

1 Beta-HPV types in patients with head and neck pathology and in healthy subjects

2

3 Ivan Sabol ^{a,e}, Jana Smahelova ^{a,f}, Jan Klozar ^b, Marinka Mravak-Stipetic ^c, Tarik Gheit ^d,

4 Massimo Tommasino ^d, Magdalena Grce ^e, Ruth Tachezy ^{a,f,*}

5

6 ^a Department of Immunology, Institute of Hematology and Blood Transfusion, Prague,

7 Czech Republic ^b Department of Otorhinolaryngology and Head and Neck Surgery, First

8 Faculty of Medicine, Charles University in Prague, Motol University Hospital, Prague,

9 Czech Republic ^c Department of Oral Medicine, School of Dental Medicine, University of

10 Zagreb, Zagreb, Croatia ^d Infections and Cancer Biology Group, International Agency for

11 Research on Cancer - World Health Organization, Lyon, France ^e Department of Molecular

12 Medicine, Ruđer Bošković Institute, Zagreb, Croatia ^f Department of Genetics and

13 Microbiology, Faculty of Science, Charles University in Prague, Czech Republic

14

15 * Corresponding author:

16 Ruth Tachezy

17 Department of Immunology

18 Institute of Hematology and Blood Transfusion

19 U Nemocnice 2094/1

20 Prague 2, CZ-12820

21 Czech Republic

22 Ruth.Tachezy@uhkt.cz

23

24 **Abstract**

25 Background

26 Human papillomaviruses (HPV) are a heterogeneous group of viruses classified into five
27 genera. The beta-HPV type (beta-PV) infection is very common but mostly asymptomatic in
28 immunocompetent individuals. However, beta-PVs play a role in *epidermodysplasia*
29 *verruciformis* and possibly in non-melanoma skin cancer. Head and neck cancer (HNC) is a
30 common cancer type worldwide and high-risk alpha-PV involvement in HNC has been
31 extensively studied but beta-PV types have rarely been the focus of such studies.

32 Objectives

33 To evaluate the prevalence of beta-PV types in HNC, subjects with non-malignant or
34 potentially pre-malignant oral lesions, and healthy controls.

35 Study design

36 The frequency of different beta-PVs in samples from oral (n=35) and oropharyngeal (n=35)
37 cancer patients, gender- and age-matched healthy controls (n=70), and subjects with various
38 non-malignant or potentially pre-malignant oral lesions (n=102) was assessed by a highly
39 sensitive, bead-based, multiplex genotyping assay.

40 Results

41 Overall, 54.8% of all tested samples contained at least one beta-PV type. Even though the
42 correlation between types found in lavage and tissue specimens from cancer patients was low,
43 there was a large statistically significant difference between oropharyngeal cancer patients
44 and matched controls for HPV5 (P=0.003; OR=15.58) and between both oral (P=0.026;
45 OR=5.7) and oropharyngeal cancer patients (P=0.002; OR=25.5) and controls for HPV122. In

46 addition, there was no correlation between the prevalence of alpha and beta-PVs in the study
47 patients.

48 Conclusion

49 The data describes the prevalence of beta-PVs in HNC. The significant association of
50 HPV122 with HNC might warrant further study as this type has not been extensively studied
51 so far.

52

53 **Key words:**

54 Oral, oropharyngeal, lesion, cancer, beta-HPV, alpha-HPV

55

56 **Abbreviations:**

57 HPV, human papillomavirus; beta-PV, beta-genus papillomaviruses; HNC, head and neck
58 cancer; HR-HPV, high-risk HPV; alpha-PV, alpha-genus papillomaviruses; LR-HPV, low-
59 risk HPV; IARC, International Agency for Research on Cancer; SCC, squamous cell
60 carcinoma; TS-MPG, type-specific bead-based multiplex genotyping assay; FFPE, formalin
61 fixed paraffin embedded; PCR, polymerase chain reaction; OR, odds ratio; HIV, human
62 immunodeficiency virus

63

64

65

66

67

68 **Introduction**

69 Human papillomaviruses (HPV) are a large and heterogeneous group of viruses of the
70 *Papillomaviridae* family, which are classified into five genera. Currently, more than 200 HPV
71 types have been identified [1]. A subset of mucosal alpha-genus papillomaviruses (alpha-PV)
72 classified as high-risk human papillomaviruses (HR-HPV) cause almost all cervical and a part
73 of vaginal, vulvar, penile, anal, and oropharyngeal cancers. Low-risk alpha-PVs (LR-HPV)
74 are involved in anogenital condyloma in men and women as well as in laryngeal
75 papillomatosis [2]. HPV infection of the skin, usually with the beta-genus papillomaviruses
76 (beta-PV), is common but is mostly asymptomatic in immunocompetent individuals. In 2009,
77 the International Agency for Research on Cancer (IARC) classified two beta-PVs, HPV5 and
78 HPV8, as possibly carcinogenic in *epidermodysplasia verruciformis* patients, who have a high
79 susceptibility to beta-PV infection and skin cancer [1–5].

80 Head and neck cancer (HNC) is a diverse group of malignancies encompassing cancers of the
81 upper aerodigestive tract, paranasal sinuses, and salivary glands [6]. The most common
82 histological type is squamous cell carcinoma (SCC). HR-HPVs are implicated in the
83 development of a subset of the HNCs [7], most notably in the tonsils. According to a recent
84 comprehensive review, in oral samples from healthy individuals, alpha-PVs can be found in
85 approximately 4.5% of cases [8]; however, other studies have shown slightly greater
86 prevalence rates of 5-12% [9- 12]. There are also studies showing higher prevalence of alpha-
87 PVs in potentially premalignant oral lesions [12, 13].

