male samples. No significant  $F_{st}$  values ( $p < 0.005$ ) after Bonferroni correction were detected in either  
153 case (**Supplementary material 1 Table S5**). Locus-by-locus AMOVA also confirmed these results (data

154 not shown). Therefore, all subsequent calculations were performed using a complete Croatian  
155 dataset of 995 samples.

156 Our results are in concordance with previous study where the variability of Croatian population had  
157 been analyzed based on Y-chromosomal haplogroup distribution, and strong similarity between the  
158 five aforementioned Croatian regions was found [6]. Consequently, there is no need for separate  
159 databases in forensic interpretation of both X-STR and Y-STR profiles obtained by corresponding kits.  
160 However, it is noteworthy that samples from small, isolated populations (e.g. islands and remote  
161 villages), where more pronounced genetic difference is expected, have not been accounted for  
162 establishing the common X-STR forensic database.

163

#### 164 *Alleles and haplotypes in forensic context*

165 In the entire Croatian data set of 995 samples (1441 chromosomes), no significant departure from  
166 HWE was detected at any locus. The total of 223 alleles across all loci was detected with frequency  
167 span from 0.00069 to 0.42679 (**Supplementary material 2 Table S1**). Marker DXS10135, comprising  
168 35 alleles in total, is the most informative (PIC=0.93), with the allele 25 as the most common  
169 (frequency of 0.103). Marker DXS8378, with the total of 7 alleles, is the least informative (PIC=0.93),  
170 and the sum of frequencies of three alleles alone (10, 11, 12) equals 0.949. Both DXS10135 and  
171 DXS8378 belong to the LG1. As expected, results for allele variability are very similar to the already  
172 published data for other European populations [13, 16, 17, 20, 24].

173 Haplotype frequencies, estimated directly from the pooled population data for each LG, are  
174 presented in **Supplementary material 2 Table S2**. The most informative linkage group is LG1 (PIC=  
175 0.9955), followed by LG4, LG2 and LG3, with frequency of the most common haplotypes equaling  
176 0.0128, 0.0182, 0.031 and 0.0364, respectively. All LGs display gene diversity value of over 0.99, with  
177 LG1 being the most diverse (0.9974), which is further corroborated by the highest PIC value of 0.9955  
178 for LG1 (**Supplementary material 2 Table S3**). LG1 is also the most polymorphic in other worldwide  
179 populations typed by Investigator® Argus X-12 Kit [13, 14, 16, 17, 24]. Of all possible haplotypes for  
180 each linkage group, 320 (4.4%), 189 (11.4%), 201 (8.9%) and 269 (3.9%) were observed in LG1, LG2,  
181 LG3 and LG4, respectively. Of the observed haplotypes, 60.9% of them are unique for LG1, 50% for  
182 LG2, 46.8% for LG3 and 57.6% for LG4. In the entire sample pool, no shared 12-marker haplotype was  
183 found.

184 To test the suitability of Investigator® Argus X-12 Kit for forensic casework in Croatian population,  
185 forensic parameters were calculated and summarized in **Table 1**. All tested parameters  
186 demonstrated high combined values, especially PD in females (0.9999999999999996), which proves  
187 the applicability of Investigator® Argus X-12 Kit in both forensic identification and kinship analysis.

188

#### 189 *Linkage disequilibrium analysis*

190 In the male sample pool, linkage disequilibrium was confirmed ( $p < 0.0008$  after Bonferroni  
191 correction) for: LG2 marker pair DXS10079-DXS10074, all three marker pairs within LG3, and LG4  
192 marker pairs DXS10134-DXS10146 and DXS7423-DXS10134 (**Supplementary material 2 Table S4**).  
193 Confirmation of the obtained results was found in female samples for LG3 marker pairs DXS10101-  
194 DXS10103 and DXS10101-HPRTB, as well as for LG4 marker pair DXS10134-DXS10146. Moreover, one  
195 additional marker pair from LG4 (DXS7423-DXS10146) was found in females (**Supplementary**  
196 **material 2 Table S5**). In total, seven out of 12 possible marker pairs across all LGs are in linkage  
197 disequilibrium. LD is confirmed for all marker pairs of LG3 and LG4. No statistically significant LD  
198 occurred between any marker pair belonging to different LGs.

