Is the digestive gland of <u>Mytilus galloprovincialis</u> a tissue of choice for estimating cadmium exposure by means of metallothioneins?

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Abstract

A study performed over 12 months with caged mussels <u>M. galloprovincialis</u> in the coastal marine zone which is under the urban pressure, reveales a temporal variation of digestive gland mass which causes "biological dilution" of cytosolic metallothionein (MT) and trace metal (Cd, Cu, Zn, Fe, Mn) concentrations. The dilution effect was corrected by expressing the cytosolic MT and metal concentrations as the tissue content. Consequently, the changes of the average digestive gland mass coincide with the changes of MT and trace metal contents. In the period from February to June, MT contents are nearly twice and trace metal contents nearly three times higher than the remaining months. The period of increased average digestive gland mass, of MT and trace metal contents is probably overlapping with the sexual maturation of mussels (gametogenesis) and the enhanced food availability. Due to the fact that natural factors contribute more to the MT content than the sublethal levels of Cd, digestive gland of <u>M</u>. galloprovincialis is not considered as a tissue of choice for estimating Cd exposure by means of MTs.

<u>Key words</u>: caged bivalves, <u>Mytilus galloprovincialis</u>; digestive gland mass variation; tissue content of metallothioneins and trace metals

Introduction

Coastal seawater is often under the urban and industrial pressure due to metal inputs. Metals are persistent and some are bioaccumulated in marine organisms. Filter-feeding bivalves are abundant in coastal waters and thus often used for monitoring trace metal burden as the consequence of anthropogenic activities (Giusti et al., 1999). By means of specific, inducible proteins, such as metallothioneins (MTs), heavy metal homeostasis and detoxification in marine invertebrates occurs (Viarengo and Nott, 1993). Induction of MTs is regarded as a biochemical response to metal exposure, but biotic and abiotic factors additionally influence MT levels and have to be considered in the assessment of metal exposure (Langston et al., 1998). The MT response to metals is tissue specific. As main metabolic and storage tissue for metals, digestive gland of bivalves has high basal MT level, which probably reflects high tissue content of Cu and Zn (Geffard et al., 2001). Within the framework of the Mediterranean Action Plan (UNEP/MAP, 1999) digestive gland of Mytilus galloprovincialis has been selected as target tissue for survey of metal exposure by means of MTs as a specific biomarker. While controlled exposure experiments with Cd clearly indicate de novo synthesis of MT proteins in the digestive gland of mussels (Raspor et al., 1987; Bebianno and Langston, 1991), the field experiments with oysters do not clearly differentiate MT levels in the digestive gland at metal-rich and the control sites (Geffard et al., 2001).

The aim of our study was to conduct field experiment over one year with the defined stock of mussels <u>M. galloprovincialis</u>, transplanted from an aquaculture area to 4 different sites in the part of the Adriatic Sea which is under urban pressure (Kaštela Bay, Eastern Adriatic Sea). Biochemical (MT) and chemical (metals) parameters were

determined in the heat-treated cytosol of the composite sample of mussel digestive glands of caged organisms. The study is based on the same stock of mussels, of the same size, age and origin, which is the prerequisite to optimize resolution power of chemical stress at different locations (De Kock and Kramer, 1994). Such set-up enabled us to follow up temporal fluctuations of digestive gland mass and consequently cytosolic MT and metal levels with the purpose to clarify if the digestive gland of <u>M</u>. <u>galloprovincialis</u> is a tissue of choice for estimating cadmium exposure by means of MTs.

Materials and Methods

Mussel caging and sampling

Filter-feeding <u>M. galloprovincialis</u> of defined length $(5.1\pm0.2 \text{ cm})$ and age $(12\pm1 \text{ months})$ were transplanted from an aquaculture area and deployed at 4 different sites within Kaštela Bay (Fig. 1, sites A to D), a recipient of most of urban and industrial wastewater of the area. Deployment sites were selected taking into account the hydrographic, chemical and biological characteristics of the Bay (Kušpilić et al., 1991; Barić et al., 1992; Beg-Paklar and Gačić, 1997). Within the Bay prevailing anticlockwise circulation of water masses exists and their average retention time amounts to 30 days. According to numerous indicators, including primary production assessment, the Kaštela Bay could be categorized as a highly eutrophic area (Barić et al., 1992). At 200 to 500 m from the shore (site B) the average annual organic matter content in the suspended particles amounts to 10% (Tudor, 1993). In this shallow area seasonal variability of the suspended particles concentration and resuspension of sediment particles of high organic matter content (>20%) contribute to the metal content uptake

by the filter feeding organisms, like mussels. Mass partition of metals (expressed on dry mass basis) in the fine-grained fraction of the surface marine sediments within the studied area amounts to 100 mg kg⁻¹ for Zn, 40 mg kg⁻¹ for Cu and 0.4 mg kg⁻¹ for Cd (Bogner et al., 1998). The decrease of Zn and Cu mass partition with sediment depth indicates to the anthropogenic sources of these metals.

