

Manuscript Number: STOTEN-D-13-02921

Title: Assessment of stability of drug biomarkers in municipal wastewater as a factor influencing the estimation of drug consumption using sewage epidemiology

Article Type: SI: Illicit Drugs

Keywords: illicit drugs; stability; urinary biomarkers, degradation, transformation, municipal wastewater, LC-MS-MS

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Abstract: Stability of the selected urinary biomarkers of six illicit drugs and two therapeutic opioids in municipal wastewater was studied in order to determine errors associated with their possible transformation in the sewer. The stability was assessed in experiments conducted at 10 °C and 20 °C in order to simulate typical winter and summer temperature conditions in the sewer system. Among fourteen substances tested, the most unstable compounds were morphine-3-β-D glucuronide (MG), 6-acetyl morphine (6-AM), cocaine (COC) and 6-acetyl codeine (6-AC), while all other investigated compounds appeared to be highly stable over a period of 72 hours. The transformation of all degradable compounds followed pseudo-first order kinetics with significantly longer half-times ($t_{1/2}$) at winter conditions. At 20 °C, $t_{1/2}$ of MG, 6-AM, COC and 6-AC was 7 h, 77 h, 35 h and 58 h, respectively, while the corresponding $t_{1/2}$ values at 10 °C were 18 h, 139 h, 173 h and 87 h. The main transformation mechanism of MG, 6-AM and 6-AC was most probably their enzymatic hydrolysis to morphine (MOR), while COC transformation to benzoylecgonine (BE) was primarily governed by chemical hydrolysis. The results from this study indicate that the observed degradation of COC and 6-AM would not significantly affect the estimates of COC and heroin consumption if the in-sewer hydraulic retention time is lower than 12 h. Acidification of the wastewater samples proved to be the good way to stabilise the wastewater samples for the analysis of all selected compounds, except for 11-nor-9-carboxy-Δ⁹-tetrahydrocannabinol (THC-COOH). This finding should be taken into account when selecting the preservation technique for multiresidual analyses of different groups of illicit drugs.

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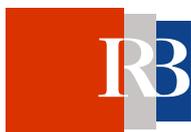
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Zagreb, 06 September 2013

Dear Editor,

please find enclosed our manuscript on *Assessment of stability of drug biomarkers in municipal wastewater as a factor influencing the estimation of drug consumption using sewage epidemiology*.

We hope that you will find this study suitable for publication in the special issue of the Science of the Total Environment.

Please send all further correspondence to me (terzic@irb.hr).

Sincerely yours,

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Highlights

- Stability of fourteen urinary drug biomarkers in municipal wastewater was evaluated
- The most unstable compounds were cocaine, morphine glucuronide and acetyl morphine
- The degradation followed pseudo first order kinetics and was temperature dependant
- In-sewer changes of common drugs do not strongly affect their consumption estimates

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1 Assessment of stability of drug biomarkers in municipal wastewater as a factor influencing
2 the estimation of drug consumption using sewage epidemiology

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4 23 **Abstract**
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8 25 Stability of the selected urinary biomarkers of six illicit drugs and two therapeutic opioids
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10 26 in municipal wastewater was studied in order to determine errors associated with their
11 27 possible transformation in the sewer. The stability was assessed in experiments conducted
12 28 at 10 °C and 20 °C in order to simulate typical winter and summer temperature conditions
13 29 in the sewer system. Among fourteen substances tested, the most unstable compounds were
14 30 morphine-3-β-D glucuronide (MG), 6-acetyl morphine (6-AM), cocaine (COC) and 6-
15 31 acetyl codeine (6-AC), while all other investigated compounds appeared to be highly stable
16 32 over a period of 72 hours. The transformation of all degradable compounds followed
17 33 pseudo-first order kinetics with significantly longer half-times ($t_{1/2}$) at winter conditions. At
18 34 20 °C, $t_{1/2}$ of MG, 6-AM, COC and 6-AC was 7 h, 77 h, 35 h and 58 h, respectively, while
19 35 the corresponding $t_{1/2}$ values at 10 °C were 18 h, 139 h, 173 h and 87 h. The main
20 36 transformation mechanism of MG, 6-AM and 6-AC was most probably their enzymatic
21 37 hydrolysis to morphine (MOR), while COC transformation to benzoylecgonine (BE) was
22 38 primarily governed by chemical hydrolysis. The results from this study indicate that the
23 39 observed degradation of COC and 6-AM would not significantly affect the estimates of
24 40 COC and heroin consumption if the in-sewer hydraulic retention time is lower than 12 h.
25 41 Acidification of the wastewater samples proved to be the good way to stabilise the
26 42 wastewater samples for the analysis of all selected compounds, except for 11-nor-9-
27 43 carboxy-Δ9-tetrahydrocannabinol (THC-COOH). This finding should be taken into account
28 44 when selecting the preservation technique for multiresidual analyses of different groups of
29 45 illicit drugs.
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49 **Keywords:** illicit drugs; stability; urinary biomarkers, degradation, transformation,
50 municipal wastewater, LC-MS-MS

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4 53 **1. Introduction**
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8 55 Municipal wastewater effluents are usually regarded as one of the main sources of input of
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10 56 different types of contaminants into the environment. However, the analysis of municipal
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12 57 wastewater has been recently increasingly used as a valuable source of information about a
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14 58 given community, including the estimation of collective drug abuse (Karolak et al., 2010;
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16 59 Kasprzyk-Hordern et al., 2009; Postigo et al., 2010; Terzic et al., 2010; Thomas et al.,
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18 60 2012; van Nuijs et al., 2011; Zuccato et al., 2005; Zuccato et al., 2008). Namely, municipal
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20 61 wastewater contains a very large number of versatile compounds excreted by humans after
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22 62 the consumption of different illegal and legal drugs. Having the data on the metabolic
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24 63 pathways of selected drugs of abuse and wastewater flow, the concentrations of selected
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26 64 urinary drug biomarkers could be used to estimate collective drug consumption. This
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28 65 innovative approach has a potential to become a rather useful complementary tool to the
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30 66 existing epidemiological methods, although further evaluation and standardisation is
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32 67 needed. The reliability of the consumption estimates is not dependent only on the accuracy
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34 68 of the chemical measurements but, among other things, on the stability of the selected
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36 69 urinary biomarker in the sewer system, as well as during the sample collection and storage
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38 70 (Castiglioni et al., 2013). Stability of the selected urinary drug biomarkers in the
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40 71 wastewater has already been assessed by several research groups (Baker and Kasprzyk-
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42 72 Hordern, 2011; Castiglioni et al., 2006, 2011; Chiaia et al., 2008; Gonzalez-Marino et al.,
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44 73 2010; Plosz et al., 2013; van Nuijs et al., 2012). The setup of these experiments was rather
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46 74 different in terms of temperature and pH conditions used, number and type of compounds
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48 75 studied, as well as the duration of the experiment. Most of the published stability
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50 76 experiments covered time scales from 12 to 72 h. In most of the cases, the samples were
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52 77 analysed only at the beginning and at the end of the experiment, while only two studies
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54 78 included sampling at multiple shorter time intervals (van Nuijs et al., 2012, Plosz et al.,
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56 79 2013). Furthermore, the experiments were focused mainly on the stability of wastewater
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58 80 samples during the collection and storage (Castiglioni et al., 2006, 2011; Gonzalez-Marino
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60 81 et al., 2010), while only limited number of them was performed applying the temperature
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62 82 and pH conditions typical for sewer systems. (Baker and Kasprzyk-Hordern, 2011; Plosz et

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4 83 al., 2013; van Nuijs et al., 2012). The compilation of the literature data obtained in different
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6 84 stability studies (van Nuijs et al., 2012, Castiglioni, 2013) show that the results are not
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8 85 always consistent and additional data are needed in order to better understand the fate of
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10 86 drug target residues in the sewer system. For instance, Baker and Kasprzyk-Hordern (2011)
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12 87 reported the significant increase of amphetamine concentration ($\approx 50\%$) after 12 h-
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14 88 experimental period (19 °C, pH 7.4), while no significant change in the concentration of
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16 89 this compound was observed in the experiments performed by Castiglioni et al. (2006) and
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18 90 van Nuijs et al. (2012) at fairly similar experimental conditions (20 °C, pH 7.4-7.5).
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20 91 Furthermore, Gonzalez-Marino et al. (2010) reported a complete loss of methadone after 72
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22 92 h at 4 °C, while other studies reported its high stability in the wastewater (Castiglioni et al.,
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24 93 2006; Baker and Kasprzyk-Hordern 2011; van Nuijs et al., 2012). Besides that, all
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26 94 experiments mimicking the sewer conditions were performed at the typical summer
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28 95 temperature conditions (19 °C or 20 °C).