199 Investigator® Argus X-12 Kit consists of four groups of closely linked STR markers which should be  
200 treated as haplotypes that are passed to the offspring unchanged [29]. Non-random association of  
201 alleles should be tested at the population level, using the actual haplotype frequencies in male  
202 samples. In populations with high genetic variability, large sample size is needed to establish all allele  
203 pairs that are in LD. This is well illustrated by typing Croatian population region by region, where the  
204 evidence for LD is demonstrated for maximum three marker pairs on a smaller sample size

205 (approximately 100 male samples). By enlargement of sample size to 549 male profiles, LD is  
206 demonstrated for six marker pairs in male samples and one additional in female samples. Lack of LD  
207 evidence for any marker pair within LG1 is probably due to the fact that LG1 is the most diverse  
208 group, including by far the greatest number of haplotypes. Therefore, in this case, even a larger  
209 sample size would be needed to confirm LD for all linked markers.

210

#### 211 *Inter-population genetic distance*

212 Haplotype frequencies were used to compare Croatian population with 13 European populations also  
213 typed by Investigator® Argus X-12 Kit. After Bonferroni correction, no significant difference  
214 ( $p < 0.00078$ ) was found in any LG (**Supplementary material 2 Table S6**). It was already shown, by us  
215 and others who addressed this issue, that European populations are indeed homogenous according  
216 to Argus X-12 markers [4, 16, 20]. There is no notable genetic variability, except for the populations  
217 that underwent specific population processes (genetic drift, interbreeding etc.), like Ibiza [19],  
218 Greenland [22], Sardinia and Southern Italy [24]. For the purpose of inter-population comparison, we  
219 chose not to include populations of different continental origins, because we had already shown  
220 correlation between genetic and geographical distance for southern Croatian population [4]. Instead,  
221 in order to obtain better resolution and determine positioning of different Croatian regions within  
222 European context of more geographically closer populations, we created a MDS plot including  
223 northern, central, southern, eastern and western Croatian populations along with 13 European  
224 populations (**Figure 2**). Left-to-right positioning of the dots representing populations along the first  
225 dimension axis of MDS plot correlates with south-to-north geographic distribution. The first  
226 dimension zero axis aligns with south-to-central Europe dividing line. Southern Croatia is positioned  
227 within the more scattered group of south European populations, while eastern, western and central  
228 Croatia belong to a tighter group consisted of central and north European populations. These results  
229 fit well with previous similar inter-population comparisons [4, 20]. Interestingly, Denmark and  
230 northern Croatia stand out as outliers at the rightmost end of the plot. Indications of more distant  
231 position of northern Croatia are visible from intra-population comparisons, where the lowest  $F_{st}$   $p$ -  
232 values (statistically significant prior to Bonferroni correction) were established between northern and  
233 eastern Croatia (**Supplementary material 1 Table S5a**), as well as northern and southern Croatia  
234 (**Supplementary material 1 Table S5b**). Besides its distant position on the plot, it is interesting to  
235 notice that northern Croatia lies nearest to Hungary. That might reflect geographic proximity and  
236 historical coexistence of Hungarian and north Croatian entities within the Austro-Hungarian Empire.

237

#### 238 *Aberrant X-STR profiles*

239 Out of 100 analyzed female profiles from eastern Croatia, two of them exhibited unexpected  
240 electropherogram patterns. One profile displayed three triple-peak patterns: at LG2 loci (DXS10079  
241 and DXS10074) and LG3 locus DXS10101 (**Supplementary material 3 Figure S11**). The other profile  
242 displayed eight triple-peak patterns: at LG1 loci (DXS10135 and DXS8378), all LG2 loci (DXS7132,  
243 DXS10079 and DXS10074), LG3 locus DXS10101 and LG4 loci (DXS10134 and DXS7423)  
244 (**Supplementary material 3 Figure S12**).