88 Since the initial discovery that certain HNCs could be associated with HPV [14], the
89 involvement of HR-HPVs in HNC has been extensively studied [15–17]. However, HPVs of
90 beta- and gamma-genera (cutaneous types) have rarely been the focus of such studies [18–20].
91 Cutaneous HPVs have been frequently detected in oral cavity specimens [19, 20], but the

92 prevalence of cutaneous types in cancer tissues has been inconsistent [18, 21]. Those studies
93 mostly used the established FAP59/FAP64 primers amplifying the L1 region of 67 different
94 types [22] or CP65/CP70 and CP66/CP69 nested system amplifying the L1 region of at least
95 19 HPVs [23].

96 Herein, the frequency of different beta-PVs in samples from the oral cavity and oropharyngeal
97 cancer patients as well as from clinically distinct oral lesions and gender- and age-matched
98 healthy controls is assessed by a highly sensitive, type-specific, bead-based, multiplex
99 genotyping assay (TS-MPG), which is suitable for the detection of low-copy number HPV
100 cutaneous types and multiple infections in epidemiological analyses [24–27].

101

102 **Materials and methods**

103 Study population

104 Samples from patients with primary SCC of the oral cavity (n=35) or oropharynx (n=35),
105 collected [28] from 2001 to 2007 in the Department of Otolaryngology and Head and Neck
106 Surgery, First Faculty of Medicine, Charles University in Prague and University Hospital
107 Motol, with signed informed consent forms, were selected for analysis. For each patient, a
108 control oral rinse sample from a gender- and age-matched healthy individual was collected
109 (n=70). In addition, oral swab samples from 102 individuals with clinically distinct oral
110 lesions (potentially malignant lesions: 40.2% with *leukoplakia*, 30.4% with *lichen ruber*
111 *planus*, and 2.9% other lesions including two cases of *erythroleukoplakia* and one case of
112 *erythroplakia*; benign proliferative lesions: 10.8% papilloma and 18.6% other lesions
113 including *stomatitis simplex*, *verruca*, *chelitis*, and *aphtae*), collected during dental
114 examination at the Department of Oral Medicine, School of Dental Medicine, University of

115 Zagreb from 1995 to 2011 [12], were included in this study. The study received the official
116 institutional and ethical approval from the participating institutions.

117

118 Patient samples

119 From oral/oropharyngeal cancer patients, both oral exfoliated cells (lavage) and tumor biopsy
120 specimens were collected as described in detail previously [28]. The corresponding controls
121 provided only lavages. DNA was extracted from a minimum of 250,000 cells using Puregene
122 Core Kit A (Gentra Qiagen, Hilden, Germany) and from two 20 µm formalin fixed paraffin
123 embedded (FFPE) biopsy sections using the Ambion RecoverAll™ Total Nucleic Acid
124 Isolation Kit for FFPE Tissues (Applied Bioscience, Austin, TX). A cytobrush (Medscand
125 AB, Sweden) was used for scraping the site(s) of clinically visible lesions, which were
126 classified either by morphology or clinical diagnosis [12] and further confirmed by
127 histopathology [29]. DNA from the collected samples in TES buffer was extracted by the high
128 salt method [12].

129

130 Beta-PV detection by Luminex

131 All samples were analyzed for the presence of beta-PV types using the TS-MPG assay (IARC,
132 Lyon, France), which combines multiplex PCR with a bead-based Luminex technology [24,
133 30]. This assay detects 43 different beta-PV types, with the human beta-globin gene serving
134 as a control. Following PCR, 10 µl of each reaction mixture was analyzed using the Luminex
135 instrument (Luminex Corporation, Austin, TX) as described previously [25,31].

136

137

138 Data analysis

139 Samples with multiple infection are those with two or more HPV types detected. Such
140 samples were counted as positive for each type of HPV found. The type-specific HPV
141 prevalence is expressed as the percentage of all subjects tested for HPV, and thus represents
142 the HPV prevalence in both single and multiple infections. The odds ratio (OR) and chi-
143 square test values were calculated in Microsoft Excel (v. 2013).

144

145 **Results**

146 In this study, 242 patients and controls were tested for beta-PV presence. Overall 312 samples
147 were analysed. Oral cancer patients (median age 55 years, range 35-84) were mostly male
148 (80%), as were the oropharyngeal cancer patients (68.6%, median age 57 years, range 35-75
149 years). Only 32.4% of patients with oral lesions (median age 55 years, range 16-87) were
150 male.

151 Thirteen samples in which the beta-globin and HPV amplification was unsuccessful were
152 removed from further calculations. Those were six oral cancer samples, a lavage from one
153 oral cancer patient, three lavages from oropharyngeal controls, and three swabs of oral
154 lesions.

155 The overall beta-PV positivity is summarized in Figure 1. Table 1 shows detailed comparison
156 of the prevalence of beta-PVs in different groups of patients, matched control group, and
157 different types of samples from the same patient. The oral lesions group contained the most of
158 beta-PV positive samples (67.7%), with the greatest average number of HPV types (5.1) per
159 positive sample, followed by the oral cancer group (55.2%) and oropharyngeal tumor group
160 (45.7%). The lowest positivity rate was found in the lavage samples from patients with oral
161 cancer (17.6%), with the lowest average number of HPV types (1.2) per sample. In contrast,

162 the oral lavage samples from patients with oropharyngeal cancer were very often positive
163 (60%). The prevalence rate of beta-PV types in the control group was also very high (56.7%).
164 The beta-PV positivity per patient was evaluated by combining the data from lavage and/or
165 tissue samples of the same patient (Table 1; “Person” column). If either his/her lavage or
166 tissue sample was positive for a beta-PV type, the study subject was considered as positive for
167 that particular beta PV-type. Overall, the beta-PV type most commonly detected in study
168 subjects was HPV5 (35/236; 14.8%), but it was only the third most common one in oral
169 cancer (11.4%) after HPV122 (25.7%) and HPV151 (17.1%).