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30 96 The aim of this paper was to study the stability of fourteen selected urinary biomarker
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32 97 compounds excreted after the consumption of six illicit and two licit drugs at the typical
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34 98 winter and summer in-sewer temperatures and to assess the impact of their potential
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36 99 degradation/formation on the estimation of drug consumption based on the wastewater
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38 100 analysis. Additionally, the stability of the selected drugs during the collection of the 24-h
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40 101 composite wastewater samples was also assessed as a possible source of error in the
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42 102 estimation of drug abuse.

43 103 44 104 45 105 **2. Experimental**

46 106 47 107 *2.1. Selection of analytes*

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50 109 The stability experiments encompassed 13 substances that are excreted after consumption
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52 110 of 6 illegal drugs and 2 therapeutic opioids. The target analytes included morphine (MOR),
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54 111 6-acetyl morphine (6-AM) and morphine-3- β -D glucuronide (MG) as principal heroin-
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56 112 derived substances, while 6-acetyl codeine (6-AC) was selected as a structural analogue of
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113 6-AM. Cocaine (COC) and its main metabolite benzoylecgonine (BE) were selected as the
114 main urinary biomarkers of COC. The amphetamine-type drugs included amphetamine
115 (AMP), 3,4-methylenedioxymethamphetamine (MDMA, ecstasy) and methamphetamine
116 (MAMP), while 11-nor-9-carboxy- Δ^9 -tetrahydrocannabinol (THC-COOH) and 11-
117 hydroxy- Δ^9 -tetrahydrocannabinol (THC-OH) were selected as biomarkers of cannabis
118 consumption. Methadone (MTHD) and its metabolite 2-ethylidene-1,5-dimethyl-3,3-
119 diphenylpyrrolidine (EDDP) were monitored as representatives of therapeutic drugs used in
120 the treatment of heroin addicts. The list of all investigated compounds is presented in the
121 Electronic supplementary material (Table S1).

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123 2.2. Chemicals and materials

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125 Standard solutions of all target analytes and their deuterated analogues (Table 1) were
126 purchased from Lipomed AG (Switzerland) at concentration of 1 mg/mL and 0.1 mg/mL,
127 respectively. Mixed standard solutions of the analytes and their deuterated analogues were
128 prepared in methanol (MeOH) at the concentration level of 10 μ g/mL and 2 μ g/mL,
129 respectively, and kept in the dark at -20 °C. MeOH (J.T.Baker, Deventer, the Netherlands)
130 and acetic acid (Fluka, Switzerland) were LC-MS grade. Aqueous ammonia (NH₃) solution
131 (25%), phosphoric acid (H₃PO₄) and acetic acid (CH₃COOH) were also supplied by Fluka
132 (Fluka, Switzerland). Water was purified using Elix-Milli-Q system (Millipore, Bedford,
133 USA).

134 Oasis HLB (200 mg/6 mL) and Oasis MCX (150 mg/6 mL) cartridges were purchased from
135 Waters (Milford, MA, USA), while Strata NH₂ cartridges (200 mg/3 mL) were delivered by
136 Phenomenex (Torrance, California, USA). HPLC columns, used in this study, were
137 manufactured by Phenomenex. Chromatographic separation of the basic drugs was
138 achieved on Synergy 4 μ POLAR-RP 80 Å 150 x 3 mm column, while Kinetex 2.6 μ m PFP
139 100 Å 11 x 2.1 mm was used for the analyses of cannabinoids.

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141 2.3. Experimental setup

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143 Stability of the selected drugs in the sewer system was assessed using a series of
144 laboratory die-away experiments. The experiments were performed at 10 °C and 20 °C,
145 respectively, in order to simulate typical winter and summer in-sewer temperatures.
146 Wastewater samples (2L), collected from the main sewer system of the city of Zagreb at the
147 entrance to WWTP, were spiked with a mixture of all selected drugs at the environmentally
148 relevant concentrations of 200 ng/L of each individual compound, with the exception of
149 cannabinoid compounds, which were spiked at 1 µg/L. Municipal wastewater of the city of
150 Zagreb contains residues of all investigated drugs (Terzic et al., 2010) but some of them are
151 present at very low levels, so that for the purpose of this experiment, additional spiking of
152 original wastewater sample was necessary. The experiment was carried out at the original
153 pH of the wastewater (7.5). In addition, control experiments were performed at pH 2. The
154 samples for the control experiments were prepared by spiking 2 L of wastewater acidified
155 with H₃PO₄ with the same amounts of drugs. After spiking, both sets of wastewater
156 samples were well homogenised by shaking and divided into 7 identical aliquots of 250
157 mL. The aliquots were placed in the glass bottles (300 mL), and capped with cotton plugs.
158 The die-away experiments were performed in the dark using a thermostated cabinet. The
159 aliquots of the initial samples were processed immediately after spiking, while extraction of
160 other aliquots was performed in the time-intervals of 2, 4, 6, 24, 48 and 72 hours. Surrogate
161 standards were spiked to the filtered samples just prior to the extraction.

162 A separate experiment was designed to assess the stability of MOR and COD and to
163 explain the mechanisms responsible for the degradation of COC. In this experiment,
164 municipal wastewater of the city of Zagreb (pH 7.5) was spiked with an enhanced
165 concentration of MOR, COD and COC (4 µg/L of each) in order to minimise the potential
166 interfering effects of drug residues already present in the original wastewater (MG: 4.7
167 ng/L; 6-AM: 3.1 ng/L; 6-AC: 1 ng/L; BE: 125 ng/L), on the results. In this experiment, two
168 different types of control samples were prepared: the sample acidified to pH 2 (H₃PO₄) and
169 the sample poisoned with mercury chloride (HgCl₂; 50 mg/L). All prepared samples were
170 well homogenised by shaking, divided into aliquotes of 250 mL and processed in the
171 identical way as described above for the first two experiments.

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4 172 To evaluate the stability of the investigated compounds during the collection of 24-hour
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6 173 composite wastewater samples, 500 mL of raw wastewater (RW) and secondary effluent
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8 174 (SE) from WWTP of the city of Zagreb were spiked with 500 ng of each analyte. The
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10 175 spiked samples were homogenised and divided in 2 subsamples of 250 mL. One of each
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12 176 paralel sample was then acidified to pH 2 by the addition of H₃PO₄. Acidified and
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14 177 nonacidified samples were further divided in 40 mL aliquots, which were placed in HD
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16 178 polypropylene bottles and placed at the thermostated dark place at 4 °C. In this experiment,
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18 179 the samples were analysed at the beginning and at the end of 24-hour experimental period.
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20 180 All analyses in this experiment were conducted in duplicate.
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23 182 *2.4. Sample preparation and analyses* 24 25 183

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28 184 After selected time-intervals the samples were immediately filtered and prepared for the
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30 185 analyses. The sample preparation and liquid chromatography-tandem mass spectrometry
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32 186 (LC-MS-MS) analyses were performed identically as described for the dissolved phase
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34 187 analyses by Senta et al.(2013).