245 Out of 48 analyzed female profiles from western Croatia, one of them exhibited six triple-peak  
246 patterns: at LG1 loci (DXS10148 and DXS10135), LG2 locus DXS7132, LG3 loci (HPRTB and DXS10101)  
247 and LG4 locus DXS10146 (**Supplementary material 3 Figure S13**). Out of 146 analyzed male profiles  
248 from western Croatia, two of them showed unexpected electropherogram patterns. One profile  
249 displayed eight double-peak patterns: at LG1 loci (DXS10135 and DXS8378), LG2 loci (DXS7132 and  
250 DXS10079), LG3 loci (DXS10103 and HPRTB) and LG4 loci (DXS10146 and DXS10134) (**Supplementary**  
251 **material 3 Figure S14**). The other profile displayed two peaks at locus DXS10079, corresponding to  
252 alleles 20 and 21 (**Table 3**).

253 Taking into account all 995 Croatian samples genotyped over time by the Investigator® Argus X-12 Kit  
254 in our laboratory, as much as 14 profiles (1.4%) displayed additional peaks at either single or multiple  
255 loci. It is obvious that such percentage should not be ignored because in forensic context, aberrant

256 profiles can add complexity to data interpretation, especially in the interpretation of partial and  
257 mixed profiles. Characterization and quantification of X-chromosome instability at the population  
258 level would therefore be of help in subsequent DNA casework. In Croatian sample pool, out of 14  
259 aberrant profiles, four of them had additional peaks at multiple loci indicating aneuploidy  
260 **(Supplementary material 3 Figures S11-S14)**, while ten profiles had additional peaks at a single locus  
261 **(Supplementary material 3 Figures S1-S10)**. Single locus events are summarized in **Table 2**, and the  
262 phenomena of by far the most represented aberration (biallelic pattern in six male profiles and  
263 triallelic pattern in two female profiles at locus DXS10079) was elaborated in our recent study [4].  
264 This exemplify the importance of documenting mutational hotspots within population, considering  
265 the fact that the additional peak at DXS10079 might be expected in 0.45% female and even 1.1%  
266 male profiles in Croatian population.

267 Locus DXS10079 is a tetranucleotide STR marker located in pericentromeric region of Xq12, flanked  
268 by upstream marker DXS7132 and downstream marker DXS10074, with approximate distance of  
269 2.3Mb between each other. In humans, pericentromeric regions are by six to seven fold enriched  
270 with duplicated sequences [30]. Mutation patterns at DXS10079 in Croatian population indicate Xq12  
271 duplication of possibly rather small region between loci DXS7132 and DXS10074, followed by  
272 tetranucleotide tract length mutation. The size of duplication is assumed given that flanking markers  
273 never display duplication/mutation pattern, and that larger duplications would likely have profound  
274 clinical impact on an affected person, as has been documented for approximately 9Mb duplication  
275 comprising Xq12 region [9, 31]. Aforementioned authors hypothesize that Xq12-Xq13.3 duplication  
276 causes increased dosage of several duplicated genes that contribute to neurobehavioral phenotype  
277 [9, 31]. One of those genes, androgen receptor (AR), is located 48 kb downstream of locus DXS10079  
278 and directly upstream of locus DXS10074. It makes AR the most probable coding candidate to be  
279 duplicated in biallelic/triallelic DXS10079 samples. Expression of AR was linked to male aggression in  
280 both humans and animals [32, 33]. Although it is impossible for us to further investigate potential AR  
281 gene duplication in DXS10079 biallelic samples because of ethical considerations, it is most appealing  
282 to speculate that the reference casework sample collection might be enriched by individuals with  
283 increased androgen receptor dosage, resulting with enhanced testosterone signaling and consequent  
284 aggressive behavior. In the context of potential functional disomy, it would also be interesting to  
285 establish frequency of duplication events without subsequent addition/loss of tetranucleotide  
286 repeats. The frequency would presumably be higher, with more profound effect at the population  
287 level. However, since definitive confirmation of such events cannot be carried out by routine forensic  
288 DNA typing methods, subsequent analysis such as comparative genomic hybridization should be  
289 performed.