Table 1. Beta-PV types detected in different study patient groups and gender- and age-matched controls.

	Control (n=67)	Oral cancer			Oropharyngeal cancer			Oral lesion (n=99)
		Lavage (n=34)	Tissue (n=29)	Person** (n=35)	Lavage (n=35)	Tissue (n=35)	Person** (n=35)	
HPV type*	N [%]	N [%]	N [%]	N [%]	N [%]	N [%]	N [%]	N [%]
Positive	38 (56.7)	6 (17.6)	16 (55.2)	22 (62.9)	21 (60.0)	16 (45.7)	29 (82.9)	67 (67.7)
Single	16 (23.9)	5 (14.7)	5 (17.2)	8 (22.9)	7 (20.0)	8 (22.9)	7 (20.0)	16 (16.2)
Multiple	22 (32.8)	1 (2.9)	11 (37.9)	14 (40.0)	14 (40.0)	8 (22.9)	22 (62.9)	51 (51.5)
HPV5	6 (9.0)	0 (0.0)	4 (13.8)	4 (11.4)	6 (17.1)	5 (14.3)	11 (31.4)	14 (14.1)
HPV8	7 (10.4)	0 (0.0)	1 (3.4)	1 (2.9)	4 (11.4)	4 (11.4)	7 (20.0)	7 (7.1)
HPV9	0 (0.0)	0 (0.0)	1 (3.4)	1 (2.9)	1 (2.9)	1 (2.9)	2 (5.7)	9 (9.1)
HPV12	4 (6.0)	0 (0.0)	0 (0.0)	0 (0.0)	3 (8.6)	0 (0.0)	3 (8.6)	9 (9.1)
HPV14	1 (1.5)	0 (0.0)	1 (3.4)	1 (2.9)	2 (5.7)	1 (2.9)	3 (8.6)	2 (2.0)
HPV15	0 (0.0)	1 (2.9)	0 (0.0)	1 (2.9)	2 (5.7)	1 (2.9)	3 (8.6)	5 (5.1)
HPV17	2 (3.0)	1 (2.9)	0 (0.0)	1 (2.9)	0 (0.0)	0 (0.0)	0 (0.0)	7 (7.1)
HPV19	3 (4.5)	2 (2.7)	0 (0.0)	2 (5.7)	0 (0.0)	0 (0.0)	0 (0.0)	9 (9.1)
HPV20	4 (6.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	6 (6.1)
HPV21	2 (3.0)	1 (2.9)	1 (3.4)	2 (5.7)	2 (5.7)	0 (0.0)	2 (5.7)	12 (12.1)
HPV22	3 (4.5)	0 (0.0)	0 (0.0)	0 (0.0)	4 (11.4)	2 (5.7)	6 (17.1)	12 (12.1)
HPV23	2 (3.0)	0 (0.0)	0 (0.0)	0 (0.0)	2 (5.7)	0 (0.0)	2 (5.7)	10 (10.1)
HPV24	7 (10.4)	0 (0.0)	0 (0.0)	0 (0.0)	3 (8.6)	2 (5.7)	5 (14.3)	17 (17.2)

HPV25	1 (1.5)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	3 (3.0)
HPV36	5 (7.5)	1 (2.9)	0 (0.0)	1 (2.9)	1 (2.9)	0 (0.0)	1 (2.9)	18 (18.2)
HPV37	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	2 (2.0)
HPV38	8 (11.9)	0 (0.0)	0 (0.0)	0 (0.0)	5 (14.3)	0 (0.0)	5 (14.3)	18 (18.2)
HPV47	2 (3.0)	0 (0.0)	3 (10.3)	3 (8.6)	4 (11.4)	4 (11.4)	8 (22.9)	8 (8.1)
HPV49	4 (6.0)	0 (0.0)	0 (0.0)	0 (0.0)	1 (2.9)	1 (2.9)	2 (5.7)	15 (15.2)
HPV75	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	2 (2.0)
HPV76	6 (9.0)	0 (0.0)	4 (13.8)	4 (11.4)	3 (8.6)	2 (5.7)	4 (11.4)	10 (10.1)
HPV80	3 (4.5)	0 (0.0)	0 (0.0)	0 (0.0)	2 (5.7)	0 (0.0)	2 (5.7)	9 (9.1)
HPV92	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	1 (1.0)
HPV93	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	1 (2.9)	0 (0.0)	1 (2.9)	3 (3.0)
HPV96	4 (6.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	3 (3.0)
HPV98	2 (3.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	4 (4.0)
HPV99	1 (1.5)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	10 (10.1)
HPV100	5 (7.5)	0 (0.0)	0 (0.0)	0 (0.0)	1 (2.9)	1 (2.9)	2 (5.7)	1 (1.0)
HPV104	1 (1.5)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	2 (5.7)	2 (5.7)	2 (2.0)
HPV105	3 (4.5)	0 (0.0)	2 (6.9)	2 (5.7)	4 (11.4)	2 (5.7)	5 (14.3)	5 (5.1)
HPV107	3 (4.5)	1 (2.9)	4 (13.8)	4 (11.4)	3 (8.6)	1 (2.9)	4 (11.4)	13 (13.1)
HPV110	8 (11.9)	0 (0.0)	0 (0.0)	0 (0.0)	2 (5.7)	0 (0.0)	2 (5.7)	12 (12.1)
HPV111	3 (4.5)	0 (0.0)	0 (0.0)	0 (0.0)	2 (5.7)	3 (8.6)	4 (11.4)	12 (12.1)
HPV113	0 (0.0)	0 (0.0)	1 (3.4)	1 (2.9)	0 (0.0)	0 (0.0)	0 (0.0)	5 (5.1)
HPV115	1 (1.5)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	2 (2.0)