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36 188 Briefly, the samples were filtered through glass-fiber filters (Whatman, GF/D). The
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38 189 dissolved fraction was acidified to pH 2 and spiked with 30 ng of individual deuterated
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40 190 surrogate standards. The samples were enriched using preconditioned Oasis MCX
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42 191 cartridges. Before elution of adsorbed analytes, the cartridges were washed with 6 mL of
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44 192 ultrapure water and subsequently dried with N₂ (30 min). The elution of the enriched drugs
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46 193 was performed by 6 mL of MeOH (cannabinoid fraction) followed by 6 mL of 0.5% NH₃ in
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48 194 MeOH (basic drug fraction). Before the analysis, the cannabinoid fraction was additionally
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50 195 cleaned up using Strata NH₂ cartridges. The extracts were evaporated to dryness under N₂
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52 196 using a TurboVap evaporator (Caliper Life Sciences, Hopkinton, MA, USA) The dry
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54 197 residues of basic drug fraction and cannabinoid fraction were redissolved in 0.5 mL of
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56 198 H₂O/MeOH (8/2; v/v), containing 0.1% of acetic acid and 0.5 mL of H₂O/MeOH (3/7; v/v),
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58 199 respectively, and analysed by LC-MS-MS.
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200 All LC-MS-MS analyses were performed on a Thermo Electron HPLC system, equipped
201 with an autosampler (Surveyor, Thermo Electron, USA) and HPLC pump (MS Pump,
202 Thermo Electron, USA) interfaced to a triple-quadrupole mass spectrometer (Quantum
203 AM, Thermo Electron, USA), equipped with an electrospray ionisation source. The
204 chromatographic separation of basic drugs was achieved on Phenomenex Synergy 4 μ
205 POLAR-RP column (Phenomenex, 150 x 3 mm), while Kinetex PFP (Phenomenex, 100 x
206 2.1 mm) was used for the analyses of cannabionoid compounds.

207 The analysis of basic drugs was performed in positive ionisation polarity (PI), while
208 cannabinoids were analysed under negative ionisation conditions (NI). The capillary
209 voltage under PI and NI conditions were 3500 V and 3000 V, respectively. For both
210 ionisation modes, the capillary temperature was 350 °C. The desolvation (40 arbitrary
211 units) and auxiliary (10 arbitrary units) gas was N₂, while Ar was applied as a collision gas.
212 The collision energy and tube lens offset were optimised for each analyte and surrogate
213 separately. Identification and quantification was performed using two characteristic
214 transitions for the analysed compounds (MRM mode). Quantification of all analytes was
215 performed using corresponding deuterated internal standards.

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3. Results and discussion

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220 The stability of fourteen target compounds in municipal wastewater at typical summer (20
221 °C) and winter (10 °C) temperatures is presented in the Fig 1. Since no significant
222 differences were obtained between the control experiments (pH 2) performed at different
223 temperatures, for sake of clarity, the results from the control experiments at 10 °C were
224 omitted from the Fig 1. The measurements were fitted by die-away curves assuming
225 pseudo-first order kinetics as follows:

$$c = c_0 * e^{-kt} \quad (I)$$

227 were c and c₀ represent concentrations at times t and t₀, respectively, and k is the
228 degradation rate constant.

229 Consequently, the die-away half-life (t_{1/2}) was calculated from the

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$$t_{1/2} = \ln 2/k \quad (\text{II})$$

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8 232 Similar model of degradation kinetics was applied by Plosz et al.. (2013) in their study of
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10 233 cocaine stability. The stability of the individual compounds in our study highly varied
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12 234 depending on the compound type and the applied temperature conditions (Fig 1). It should
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14 235 be pointed out that in all control experiments drug biomarker concentrations were rather
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16 236 stable, indicating only negligible changes during the period of 72 hours in acidified
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18 237 samples. Die-away curves of amphetamine-type drugs (AMP, MAMP, MDMA),
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20 238 cannabinoid compounds (THC-COOH, THC-OH) as well as of MTHD and EDDP were not
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22 239 significantly different from the stability curves for the control experiments (pH 2). In fact,
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24 240 the estimated degradation half-lives of these drug biomarkers were rather long ($\gg 200$ h)
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26 241 at both temperatures examined, which indicated that these compounds could be considered
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28 242 virtually stable in a typical sewer system. This is in a good agreement with the results
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30 243 reported by van Nuijs at al. (2012). In contrast to amphetamines, cannabinoids and
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32 244 therapeutic opioids, significant changes were observed for MG, 6-AM, 6-AC, COC, BE,
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34 245 MOR and COD. The die away curves for MG, 6-AM, 6-AC and COC followed the first
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36 246 order kinetics with pronounced temperature dependence (Table 1). The $t_{1/2}$ of MG, 6-AM,
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38 247 6-AC and COC at 20 °C was 7 h, 77 h, 58 h and 35 h, respectively, while the corresponding
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40 248 $t_{1/2}$ values at 10 °C were 18 h, 139 h, 87 h and 173 h, respectively. These results indicated a
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42 249 pronounced seasonal variability of the MG, 6-AM, 6-AC and COC stability in the sewer.
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44 250 However, the significance of the seasonal differences in drug stability on the reliability of
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46 251 drug consumption estimates strongly depends on in-sewer hydraulic retention time. In the
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48 252 sewer systems, like the one in the city of Zagreb (Croatia), having relatively short average
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50 253 residence times (4 hours), the seasonal impact on in-sewer losses becomes significant only
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52 254 for MG (33% and 16%, in summer and winter, respectively). Nevertheless, for the systems
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54 255 having retention times longer than 12 h, the seasonal differences would become more
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56 256 prominent for all four compounds.

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58 257 As opposed to MG, 6-AM, 6-AC and COC, the stability curves of BE, MOR and COD
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60 258 showed an increasing trend indicating that significant transformations of COC to BE, 6-AM
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62 259 and MG to MOR as well as of 6-AC to COD. Obviously, these transformation processes

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260 were faster than the possible degradation of BE, MOR and COD themselves. Analogous
261 behaviour of 6-AM and 6-AC further indicated that the hydrolysis of acetyl group on the 6
262 position, was probably the common transformation mechanism of these compounds in the
263 wastewater. On the other hand, MG was most probably transformed to MOR by
264 glucuronidase enzymes of the bacteria present in the wastewater (e.g. Ternes 1998). This
265 transformation was efficiently prevented in the control experiment by lowering pH to 2.

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3.1. Stability of cocaine biomarkers and their impact on cocaine consumption estimates

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270 Stability of COC and its potential transformation to BE is an important issue in sewage
271 epidemiology since BE, as the main human metabolite of COC, has been most frequently
272 used for the estimation of COC consumption (e.g. Postigo et al., 2010; Terzic et al., 2010;
273 Thomas et al., 2012; van Nuijs et al., 2011; Zuccato et al., 2005, 2008). Several reports
274 from the literature suggested that COC could be partially transformed to BE during the
275 passage through the sewer system (for example Gheorghe et al., 2008; van Nuijs et al.,
276 2012). The results from our study are in a full agreement with these findings (Fig 1). In
277 order to clarify the mechanisms leading to the degradation of COC, an additional
278 experiment, in which the wastewater sample was spiked with enhanced concentration of
279 COC (4 µg/l) was performed (see Experimental part for the details). To calculate the
280 amount of newly-formed BE, the initial BE concentration was subtracted from the BE
281 concentrations measured for each sampling point. The results of this experiment are
282 presented in Table 2 and Fig S1-A. The die-away curve of COC (Fig S1-A) was rather
283 similar to the corresponding curve obtained in the first experiment (Fig 1). Furthermore, the
284 degradation curves of COC in the non-preserved sample (pH 7.5) and the control sample
285 preserved with mercury chloride to prevent biological activity (pH 7.5; HgCl₂) were quite
286 similar (see Fig S1-A) and showed gradual decrease of COC. This suggested that the main
287 mechanism governing the degradation of COC in our experiments was not biodegradation.
288 On the contrary, the concentration of COC in the sample acidified to pH 2 was virtually
289 stable during the whole experiment, which indicated that COC degradation was caused

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290 almost exclusively by chemical hydrolysis. In contrast, recent study by Plosz et al. (2013)
291 reported on a significant biodegradability of COC and BE at 21 °C, both under aerobic and
292 anaerobic conditions. This discrepancy possibly suggests that biodegradability of COC
293 depends on specific conditions in wastewater, including the composition and pre-adaptation
294 of the present microbial consortium, as well as the total heterotrophic biomass. These
295 authors concluded that the biotransformation of COC must be taken into account when
296 estimating the COC consumption based on BE determination, especially during the festival
297 periods, characterised by enhanced relative concentrations of COC. On the other hand, van
298 Nuijs et al.. (2012) suggested that the influence of formation of BE in sewage epidemiology
299 back-calculations was supposed to be low, even for in-sewer residence times longer than 12
300 hours. Therefore, further research is needed to clarify these issues.