290 Defining DXS10079 as a duplication hotspot might go beyond Croatian population, since the  
291 occurrence of mutational events are documented in Greek [18] and Cabo Verde [34] populations on  
292 a much smaller sample size compared to Croatia. Moreover, the inauguration study for DXS10079  
293 locus described three families, out of approximately 333 parent-child trios from German population,  
294 with apparent small duplication encompassing DXS10079 [35]. In total, three female triallelic profiles  
295 and one male biallelic profile were found with one-step or two-step shift in the repeat number [35].  
296 In our study, where no parental samples were available, we simply excluded the possibility of close  
297 relatedness and endogamy of individuals from Croatian sample pool bearing DXS10079 mutations  
298 [4]. Nevertheless, it is interesting to notice that all cases except one are found in central and  
299 southern Croatia **(Table 2)**, and two remaining triallelic profiles at loci DXS10134 and DXS10146 are  
300 found in southern Croatia **(Table 2)**. It indicates some kind of regional localization, but more  
301 population and experimental data, including mitochondrial DNA and Y-chromosome haplotyping  
302 would be needed to confirm potential non-random distribution. In any case, forensic community  
303 should be encouraged to report “duplication” profiles that presently cannot be taken into account  
304 for statistical STR calculations. That practice would enhance the development of statistical calculation  
305 strategies able to account for the phenomena present in actual populations.

306 One male and three female X-STR profiles exhibited patterns consistent with aneuploidy **(Table 3)**. In  
307 all female samples we detected several triallelic loci, which indicates the presence of an additional X

308 chromosome (Triple X syndrome - 47, XXX) (**Supplementary material 3 Figures S11-S13**). In the male  
309 sample profile, we found eight biallelic loci which indicates the presence of an additional X-  
310 chromosome (Klinefelter syndrome - 47, XXY) (**Supplementary material 3 Figure S14**). Distribution of  
311 aberrant loci across different LGs, which are scattered over the entire chromosome, suggests an  
312 extra X, rather than duplication of a large chromosomal fragment (**Table 3**). This finding immediately  
313 raises the question of potential contamination that might have arisen upon re-processing of stored  
314 FTA cards. However, it is highly unlikely because all reference samples were initially genotyped when  
315 collected during casework process, and no contamination was found. Nevertheless, after repeated X-  
316 STR analysis, we performed additional AmpFLSR®SEfiler Plus™ amplification, which resulted in  
317 normal, diploid profiles with homo- or heterozygous loci in all autosomal STR markers  
318 (**Supplementary material 4 Figures S1-S4**). In addition, amelogenin peak heights also reflect an  
319 increased dosage of X-chromosome in all profiles (**Supplementary material 4 Figures S1-S4**).

320 Triple X syndrome in human population is not a rare disorder, affecting about 1 in 1000 female  
321 births. It often goes undiagnosed due to mild symptoms [36]. Our results of 0.67% female individuals  
322 affected by Trisomy X considerably exceed the established incidence rate of 0.1%. Again, as discussed  
323 for DXS10079 duplication, sample size is not sufficient neither to exclude high occurrence by chance,  
324 nor to speculate that increased percentage of affected individuals that are at risk of developing  
325 social-emotional difficulties [37] might be overrepresented in reference sample pool from casework.  
326 The finding of a single male profile indicating Klinefelter syndrome among the 549 analyzed is  
327 consistent with worldwide estimations ranging from 1 case in 500 to 1 case in 1000 live male births.

## 328 329 CONCLUSION

330 In an effort to introduce X-STR analysis for identification and kinship testing, an extensive database  
331 including 995 profiles typed by Investigator® Argus X-12 Kit has been established for the Croatian  
332 population. We conclude that Investigator® Argus X-12 Kit is suitable for forensic casework and the  
333 reference database is universally applicable in the entire Croatia.

334 Some STR markers incorporated in Investigator® Argus X-12 Kit are affected by the occurrence of  
335 additional electropherogram peaks. In Croatian population, locus DXS10079 exhibits the greatest  
336 instability manifested as biallelic pattern in males and triallelic pattern in females. Hence we  
337 conclude that locus DXS10079 is potential mutational hotspot.