HPV118	1 (1.5)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)
HPV120	3 (4.5)	0 (0.0)	0 (0.0)	0 (0.0)	3 (8.6)	2 (5.7)	5 (14.3)	14 (14.1)
HPV122	2 (3.0)	0 (0.0)	9 (31.0)	9 (25.7)	4 (11.4)	5 (14.3)	9 (25.7)	9 (9.1)
HPV124	1 (1.5)	0 (0.0)	0 (0.0)	0 (0.0)	2 (5.7)	2 (5.7)	4 (11.4)	11 (11.1)
HPV143	2 (3.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	4 (4.0)
HPV145	5 (7.5)	0 (0.0)	1 (3.4)	1 (2.9)	1 (2.9)	3 (8.6)	3 (8.6)	9 (9.1)
HPV151	2 (3.0)	0 (0.0)	6 (20.7)	6 (17.1)	1 (2.9)	3 (8.6)	3 (8.6)	14 (14.1)

*A particular type was counted in each sample (single or multiple infection) positive for that type (maximum attribution) and thus the sum of detected types exceeds the total of the samples tested.

**Lavage and tissue results were aggregated per person tested.

1 When the cancer samples were compared to their respective matched controls, the only
2 statistically significant differences for the oral cancer subset were in the prevalence of
3 HPV122 (P=0.026; OR=5.71) and HPV24 (P=0.012; OR=0.06), while in the oropharyngeal
4 cancer subset differed significantly in the prevalence of HPV122 (P=0.002; OR=25.45),
5 HPV5 (P=0.003; OR=15.58), and HPV47 (P=0.003, OR=21.95).

6 When all cancer samples were compared with all other non-cancer samples (control and oral
7 non-cancer lesions), statistically significant differences were observed in the prevalence of
8 more HPV types: HPV122 (P<0.001; OR=4.97), HPV36 (P=0.011; OR=0.19), HPV47
9 (P=0.022; OR=2.96), HPV110 (P=0.028; OR=0.22), HPV20 (P=0.037; OR=0.11), HPV99
10 (P=0.037; OR=0.10), and HPV49 (P=0.044; OR=0.23). However, there were also significant
11 differences in HPV type specific positivity between the subjects with oral lesions and healthy
12 controls: HPV9 (P=0.011; OR=14.8), HPV151 (P=0.015; OR=5.6), HPV124 (P=0.016;
13 OR=8.63), HPV99 (P=0.028; OR=7.75), HPV120 (P=0.04; OR=3.68), HPV36 (P=0.043;
14 OR=2.89), and HPV21 (P=0.045; OR=4.69).

15 Another result of particular interest was a low concordance between the typing results of
16 biopsy tissue and oral rinse samples from the same cancer patient (n=70). Overall, there were
17 seven (10%) patients who had one of the two samples beta-globin negative and were excluded
18 from analysis, 32 (45.7%) patients with HPV detected in only one sample and the other
19 sample being HPV negative, 18 (25.7%) patients whose samples were both HPV negative, ,
20 and finally 13 (18.6%) patients who tested positive for HPV in both types of material. The
21 low concordance of HPV types in the 13 patients with both samples positive for at least one
22 HPV type is illustrated in Table 2.

Table 2. HPV typing concordance between different samples from the same patient (n=13).

Sample	HPV100	HPV104	HPV105	HPV107	HPV110	HPV111	HPV112	HPV120	HPV122	HPV124	HPV14	HPV145	HPV15	HPV151	HPV19	HPV21	HPV22	HPV23	HPV24	HPV38	HPV47	HPV49	HPV5	HPV76	HPV8	HPV80	HPV9	HPV93
8	OPH L		1	1	1		1						1					1						1				
	OPH T		1		1														1									
10	OPH L												1				1						1		1			
	OPH T								1																1			
13	OPH L																									1		1
	OPH T							1																				
23	OPH L						1																					
	OPH T																1											
30	OPH L				1																							
	OPH T										1								1						1			
62	OPH L																1						1					
	OPH T																								1			
79	OPH L																					1						
	OPH T																										1	
87	OPH L		1	1				1		1																1		
	OPH T					1			1			1	1															
110	OPH L						1					1	1		1						1				1			
	OPH T	1				1			1	1		1	1										1					
122	O L														1													
	O T								1				1															
167	O L			1																								
	O T			1									1															
178	OPH L									1									1	1					1			
	OPH T	1		1					1								1						1	1				
184	OPH L																				1							
	OPH T																						1					

OPH = oropharyngeal cancer; O = oral cancer; L = lavage fluid, T = tissue

1 **Discussion**

2 The alpha-PVs are a well acknowledged etiological factor in the development of some types
3 of head and neck cancers. However, data on the potential influence of beta-PVs are limited.
4 The purpose of this study was to provide additional data on the beta-PV prevalence in oral
5 and oropharyngeal cancers, potentially premalignant lesions, and corresponding controls. To
6 this end, 35 oral and 35 oropharyngeal cancer cases, 102 patients with pre- and non-malignant
7 oral mucosa lesions, and 70 healthy controls were evaluated by a highly sensitive TS-MPG
8 assay for beta-PV genotyping.

9 In this study, HPV was found in 41.8%, 54.9%, and 67.7% of samples from the oral region of
10 cancer patients and controls, oropharyngeal region of cancer patients and controls, and oral
11 region of oral lesion patients, respectively. The high average beta-PV prevalence observed in
12 this study in any group investigated is difficult to put in context with previous studies.

13 Koskinen et al. [21] tested 61 HNC tissues with a single-phase FAP59/64 PCR and nested
14 PCR with primers CP65/70 and CP66/69 to detect cutaneous HPVs but failed to find any.