301 As to the transformation products formed, our experiment suggested that COC was
302 transformed almost exclusively into BE (Table 2). The relative increase in BE
303 concentration, estimated from our experiments at summer conditions and considering
304 BE/COC ratio of 1.2 and the hydraulic wastewater retention time of 2-12 hours, was 1 to
305 6%. However, in the real wastewater, which is generally characterised by higher BE/COC
306 concentration ratio (3.3 ± 0.2 ; Terzic et al.. 2010) than in our spiked samples, the impact of
307 COC hydrolysis on the BE levels would be even lower than the one estimated above.

308 Besides the stability of COC, potential instability of BE might have an important impact on
309 the accuracy of the COC consumption estimates. The die-away curves of BE represent a
310 combined result of its formation and possible further degradation. Nevertheless, the
311 accumulation of BE (Table 2, Fig 1), indicated its relatively high stability in the wastewater
312 for at least 72 hours. Two experiments showed virtually quantitative transformation of
313 COC into BE, while the mass balance analysis of one of the experiments, conducted at 20
314 °C (Table 2) showed that the increase of BE after 72 hours was 19% lower than the amount
315 of transformed COC. This indicated either simultaneous formation of other COC
316 transformation products or further transformation of BE. Plosz et al.. (2013) showed that
317 BE could be biotransformed in the sewer, however biotransformation was much slower
318 than its in-sewer formation from COC.

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4 320 3.2. *Stability of heroin biomarkers and their impact on heroin consumption estimates*
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8 322 As can be seen in Fig. 1, clear decreasing temporal trends of MG and 6-AM were followed
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10 323 by a concomitant increase in MOR concentration. This suggested that the transformation of
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12 324 MG and 6-AM to MOR was faster than MOR degradation (Fig 1). In fact, additional
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14 325 experiment, which was performed with wastewater sample spiked solely with MOR,
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16 326 showed that MOR itself was relatively stable in the wastewater over the entire investigated
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18 327 time-period of 72 h (Fig S1). However, a detailed mass balance analysis of the experiments
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20 328 (Table 2) showed that the loss of MG and 6-AM was slightly greater than the amount of
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22 329 newly formed MOR. After 72 hours the difference was equivalent to about 14-16% of the
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24 330 theoretically expected concentration of MOR, which might be due to its further
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26 331 biotransformation. However, UPLC-QTOF screening of the extracts by UPLC/Q-TOF MS
27
28 332 did not confirm any detectable concentrations of known MOR transformation products
29
30 333 (Wick et al., 2011).

31
32 334 The transformation of both MG and 6-AM was efficiently prevented by the acidification
33
34 335 (pH 2) of the wastewater sample, suggesting enzymatic hydrolysis as the main mechanism
35
36 336 for both biomarkers.

37
38 337 MG is one of the major heroin metabolites (Baselt, 2008). However, it is generally assumed
39
40 338 that such conjugate compounds are readily re-transformed to the parent compounds in the
41
42 339 municipal wastewater due to the presence of β -glucuronidase enzymes of the fecal bacteria
43
44 340 (e.g. Ternes, 1998). However, kinetic models derived from our experiments showed that,
45
46 341 assuming the hydraulic sewer residence time of 2 and 12 h, approximately 63 to 92% of
47
48 342 non-transformed MG would remain in the sewer at winter temperature conditions. In
49
50 343 summer, these percentages decrease to 30 and 82%, but remain significant. The remaining
51
52 344 MG could be interpreted as one of the factors leading to underestimation of heroin if the
53
54 345 estimation is based on morphine measurements. Further deconjugation of MG would
55
56 346 probably occur during the composite sample collection and sample preparation.
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58 347 Nevertheless, our results suggest that MG should be measured and summed up with the
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60 348 corresponding MOR concentration in order to avoid underestimation of heroin
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62 349 consumption based on MOR measurements.

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350 The alternative way to estimate heroin consumption is based on 6-AM measurements.
351 Unlike MG and MOR, 6-AM is a minor, but exclusive metabolite of heroin. However, the
352 reliability of the heroin estimation strongly depends on 6-AM stability in the wastewater.
353 According to our results, the residual concentration of 6-AM after 2 to 12 h in the sewer
354 would be 98 to 91% and 99 to 94% of its initial concentration, assuming summer and
355 winter temperature conditions, respectively. Consequently, the errors in estimations of the
356 heroin consumption using 6-AM can become significant only at higher sewer hydraulic
357 retention times (>12 h) under summer temperature conditions.

358

359 *3.3. Stability of the drug biomarkers during the collection of 24-h composite samples*

360

361 Besides the stability in the sewer, sampling over a prolonged period is also a potential
362 source of error. In this study, we examined the stability of drug biomarkers during the
363 collection of the 24-hour composite samples. The experiment was performed at 4 °C, since
364 this is a typical temperature applied during the sample collection in automatic samplers.
365 The results obtained for both raw wastewater (RW) and secondary effluent (SE) are
366 presented in the Fig. 2. Most of the investigated compounds exhibited rather high stability
367 at the applied experimental conditions in both matrices. For most of the compounds the
368 concentration changes after 24 hours were within the error margins of the analytical method
369 and cannot be considered significant. The compound losses for the samples kept at original
370 pH (Fig. 2) were similar to the losses in the control samples kept at pH 2 (Fig. S2). A
371 significant difference between the two sample types was obtained for THC-COOH, which
372 residual percentage was much lower in the control samples (46% at pH 2) than in the non-
373 acidified samples (90% at pH 7.4). This result indicates most probably the enhanced
374 adsorption of THC-COOH at pH 2 as compared to the environmental pH. Namely,
375 according to Khan and Nicell (2012) only 1.3% of THC-COOH is expected to be adsorbed
376 on sewer-borne solids at environmental pH conditions (pH ≈ 7.5), while its adsorption at
377 pH 2 was estimated to be much higher (56.3%).

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380 **4. Conclusion**

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382 Most of the illicit drug biomarkers examined in this study, including 6 illicit drugs and 2
383 therapeutic opioids, proved to be rather stable in model experiments, simulating in-sewer
384 degradation. The most unstable biomarkers were MG, 6-AM, 6-AC and COC and the errors
385 associated with their changes should be carefully taken into account when estimating the
386 collective drug consumption. Their degradation followed pseudo-first order kinetics that
387 was much faster at summer (20 °C) than at winter temperatures (10 °C). COC degradation
388 was caused predominately by chemical hydrolysis to BE. However it was estimated that
389 this process would significantly affect the accuracy of BE-based COC consumption
390 estimates only if the in-sewer hydraulic retention time is very long (> 12 h) and/or when
391 COC/BE ratio is unusually high.

392 The heroin consumption estimates, based on MOR measurements, could be significantly
393 underestimated if MG is not measured and summed-up with MOR measurements. This
394 would be more pronounced at winter temperature conditions due to the much slower
395 transformation of MG to MOR. On the other hand, the heroin consumption estimates based
396 on 6-AM measurements are less prone to the errors due to the in-sewer transformations and
397 could become significant only at very high hydraulic retention times (>12 hours).