338 In order to avoid misinterpretation of DNA evidence in criminal casework, information on pattern  
339 and frequency of X-STR profile aberrations should be available to forensic community.

340  
341 The DNA Analysis Unit of the Forensic Science Centre “Ivan Vučetić” is accredited according to  
342 ISO/IEC 17025, and regularly participates in quality control proficiency testing provided by the  
343 German DNA Profiling group (GEDNAP). This article follows the population data publication  
344 guidelines set by the journal [38-40].

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349 and expertise to realization of this research.

350  
351 **Conflict of interest:** none.

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353  
354 **Figure 1.** Map of Croatian regions and position of Croatia in European geographical context.

355  
356 **Figure 2.** A two-dimensional multidimensional scaling plot drawn from sample bias corrected Fst\*  
357 genetic distances calculated from the allele frequencies of 12 X-chromosome STRs included in  
358 Investigator Argus X-12 kit with the POPTREE2 software. Stress=0.1336/RSQ=0.9287. NCroatia -



359 northern Croatia; SCroatia - southern Croatia; CCroatia - central Croatia; WCroatia - western Croatia;  
360 ECroatia - eastern Croatia.  
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Table 1 Forensic parameters.

Linkage Group	Locus	No. of alleles	PIC	PE	PDfemale	PDmale	MEC Krüger	MEC Kishida	MEC Desmarais Duo
LG1	DXS10148	30	0,872603	0,762303	0,975360	0,883723	0,765966	0,872570	0,783675
	DXS10135	35	0,929327	0,863863	0,991558	0,933323	0,864861	0,929306	0,871936
	DXS8378	7	0,632163	0,417471	0,844925	0,692972	0,423151	0,632139	0,486305
LG2	DXS7132	8	0,694091	0,488352	0,887762	0,737336	0,504896	0,694079	0,554401
	DXS10079	13	0,796676	0,636639	0,944287	0,819983	0,646788	0,796664	0,678962
	DXS10074	16	0,812496	0,662387	0,951353	0,833381	0,669797	0,812496	0,699606
LG3	DXS10103	9	0,706566	0,491750	0,899270	0,739366	0,529616	0,706543	0,568508
	HPRTB	10	0,721152	0,525245	0,904084	0,758974	0,540739	0,721140	0,585899
	DXS10101	25	0,895703	0,802640	0,982865	0,903531	0,805530	0,895670	0,818506
LG4	DXS10146	30	0,892907	0,797356	0,982148	0,900948	0,801360	0,892875	0,814508
	DXS10134	33	0,845420	0,714650	0,965826	0,859992	0,723767	0,845386	0,745402
	DXS7423	7	0,655976	0,438535	0,863294	0,706594	0,458952	0,655976	0,512244
Combined			0,9999999812	0,99999817069	0,999999999999996	0,99999999954	0,99999868605	0,99999999812	0,99999954317

Table 2 Single locus affected cases (10 samples, 1% from the total 995 of analyzed).

		Sample	Locus	Genotype			Croatian Region	Supplementary material 3 Figure	Reference
Triallelic loci	females	JZ26	DXS10134	33	34	38	South	S1	[4]
		JZ101	DXS10146	27	30	31	South	S2	[4]
		SRZ38	DXS10079	14	20	21	Central	S3	[3]
		SRZ91	DXS10079	18	20	23	Central	S4	[3]
Biallelic loci	males	J8	DXS10079	20	21		South	S5	[4]
		J11		20	21		South	S6	[4]
		J57		20	21		South	S7	[4]
		Z111		20	21		West	S8	This study
		J80		20	22		South	S9	[4]
		SR241		21	22		Central	S10	[3]

Table 3 Multiple loci affected cases (four samples, 0.4% from the total of 995 analyzed). Affected loci are marked with “+”.