15 Lindel et al. [18] have reported HPV in 35.0% of 51 HNC tissues where, aside from HPV6
16 and HPV16, most of the detected types belonged to the cutaneous types. Bottalico et al. [19]

17 investigated oral rinse samples from 317 HIV negative and otherwise healthy men by
18 FAP59/64 primers and hybridization and have found a beta-PV prevalence of 34.0%

19 (108/317). Paolini et al. [20] tested oral rinse/swab samples from healthy controls, non-
20 malignant lesions, and biopsy material from HNC. They used CP65/CP70 followed by

21 CP66/CP69 and FAP59/FAP64 primers. The healthy group had 25.0% beta-PV positive
22 samples, the patients with lesions had the highest beta-PV prevalence (51.0%), and only

23 20.5% of cancer tissues were beta-PV positive. The authors noted the methodological

24 differences between their study and all three previous ones and also emphasized that beta-PVs

1 are usually present in a very low copy number. Therefore, the higher prevalence of beta-PVs
2 in our study can be attributed to using a more recent and sensitive genotyping method. It
3 should also be noted that their control group had an average age of 40.7 years, while our
4 controls were age matched to the cancer patients and thus had an average age of 56.2 years,
5 which might have influenced the results [10].

6 The overall most common type in the analyzed samples was HPV5 (11.7%). This HPV type
7 was found in 9.0% samples from the combined control group, of which 11.4% and 31.4%
8 were detected in oral and oropharyngeal cancer samples, respectively. This HPV type is
9 grouped as probably carcinogenic and associated with non-melanoma skin cancer [4] but was
10 also the most common beta-PV type to be found in the oral cavity of healthy people [19].

11 Herein, there was a large statistically significant difference between the oropharyngeal cancer
12 patients and matched controls ($P=0.003$; $OR=15.58$), indicating that HPV5 should be studied
13 more closely.

14 The second leading beta-PV type in this study was HPV38 (10.4%). It is known to be highly
15 prevalent in skin cancer [32, 33] but is also very common in the plucked eyebrow hairs of
16 healthy people [34]. Antibodies against this type are also common in the general population
17 [35]. Strangely, even though HPV38 was often identified in oral cancer in a previous study
18 [36], none of the oral cancer patients had this type detectable in either lavage or biopsy
19 sample in our study. However, comparable rates of the oropharyngeal cancer patients (14.3%)
20 and controls (11.9%) had this type present in the present study.

21 The third overall most common beta-PV type was HPV122 (9.7% of samples tested; 12.3% of
22 individuals tested). Despite the data on this type being scarce, in this study, HPV122 was
23 found to be statistically significantly associated with both oral ($P=0.026$; $OR=5.7$) and

1 oropharyngeal (P=0.002; OR=25.5) cancer. Furthermore, the association was also significant
2 in the overall comparison of cancer *versus* all non-cancer samples (including control and non-
3 cancer oral lesions). The lack of other reports of the presence of this HPV type in HNC might
4 be due to the fact that it was only recently described [1], and some of the methods previously
5 used for the detection of beta-PVs do not specifically target HPV122.

6 The prevalence of alpha-PVs in the same cancer patients has been previously determined, and
7 as expected, alpha-PVs were more often found in the cancer patients (49.0%) compared with
8 the matched controls (12.0%) [11]. However, a study on a subset of the same cancer patients
9 comparing tumor tissue and lavage specimens from the same patient has observed that the
10 alpha-PV overall positivity rate is slightly lower in the rinse specimens than in tumor tissue
11 specimens (53.0% vs 59.0%), with a greater diversity of types and multiple infection being
12 more often found in the rinse specimens (12.0% vs 2.4%) [28]. The difference in alpha-PV
13 positivity of oral cancer patients between tumor tissue and rinse specimens was more
14 pronounced (16.7% vs 8.3%) in comparison with oropharyngeal cancer patients (67.5% vs
15 62.4%). Similarly, more than double difference was found in beta-PV positivity of oral cancer
16 patients between cancer tissue and rinse samples (55.2% vs 17.7%). On the other hand, in
17 oropharyngeal cancer patients, unlike the alpha-PVs, the beta-PVs were less prevalent in
18 tissue samples (45.7%) than in the rinse samples (60%). The co-positivity rates for alpha- and
19 beta-types were 1%, 6%, 10%, and 29% in controls, oral cancers, oral lesions, and
20 oropharyngeal cancers, respectively, but with no correlation (correlation coefficients ranged
21 from -0.2 to 0.09, without statistical significance) (Table 3).

22

23

1 **Table 3.** Positivity of controls and patients with oral/oropharyngeal tumors for HPVs of alpha
 2 and beta-genera.

Group of subjects	N	Alpha-HPV		Overall HPV prevalence [%]	
		Positive	Negative	Alpha	Beta
Controls	67	5	62	7.5	56.7
Positive	38	1 (1.5)	37 (55.2)		
Negative	29	4 (6.0)	25 (37.3)		
Oral lesions	99	14	85	14.1	67.7
Positive	67	10 (10.1)	57 (57.6)		
Negative	32	4 (4.0)	28 (28.3)		
Beta-HPV	63	9	54	14.3	34.9
Oral cancers	63	9	54		
Positive	22	4 (6.3)	18 (28.6)		
Negative	41	5 (7.9)	36 (57.1)		
Oropharyngeal cancers	70	41	29	58.6	52.9
Positive	37	20 (28.6)	17 (24.3)		
Negative	33	21 (30.0)	12 (17.1)		