398 Acidification proved to be a good way to stabilise wastewater samples for the analysis of
399 the drug biomarkers, except for THC-COOH, most probably due to the adsorption losses.
400 This finding should be taken into account when selecting the preservation technique for
401 multiresidual analyses of different groups of illicit drugs.

402

403

404 **Acknowledgments**

405 This work was funded by Croatian Ministry of Science, Education and Sports (Project No.
406 098-982934-2712). We are grateful to the staff of the central wastewater treatment plants of
407 the city of Zagreb for helping with wastewater sampling and providing the data about
408 WWTPs. A technical help of Nenad Muhin is also highly appreciated.

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410 **References**

411

412 Baker DR, Kasprzyk-Hordern, B. Critical evaluation of methodology commonly used in
413 sample collection, storage and preparation for the analysis of pharmaceuticals and illicit
414 drugs in surface water and wastewater by solid phase extraction and liquid
415 chromatography–mass spectrometry. *J Chromatogr A* 2011; 1218: 8036-59.

416 Baselt RC. 2008. Disposition of toxic drugs and chemicals in man, 8th ed. Biomedical
417 Publications, Foster City, California, USA, p. 730-1.

418 Castiglioni S, Zuccato E, Crisci E, Chiabrando C, Fanelli R, Bagnati R. Identification and
419 measurement of illicit drugs and their metabolites in urban wastewater by liquid
420 chromatography–tandem mass spectrometry. *Anal Chem* 2006; 78: 8421-9.

421 Castiglioni S, Bagnati R, Melis M, Panawennage D, Chiarelli P, Fanelli R, Zuccato E.
422 Identification of cocaine and its metabolites in urban wastewater and comparison with the
423 human excretion profile in urine. *Water Res* 2011; 45: 5141-50.

424 Castiglioni S, Bijlsma , Reid M, Ort C, Thomas KV, van
425 Nuijs ALN, de Voogt P, Zuccato E. Evaluation of Uncertainties Associated with the
426 Determination of Community Drug Use through the Measurement of Sewage Drug
427 Biomarkers. *Environ Sci Technol* 2013; 47: 1452-60.

428 Chiaia AC, Banta-Green C, Field J. Eliminating solid phase extraction with large-volume
429 injection LC/MS/MS: analysis of illicit and legal drugs and human urine indicators in US
430 wastewaters. *Environ Sci Technol* 2008; 42: 8841-8.

431 Gheorghe A, van Nuijs A, Pecceu B, Bervoets L, Jorens PG, Blust R, Neels H, Covaci A.
432 Analysis of cocaine and its principal metabolites in waste and surface water using solid-
433 phase extraction and liquid chromatography–ion trap tandem mass spectrometry. *Anal*
434 *Bioanal Chem* 2008; 391: 1309-19.

435 Gonzalez-Marino I, Quintana JB, Rodriguez I, Cela R. Determination of drugs of abuse in
436 water by solid-phase extraction, derivatisation and gas chromatography–ion trap-tandem
437 mass spectrometry. *J Chromatogr A* 2010; 1217: 1748–60.

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438 Karolak S, Nefau T, Bailly E, Solgadi A, Levi Y. Estimation of illicit drugs consumption
439 by wastewater analysis in Paris area (France). *Forensic Sci Int* 2010; 200: 153-60.

440 Kasprzyk-Hordern B, Dinsdale RM, Guwy AJ. Illicit drugs and pharmaceuticals in the
441 environment - Forensic applications of environmental data. Part1: Estimation of the usage
442 of drugs in local communities. *Environmental Pollut* 2009; 157: 1773-7.

443 Khan U, Nicell JA. Sewer epidemiology mass balances for assessing the illicit use of
444 metamphetamine, amphetamine and tetrahydrocannabinol. *Sci Tot Environ* 2012; 421: 144-
445 62.

446 Plosz BG, Reid MJ, Borup M, Langford KH, Thomas KV. Biotransformation kinetics and
447 sorption of cocaine and its metabolites and the factors influencing their estimation in
448 wastewater. *Water Res* 2013; 47: 2129-40.

449 Postigo C, Lopez de Alda MJ, Barcelo D. Drugs of abuse and their metabolites in the Ebro
450 River basin: Occurrence in sewage and surface water, sewage treatment plants removal
451 efficiency, and collective drug usage estimation. *Environ Int* 2010; 36: 75-84.

452 Senta I, Krizman I, Ahel M, Terzić S. Integrated procedure for multiresidue analysis of
453 dissolved and particulate drugs in municipal wastewater by liquid chromatography -
454 tandem mass spectrometry. *Anal Bioanal Chem* 2013; 405: 3255-68.

455 Ternes T. Occurrence of drugs in German sewage treatment plants and rivers. *Water Res*
456 1998; 32: 3245-60.

457 Terzic S, Senta I, Ahel M. Illicit drugs in wastewater of the city of Zagreb (Croatia)-
458 Estimation of drug abuse in a transition country. *Environ Pollut* 2010; 158: 2686-93.

459 Thomas KV, Bijlsma L, Castiglioni S, Covaci A, Emke E, Grabic R, Hernandez F,
460 Karolak, S, Kasprzyk-Hordern B, Lindberg RH, Lopez de Alda M, Meierjohann A, Ort C,
461 Pico Y, Quintana JB, Reid M, Rieckermann J, Terzic S, van Nuijs ALN, de Voogt P.
462 Comparing illicit drug use in 19 European cities through sewage analysis. *Sci Total*
463 *Environ* 2012; 432: 432-9.

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464 Van Nuijs ALN, Mougel JF, Tarcomnicu I, Bervoets L, Blust R, Jorens PG, Neels H,
465 Covaci A. Sewage epidemiology-A real-time approach to estimate the consumption of
466 illicit drugs in Brussels, Belgium. *Environ Int* 2011; 37: 612-21.

467 Van Nuijs ALN, Abdellati K, Bervoets L, Blust R, Jorens PG, Neels H, Covaci A. The
468 stability of illicit drugs and metabolites in wastewater, in important issue for sewage
469 epidemiology? *J Hazard Mater* 2012; 239-240: 19-23.

470 Wick A, Wagner M, Ternes TA. Elucidation of the Transformation Pathway of the Opium
471 Alkaloid Codeine in Biological Wastewater Treatment. *Environ Sci Technol* 2011; 45:
472 3374-85.

473 Zuccato E, Chiabrando C, Castiglioni S, Calamari D, Bagnati R, Schiarea S. Cocaine in
474 surface waters: a new evidence-based tool to monitor community drug abuse. *Environ*
475 *Health: A Global Access Science Source* 2005; 4: 14-20.

476 Zuccato E, Chiabrando C, Castiglioni S, Bagnati R, Fanelli R. Estimating Community Drug
477 Abuse by Wastewater Analysis. *Environ Health Persp* 2008; 116: 1027-32.

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480 **Figure Captions**

481

482 Figure 1. Stability of illicit drugs and their metabolites in the wastewater at winter (10 °C)
483 and summer (20 °C) sewer temperature conditions. (COC-cocaine; BE-benzoylecgonine;
484 MOR-morphine; 6-AM-6-acetylmorphine; MG- morphine-3-β-D glucuronide; 6-AC- 6-
485 acetyl codeine; MDMA-3,4-methylenedioxymethamphetamine; AMP-amphetamine;
486 MAMP- methamphetamine; MTHD-methadone; EDDP-2-ethylidene-1,5-dimethyl-3,3-
487 diphenylpyrrolidine; THC-COOH-11-nor-9-carboxy-Δ⁹-tetrahydrocannabinol; THC-OH-
488 11-hydroxy-Δ⁹-tetrahydrocannabinol)

489

490 Figure 2. Twenty-four-hour stability of different types of urinary biomarkers in the raw
491 wastewater (RW) and secondary effluent (SE) at 4 °C. (COC-cocaine; BE-
492 benzoylecgonine; MOR-morphine; 6-AM-6-acetylmorphine; MG- morphine-3-β-D
493 glucuronide; MDMA-3,4-methylenedioxymethamphetamine; AMP-amphetamine; MAMP-
494 methamphetamine; MTHD-methadone; EDDP-2-ethylidene-1,5-dimethyl-3,3-
495 diphenylpyrrolidine; THC-COOH-11-nor-9-carboxy-Δ⁹-tetrahydrocannabinol; THC-OH-
496 11-hydroxy-Δ⁹-tetrahydrocannabinol; COD-codeine)

Table 1[Click here to download Table: Table 1.docx](#)**Table 1.** Kinetic parameters determined for degradable compounds at two different temperature conditions.