Sample	LG1			LG 2			LG 3			LG 4			Supplementary material 3 Figure	Croatian Region
	DXS10148	DXS10135	DXS8378	DXS7132	DXS10079	DXS10074	DXS10103	HPRTB	DXS10101	DXS10146	DXS10134	DXS7423		
Triallelic loci in female samples	I212				+	+			+				S11	East
	I221	+	+	+	+	+			+		+	+	S12	East
	Z246	+	+	+					+	+	+		S13	West
Biallelic loci in male sample	Z55	+	+	+	+		+	+		+	+		S14	West

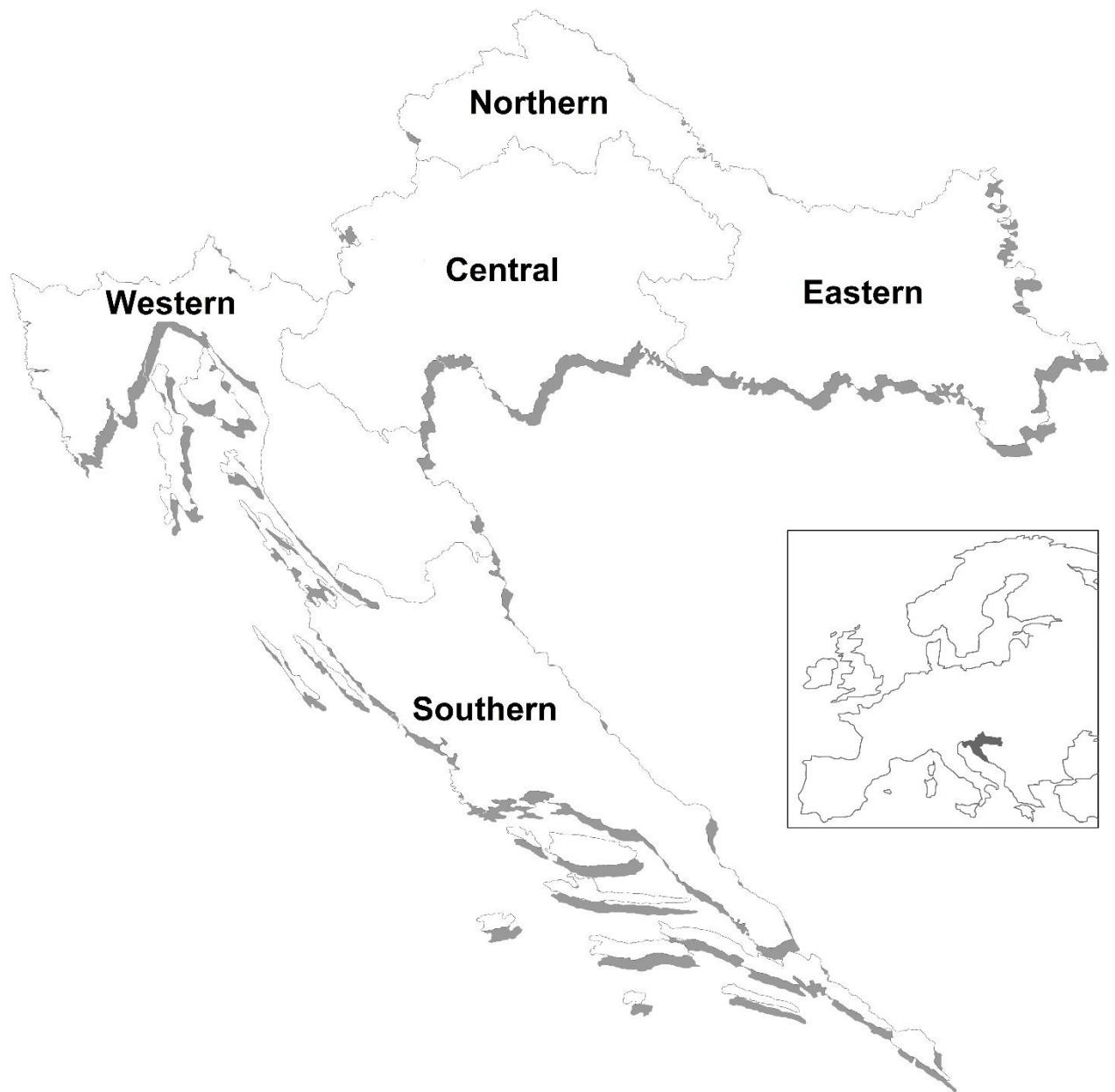


Figure 1. Map of Croatian regions and position of Croatia in European geographical context.