3 *percentages were calculated on the sample group subtotal

4

1 One of the limitations of the present study is the availability of different types of samples
2 collected from cancer patients and the respective controls. However, prior studies often used
3 the same types of clinical material [21] due to ethical and medical reasons preventing biopsy
4 sampling of healthy subjects. Thus, our data are readily comparable to those from the
5 previous studies in this regard. Furthermore, in our present study, we were able to compare
6 the prevalence and type-specificity in two different types of the clinical material taken from
7 the same patient. We have previously shown for Alpha PV types that while in tumor tissue,
8 only HR HPV types are present and multiple infection is rare, in oral rinses, multiple infection
9 is more frequent and also LR HPV types as a single infection can be detected. Nevertheless,
10 in patients with HR HPV-positive tumors, a concordant HR HPV type was present in 97.0%
11 of oral rinses, suggesting viral shedding from the tumor [29]. In contrast, the concordance was
12 poor for Beta-PV types. We hypothesize that it is due to the non-etiological association of the
13 Beta-PV types with studied type of cancers. It is important to note that the biggest difference
14 in the two types of materials analyzed in cancer patients is that the oral rinse provides
15 information about the presence of viruses in the whole oral and proximal part of the
16 oropharyngeal area while tissue sections provide information of the presence of HPV types in
17 the tumor itself and in its direct proximity.

18 One of the major strengths of this study is the use of the optimized multiplex PCR/Luminex
19 assay that can readily distinguish a large number of genotypes in a single sample. The
20 previous methods based on sequencing of PCR amplicons generated by consensus primers
21 were limited in this regard.

22 In summary, this study assessed the presence of the so far understudied beta-PVs in HNC
23 patients, matched controls, and subjects with oral lesions using the newest TS-MPG assay
24 specifically designed to target beta-PVs. The most prevalent type was the probably

1 carcinogenic HPV5, while the second most common was HPV38. The third leading type was
2 HPV122 that has been so far almost completely absent from the literature, and herein it was
3 found to be statistically significantly associated with cancer, which might warrant further
4 interest and study. Finally, poor HPV type concordance between patient matched lavage and
5 tissue samples suggests that beta-PVs are not strongly etiologically linked to HNC, even if
6 some associations are statistically significant.

7

1 **Funding**

2 MG, IS and MMS were partially supported by the National Science Foundation of Croatia
3 (grant No 4758). TG and MT were partially supported by the European Commission, grant
4 HPV-AHEAD (FP7-HEALTH-2011-282562). RT and JS were partially supported by the
5 Ministry of Education, Youth and Sports of CR within the National Sustainability Program II
6 (Project BIOCEV-FAR) LQ1604 and by the project BIOCEV (CZ.1.05/1.1.00/02.0109).

7 **Competing interests**

8 The authors have no competing interests.

9 **Ethical approval**

10 All study participants signed an informed consent form. The study received the official
11 institutional and ethical approval from the participating institutions according to the national
12 regulations.

13

1 **References**

- 2 [1] H.-U. Bernard, R.D. Burk, Z. Chen, K. van Doorslaer, H. zur Hausen, E.-M. de
3 Villiers, Classification of papillomaviruses (PVs) based on 189 PVs and proposal of
4 taxonomic amendments, *Virology*. 401 (2010) 70–79. doi:10.1016/j.virol.2010.02.002.
- 5 [2] F.X. Bosch, T.R. Broker, D. Forman, A.-B. Moscicki, M.L. Gillison, J. Doorbar, et al.,
6 Comprehensive control of human papillomavirus infections and related diseases, *Vaccine*. 31
7 Suppl 7 (2013) H1–31. doi:10.1016/j.vaccine.2013.10.003.
- 8 [3] S. Jablonska, J. Dabrowski, K. Jakubowicz, Epidermodysplasia verruciformis as a
9 model in studies on the role of papovaviruses in oncogenesis, *Cancer Res*. 32 (1972) 583–589.
- 10 [4] V. Bouvard, R. Baan, K. Straif, Y. Grosse, B. Secretan, F. El Ghissassi, et al., A
11 review of human carcinogens--Part B: biological agents, *Lancet Oncol*. 10 (2009) 321–322.
- 12 [5] B. Aldabagh, J.G.C. Angeles, A.R. Cardones, S.T. Arron, Cutaneous squamous cell
13 carcinoma and human papillomavirus: is there an association?, *Dermatol. Surg. Off. Publ.*
14 *Am. Soc. Dermatol. Surg. Al*. 39 (2013) 1–23. doi:10.1111/j.1524-4725.2012.02558.x.
- 15 [6] H. Mehanna, V. Paleri, C.M.L. West, C. Nutting, Head and neck cancer--Part 1:
16 Epidemiology, presentation, and prevention, *BMJ*. 341 (2010) c4684–c4684.
17 doi:10.1136/bmj.c4684.
- 18 [7] C. Hobbs, J. Sterne, M. Bailey, R. Heyderman, M. Birchall, S. Thomas, Human
19 papillomavirus and head and neck cancer: a systematic review and meta-analysis, *Clin.*
20 *Otolaryngol*. 31 (2006) 259–266. doi:10.1111/j.1749-4486.2006.01246.x.