	10 °C; pH 7.5				20 °C; pH 7.5			
	equation	r ²	k (h ⁻¹)	t _{1/2}	equation	r ²	k (h ⁻¹)	t _{1/2}
COC	$y = e^{-0.004x}$	0.85010	0.004	173	$y = e^{-0.02x}$	0.9897	0.020	35
6-AM	$y = e^{-0.005x}$	0.8695	0.005	139	$y = e^{-0.008x}$	0.9540	0.008	87
MG	$y = e^{-0.039x}$	0.9899	0.039	18	$y = e^{-0.1x}$	0.9335	0.100	7
6-AC	$y = e^{-0.008x}$	0.9587	0.008	87	$y = e^{-0.012x}$	0.9880	0.012	58

COC-cocaine; 6-AM-6-acetyl morphine; MG- morphine-3-β-D glucuronide; 6-AC-6-acetyl codeine

Table 2[Click here to download Table: Table 2.docx](#)

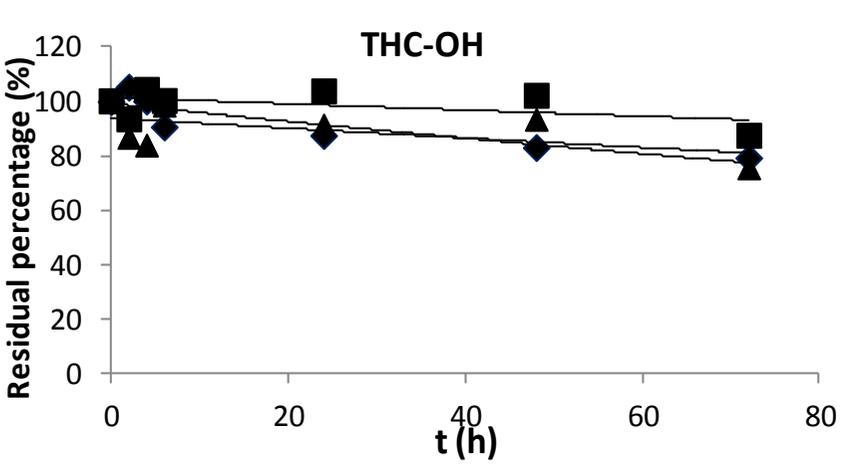
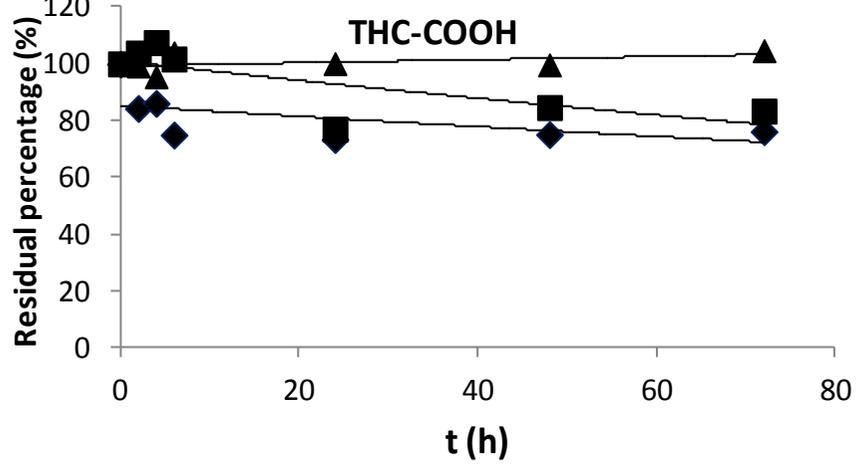
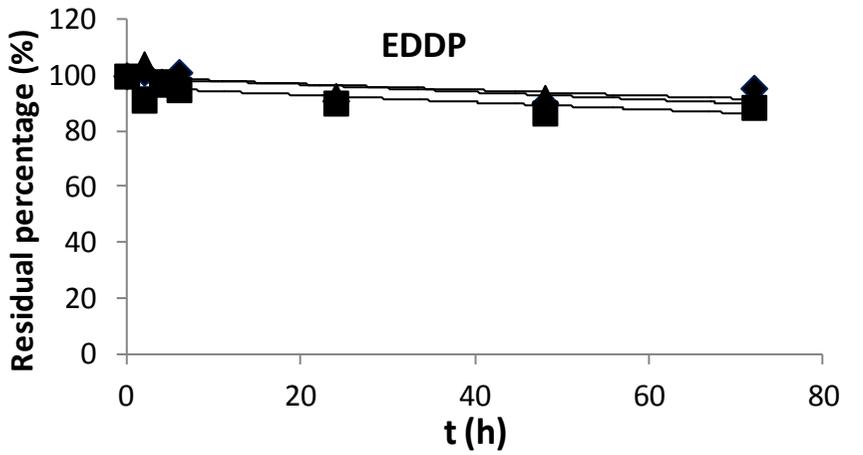
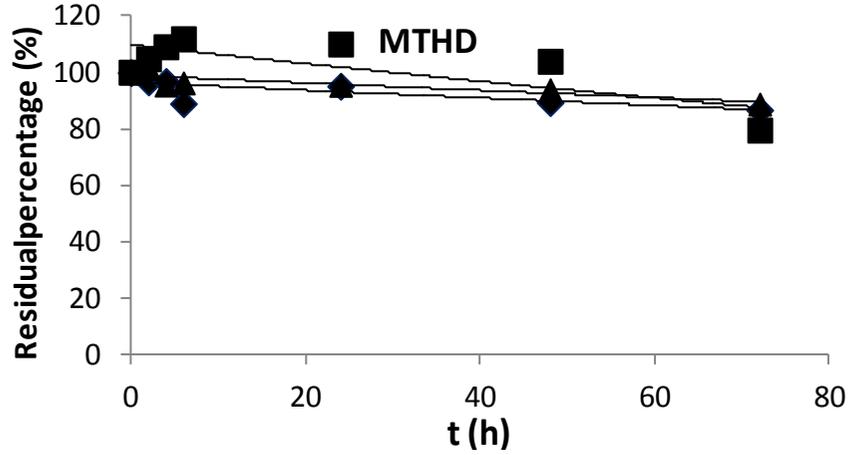
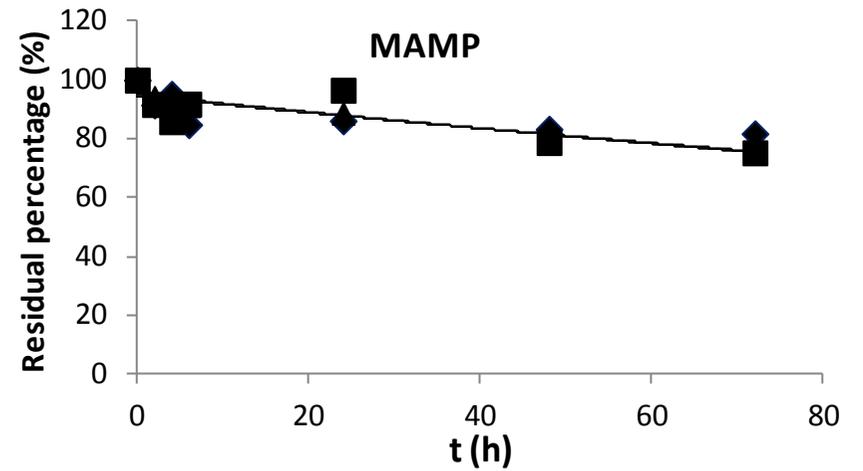
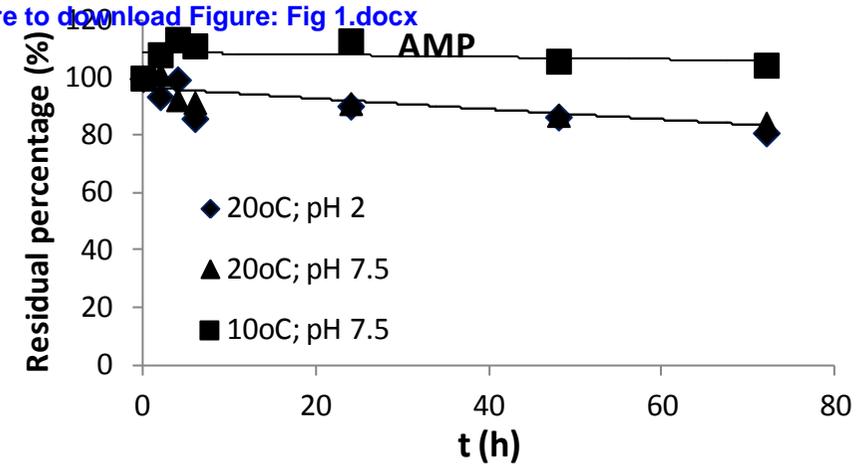
Table 2. Mass balance of cocaine (COC), benzoylecgonine (BE), morphine-3- β -D glucuronide (MG), 6-acetyl morphine (6-AM) and morphine (MOR) at the end of the performed 72-hour die-away experiments.