## Derived Stimulus Configuration

### Euclidean distance model

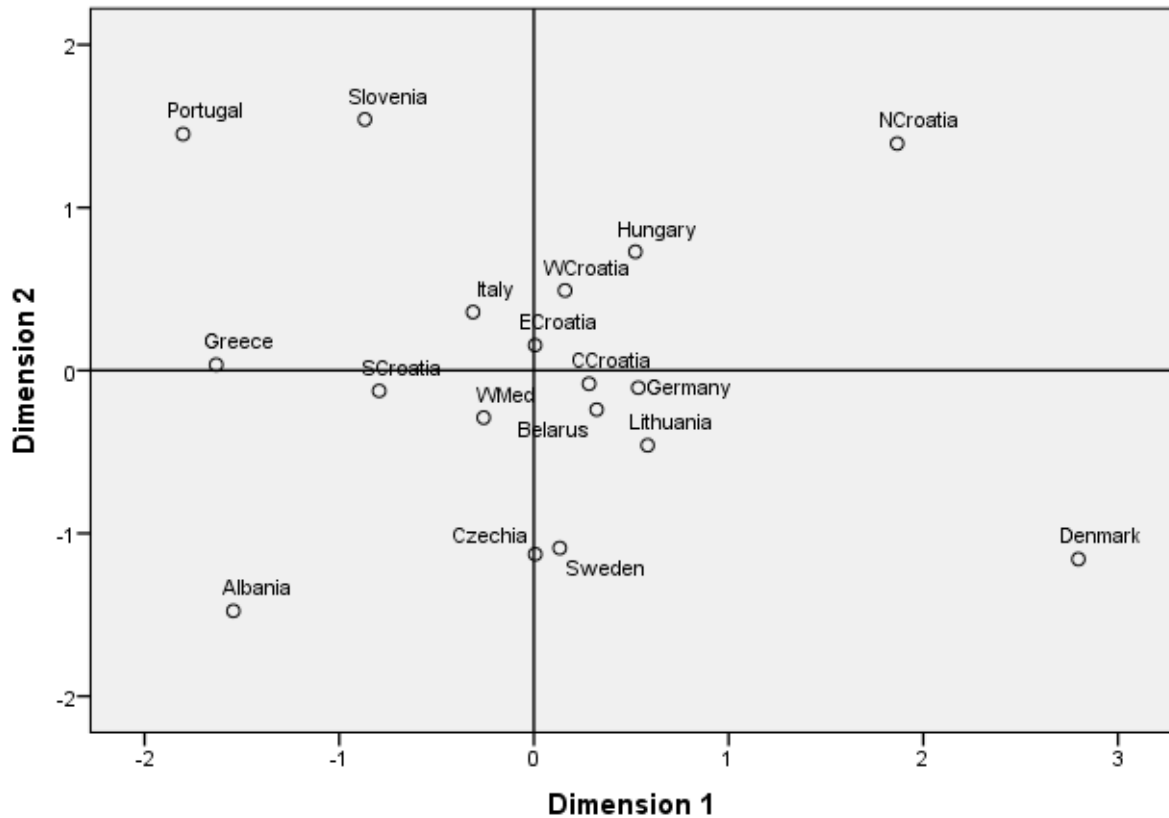


Figure 2. A two-dimensional multidimensional scaling plot drawn from sample bias corrected  $F_{st}$  genetic distances calculated from the allele frequencies of 12 X-chromosome STRs included in Investigator Argus X-12 kit with the POPTREE2 software. Stress=0.1336/RSQ=0.9287. NCroatia - northern Croatia; SCroatia - southern Croatia; CCroatia - central Croatia; WCroatia - western Croatia; ECroatia - eastern Croatia.