- 1 [8] A.R. Kreimer, R.K. Bhatia, A.L. Messeguer, P. González, R. Herrero, A.R. Giuliano,
2 Oral human papillomavirus in healthy individuals: a systematic review of the literature, *Sex.*
3 *Transm. Dis.* 37 (2010) 386–391. doi:10.1097/OLQ.0b013e3181c94a3b.
- 4 [9] G. D’Souza, N. Kluz, A. Wentz, R.M. Youngfellow, A. Griffioen, E. Stammer, et al.,
5 Oral Human Papillomavirus (HPV) Infection among Unvaccinated High-Risk Young Adults,
6 *Cancers.* 6 (2014) 1691–1704. doi:10.3390/cancers6031691.
- 7 [10] C.H. Chung, A. Bagheri, G. D’Souza, Epidemiology of oral human papillomavirus
8 infection, *Oral Oncol.* 50 (2014) 364–369. doi:10.1016/j.oraloncology.2013.09.003.
- 9 [11] R. Tachezy, J. Klozar, L. Rubenstein, E. Smith, M. Saláková, J. Smahelová, et al.,
10 Demographic and risk factors in patients with head and neck tumors, *J. Med. Virol.* 81 (2009)
11 878–887. doi:10.1002/jmv.21470.
- 12 [12] M. Mravak-Stipetić, I. Sabol, J. Kranjčić, M. Knežević, M. Grce, Human
13 Papillomavirus in the Lesions of the Oral Mucosa According to Topography, *PLoS ONE.* 8
14 (2013) e69736. doi:10.1371/journal.pone.0069736.
- 15 [13] D. Dalla Torre, D. Burtscher, M. Edlinger, E. Sölder, A. Widschwendter, M. Rasse, et
16 al., Comparison of the prevalence of human papilloma virus infection in histopathologically
17 confirmed premalignant oral lesions and healthy oral mucosa by brush smear detection, *Oral*
18 *Surg. Oral Med. Oral Pathol. Oral Radiol.* 119 (2015) 333–339.
19 doi:10.1016/j.oooo.2014.11.013.
- 20 [14] M.L. Gillison, W.M. Koch, K.V. Shah, Human papillomavirus in head and neck
21 squamous cell carcinoma: are some head and neck cancers a sexually transmitted disease?,
22 *Curr. Opin. Oncol.* 11 (1999) 191–199.

- 1 [15] R. Woods, E.M. O'Regan, S. Kennedy, C. Martin, J.J. O'Leary, C. Timon, Role of
2 human papillomavirus in oropharyngeal squamous cell carcinoma: A review, *World J. Clin.*
3 *Cases.* 2 (2014) 172–193. doi:10.12998/wjcc.v2.i6.172.
- 4 [16] T. Isayeva, Y. Li, D. Maswahu, M. Brandwein-Gensler, Human papillomavirus in
5 non-oropharyngeal head and neck cancers: a systematic literature review, *Head Neck Pathol.*
6 *6 Suppl 1* (2012) S104–120. doi:10.1007/s12105-012-0368-1.
- 7 [17] H. Mirghani, F. Amen, F. Moreau, J. Lacau St Guily, Do high-risk human
8 papillomaviruses cause oral cavity squamous cell carcinoma?, *Oral Oncol.* (2014).
9 doi:10.1016/j.oraloncology.2014.11.011.
- 10 [18] K. Lindel, B. Helmke, C. Simon, K.J. Weber, J. Debus, E.-M. de Villiers, Cutaneous
11 human papillomavirus in head and neck squamous cell carcinomas, *Cancer Invest.* 27 (2009)
12 781–787. doi:10.1080/07357900802653456.
- 13 [19] D. Bottalico, Z. Chen, A. Dunne, J. Ostoloza, S. McKinney, C. Sun, et al., The oral
14 cavity contains abundant known and novel human papillomaviruses from the
15 Betapapillomavirus and Gammapapillomavirus genera, *J. Infect. Dis.* 204 (2011) 787–792.
16 doi:10.1093/infdis/jir383.
- 17 [20] F. Paolini, C. Rizzo, I. Sperduti, B. Pichi, B. Mafera, S.S. Rahimi, et al., Both mucosal
18 and cutaneous papillomaviruses are in the oral cavity but only alpha-genus seems to be
19 associated with cancer, *J. Clin. Virol. Off. Publ. Pan Am. Soc. Clin. Virol.* 56 (2013) 72–76.
20 doi:10.1016/j.jcv.2012.09.016.