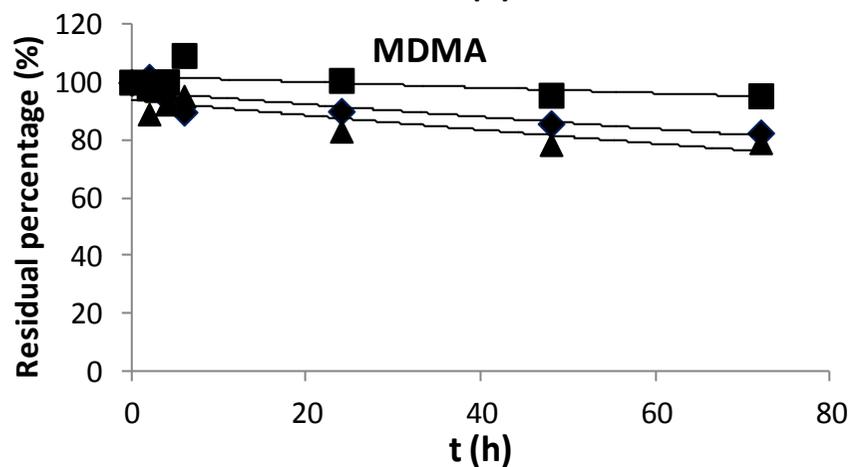
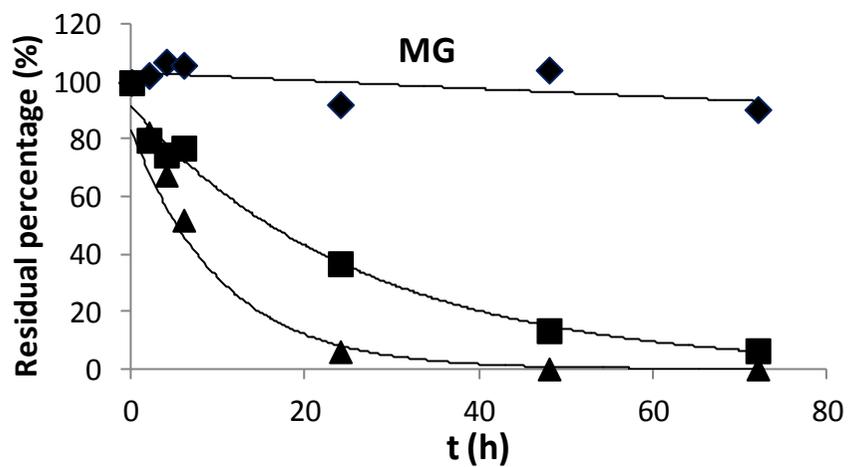
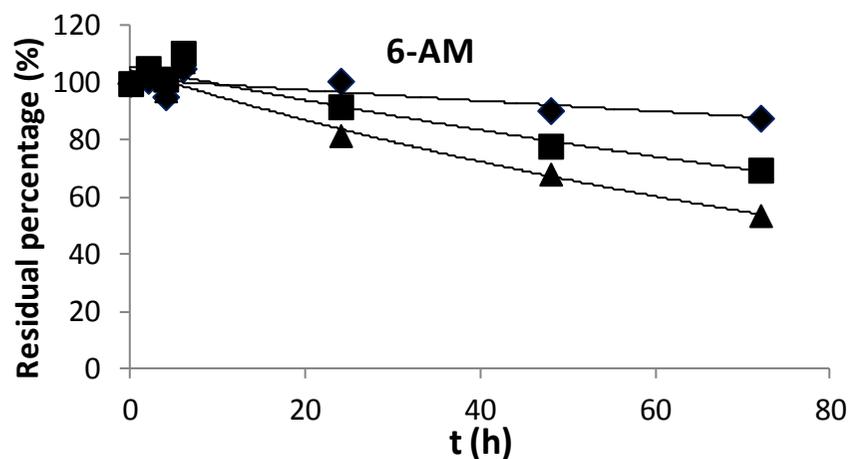
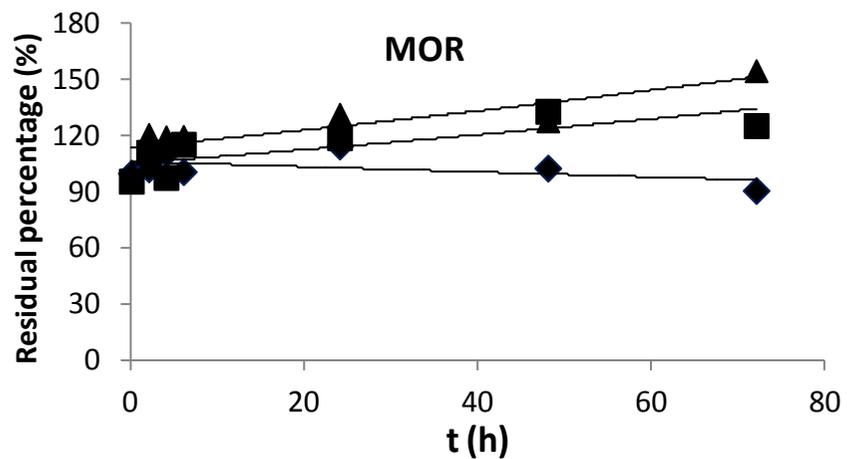
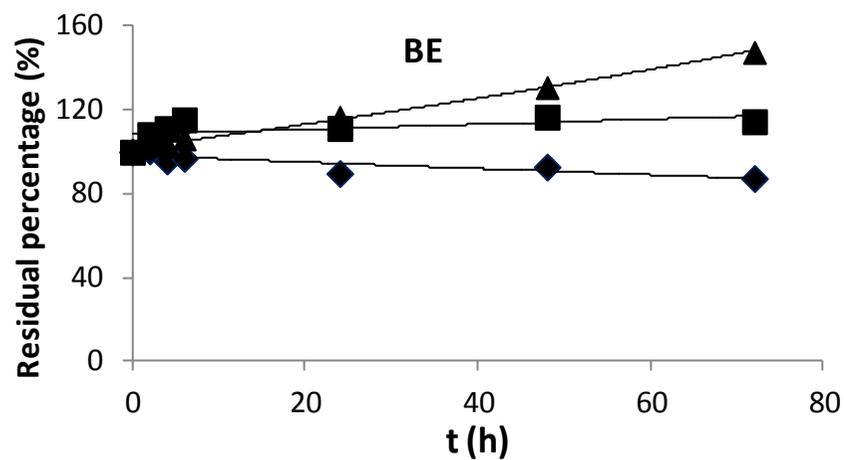
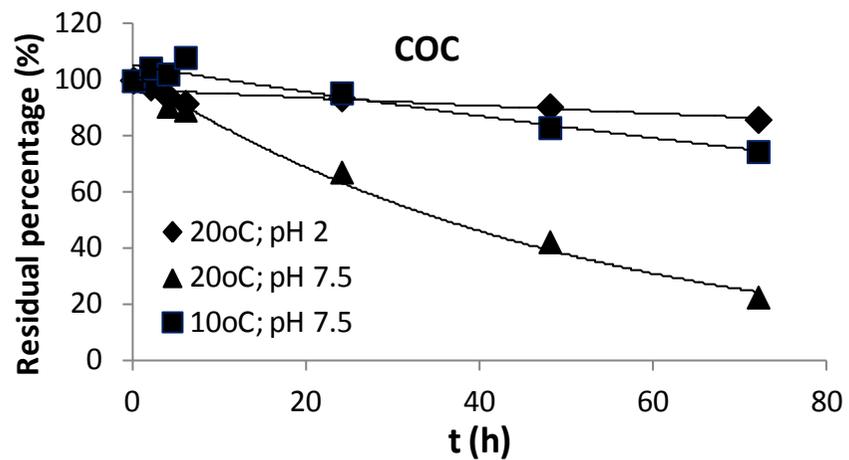
	Δc_{COC} (nmol/L)	Δc_{BE} (nmol/L)	$\Delta c_{\text{MG}} + \Delta c_{\text{6-AM}}$ (nmol/L)	Δc_{MOR} (nmol/L)
10 °C, pH 7.5; 0.2 $\mu\text{g/L}^*$	-0,18	+0.18	-0.51	+0.28
20 °C, pH 7.5; 0.2 $\mu\text{g/L}^*$	-0,62	+0.50	-0.70	+0.48
20 °C, pH 7.5; 4 $\mu\text{g/L}^*$	-7.74	+7.80	NA	NA
20 °C, pH 7.5; HgCl_2 , 4 $\mu\text{g/L}^*$	-7.05	+6.85	NA	NA
20 °C, pH 2; 4 $\mu\text{g/L}^*$	+0.09	0.03	NA	NA

NA-not applicable; * spiking level (note that a real initial concentration of individual compounds was a sum of the spiked concentration and the concentration of each biomarker already present in the wastewater sample); Δc represents the difference between the final and initial concentration (e.g.

$$\Delta c_{\text{MOR}} = c(\text{MOR}_{t72\text{h}}) - c(\text{MOR}_{t0})$$

Figure 1
[Click here to download Figure: Fig 1.docx](#)





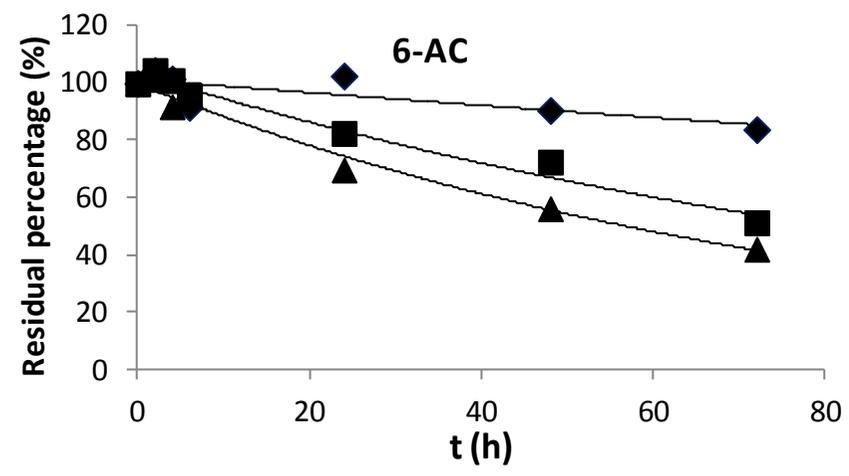
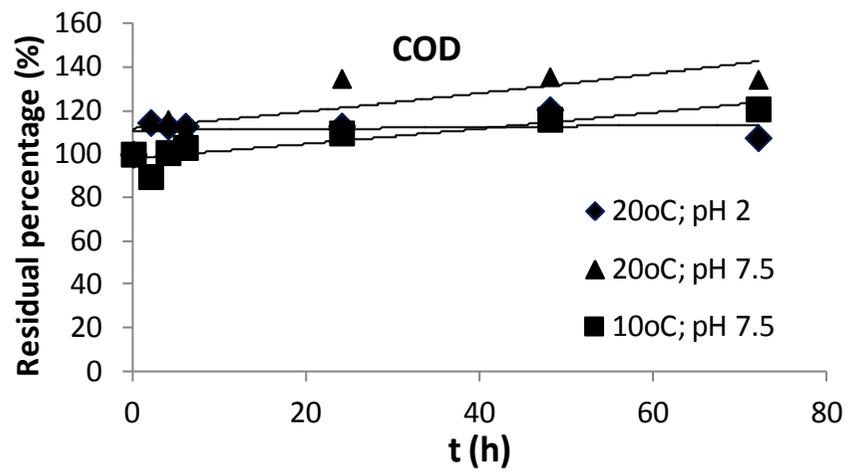


Fig. 1

Figure 2
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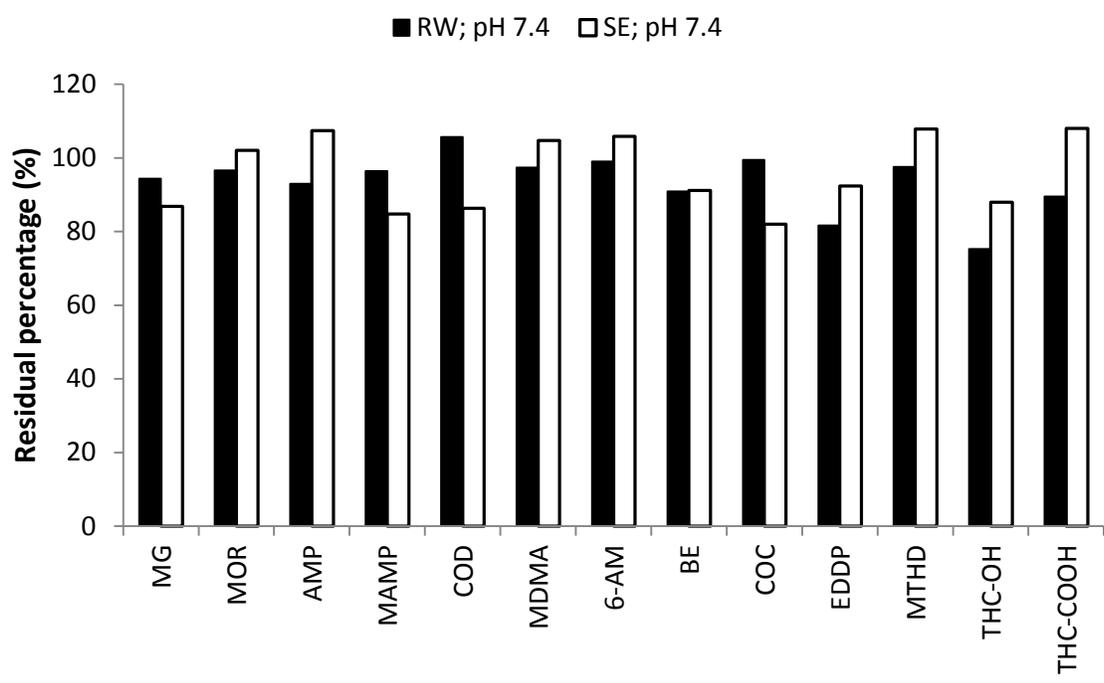


Figure 2

Supplementary Material

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