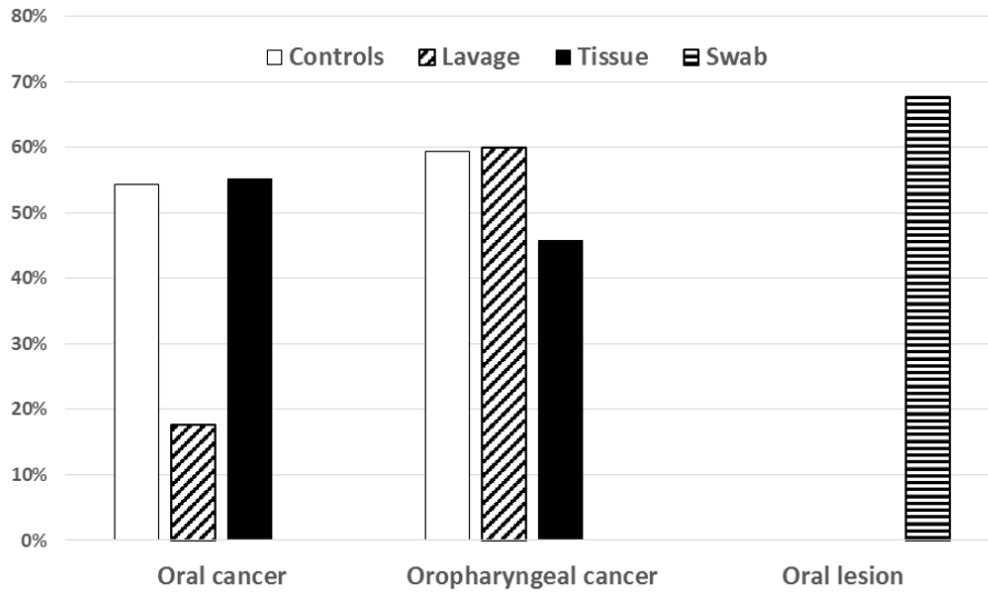
- 1 [21] W.J. Koskinen, R.W. Chen, I. Leivo, A. Mäkitie, L. Bäck, R. Kontio, et al., Prevalence
2 and physical status of human papillomavirus in squamous cell carcinomas of the head and
3 neck, *Int. J. Cancer*. 107 (2003) 401–406. doi:10.1002/ijc.11381.
- 4 [22] O. Forslund, A. Antonsson, P. Nordin, B. Stenquist, B. Goran Hansson, A broad range
5 of human papillomavirus types detected with a general PCR method suitable for analysis of
6 cutaneous tumours and normal skin, *J. Gen. Virol.* 80 (1999) 2437-2443.
- 7 [23] R.J. Berkhout, L.M. Tieben, H.L. Smits, J.N. Bavinck, B.J. Vermeer, J. ter Schegget,
8 Nested PCR approach for detection and typing of epidermodysplasia verruciformis-associated
9 human papillomavirus types in cutaneous cancers from renal transplant recipients, *J. Clin.*
10 *Microbiol.* 33 (1995) 690–695.
- 11 [24] T. Gheit, G. Billoud, M.N.C. de Koning, F. Gemignani, O. Forslund, B.S. Sylla, et al.,
12 Development of a Sensitive and Specific Multiplex PCR Method Combined with DNA
13 Microarray Primer Extension To Detect Betapapillomavirus Types, *J. Clin. Microbiol.* 45
14 (2007) 2537–2544. doi:10.1128/JCM.00747-07.
- 15 [25] M. Schmitt, B. Dondog, T. Waterboer, M. Pawlita, M. Tommasino, T. Gheit,
16 Abundance of Multiple High-Risk Human Papillomavirus (HPV) Infections Found in
17 Cervical Cells Analyzed by Use of an Ultrasensitive HPV Genotyping Assay, *J. Clin.*
18 *Microbiol.* 48 (2010) 143–149. doi:10.1128/JCM.00991-09.
- 19 [26] C.M. Pierce Campbell, J.L. Messina, M.H. Stoler, D.M. Jukic, M. Tommasino, T.
20 Gheit, et al., Cutaneous human papillomavirus types detected on the surface of male external
21 genital lesions: a case series within the HPV Infection in Men Study, *J. Clin. Virol. Off. Publ.*
22 *Pan Am. Soc. Clin. Virol.* 58 (2013) 652–659. doi:10.1016/j.jcv.2013.10.011.

- 1 [27] S.S. Hampras, A.R. Giuliano, H.-Y. Lin, K.J. Fisher, M.E. Abrahamsen, B.A. Sirak, et
2 al., Natural history of cutaneous human papillomavirus (HPV) infection in men: the HIM
3 study, *PloS One*. 9 (2014) e104843. doi:10.1371/journal.pone.0104843.
- 4 [28] E. Koslabova, E. Hamsikova, M. Salakova, J. Klozar, E. Foltynova, E. Salkova, et al.,
5 Markers of HPV infection and survival in patients with head and neck tumors, *Int. J. Cancer*.
6 133 (2013) 1832–1839. doi:10.1002/ijc.28194.
- 7 [29] S. Warnakulasuriya, N.W. Johnson, I. Van Der Waal, Nomenclature and classification
8 of potentially malignant disorders of the oral mucosa, *J. Oral Pathol. Med.* 36 (2007) 575–
9 580. doi:10.1111/j.1600-0714.2007.00582.x.
- 10 [30] T. Gheit, S. Landi, F. Gemignani, P.J.F. Snijders, S. Vaccarella, S. Franceschi, et al.,
11 Development of a sensitive and specific assay combining multiplex PCR and DNA
12 microarray primer extension to detect high-risk mucosal human papillomavirus types, *J. Clin.*
13 *Microbiol.* 44 (2006) 2025–2031. doi:10.1128/JCM.02305-05.
- 14 [31] M. Schmitt, I.G. Bravo, P.J.F. Snijders, L. Gissmann, M. Pawlita, T. Waterboer, Bead-
15 based multiplex genotyping of human papillomaviruses, *J. Clin. Microbiol.* 44 (2006) 504–
16 512. doi:10.1128/JCM.44.2.504-512.2006.
- 17 [32] E.M. de Villiers, Human papillomavirus infections in skin cancers, *Biomed.*
18 *Pharmacother. Bioméd. Pharmacothérapie.* 52 (1998) 26–33.
- 19 [33] G. Astori, D. Lavergne, C. Benton, B. Höckmayr, K. Egawa, C. Garbe, et al., Human
20 papillomaviruses are commonly found in normal skin of immunocompetent hosts, *J. Invest.*
21 *Dermatol.* 110 (1998) 752–755. doi:10.1046/j.1523-1747.1998.00191.x.

- 1 [34] K.F. Komloš, B.J. Kocjan, A. Sterbenc, M.M. Jelen, K. Seme, P. Košorok, et al.,
2 Genomic distribution of beta papillomaviruses in single eyebrow hair samples and pools of
3 eyebrow hair samples, *Acta Dermatovenerol. Alp. Panon. Adriat.* 20 (2011) 155–160.
- 4 [35] A. Antonsson, A.C. Green, K. Mallitt, P.K. O'Rourke, N. Pandeya, M. Pawlita, et al.,
5 Prevalence and stability of antibodies to 37 human papillomavirus types--a population-based
6 longitudinal study, *Virology*. 407 (2010) 26–32. doi:10.1016/j.virol.2010.07.046.
- 7 [36] A. Kojima, H. Maeda, Y. Sugita, S. Tanaka, Y. Kameyama, Human papillomavirus
8 type 38 infection in oral squamous cell carcinomas, *Oral Oncol.* 38 (2002) 591–596.
- 9

1 **Figure 1.** Beta-PVs detected in head and neck squamous cell carcinomas (HNC) and oral
2 lesions (the control groups are age and sex matched to HNC patients).

3



4