At the deployment sites the water depth was 10 m, while the distance from the shore varied from 50 to 400 m. At each caging site eight net-like baskets (each one containing 50 specimens) were deployed 1.5 m above the sea bottom on September 15th, 1997; one basket was sampled at a time. Monthly sampling was performed in October, November and December 1997, to detect any changes which might reflect the acclimation of the Mediterranean mussels to new ambient conditions. In 1998 bimonthly sampling was performed (February, April, June, August). The last one took place in September 1998 to end up the round. Within current month, sampling took place on the same date as the deployment date. At stations A and B sampling was complete (8 times), while due to rough weather at site C one sampling and at site D two samplings were not accomplished. Therefore, data at these two locations in particular periods are missing.

After sampling, mussels were kept for 24 hours in the filtered (0.45 μ m) seawater to depurate the gut content (Bordin et al., 1992; Odžak et al., 2001), and than transported to the Laboratory in Zagreb, where biometry and tissue dissection took place. Composite samples of mussel tissues were deep frozen at -80°C until further processing and analysis.

Mussel biometry

At each caging site and sampling period the composite sample of the digestive glands was made up on average from 28 specimens. Therefore, no replicates per sampling period and site exist, but results represent the average biochemical and chemical responses. Average digestive gland mass within composite tissue sample was determined in order to observe temporal fluctuations of that tissue, mainly related to reproductive cycle and food availability (Bordin et al., 1997). At the beginning of our field study the mussels' age was defined as 12 ± 1 months. Shell mass was selected as additional indicator of age, due to the fact that the calcareous shell continues to be formed even when the length increments are not observed (Fischer, 1983). Our observations confirm that during 12 months of caging, mussels' length increased on average from 5.2 to 5.9 cm, while the shell mass increased on average from 4.5 to 7.0 g.

Homogenate and cytosolic fraction

Composite digestive gland tissue was homogenized in three volumes of 0.02 M TRIScontaining leupeptine HCl buffer. pH=8.6, (0.006)mM). phenylmethylsulphonylfluoride (PMSF, 0.5 mM) and 2-mercaptoethanol (0.01%), on an ice-bath with a Potter-Elvehjem type of homogenizer. The homogenate was centrifuged in the Sorval RC28S centrifuge by Du Pont at 30000xg for 40 minutes at 4°C. The isolated supernatant (S30) contained total cytosolic proteins (Yang et al., 1995). Further on, S30 was heat-treated at 70°C for 10 minutes using The Dri Block (Techne), and subsequently centrifuged at 30000xg for 20 minutes at 4°C. This supernatant contained MTs as heat-stable cytosolic proteins (Yang et al., 1995).

Cytosolic MT concentration

MT concentration in the heat-treated supernatant (mg ml⁻¹) was determined by electrochemical method in a differential pulse mode (Raspor et al., 2001), on a Metrohm 290E hanging mercury drop electrode (HMDE). MT calibration straight line was obtained at 7°C with the commercial rabbit liver MT(I+II) from Sigma.

Heat-denatured cytosolic trace metal concentrations

Metal concentrations in the heat-treated S30 fraction (μ g ml⁻¹) were determined by means of Varian double beam flame atomic absorption spectrometer (SpectrAA 220) with multielement lamps and a deuterium lamp for baseline correction. Atomization of metals was achieved in the air-acetylene flame. Calibration was performed using Merck's standard solutions of Cd, Zn, Cu, Mn and Fe. Selected concentration ranges of Cd 0.01-0.05 μ g ml⁻¹; Zn 0.5-2.0 μ g ml⁻¹; Cu 0.1-0.5 μ g ml⁻¹; Mn 0.05-0.5 μ g ml⁻¹; and Fe 0.1-0.5 μ g ml⁻¹ for the construction of the calibration straight lines were prepared in Tris-HCl buffer (0.004 M) of the same concentration as the samples, which were prior to metal analysis fivefold diluted with redestilled water. Detection limits of the selected metals were as follows: Cd 0.003 μ g ml⁻¹; Zn 0.012 μ g ml⁻¹; Cu 0.002 μ g ml⁻¹; Mn 0.002 μ g ml⁻¹; and Fe 0.009 μ g ml⁻¹.

Statistical treatment of results

All statistical analyses (Kruskal-Wallis test, multiple regression analysis, two-way ANOVA) were performed in SigmaStat for Windows Version 1.0, except the principal component analysis (PCA), which was performed in SPSS 10.0 for Windows.

Results and Discussion

MTs, as water-soluble proteins, are present in the cytosol, which is isolated from the composite sample of mussel digestive glands, and purified by heat-treatment. Metal concentrations were also determined in the heat-treated cytosolic fraction, in order to observe the relationship between MTs and metals, particularly Cd, Zn and Cu, which are known inducers of MT synthesis (Langston et al., 1998). Fe and Mn were considered as metals essential for mussels' metabolism. At all 4 sites, the highest metal concentrations present in the heat-treated digestive gland cytosol were those of Zn and Fe with median values of 1 to 2 μ g ml⁻¹, followed by Cu \approx 0.5 μ g ml⁻¹, Mn \approx 0.2 μ g ml⁻¹, and Cd \approx 0.1 μ g ml⁻¹. Median value of MT concentration was \approx 0.6 mg ml⁻¹ (Table 1). The range of values presented in Table 1 comprises the caging period of 12 months.

Temporal variations of digestive gland mass, cytosolic MT and metal concentrations

Variation of average mussel digestive gland mass over one year, at each caging site is presented in Fig. 2. Common is the fact that at 4 caging sites the average digestive gland mass of mussels reached the maximum in April 1998, when the concentration of MTs was minimal, while the peaks of MT concentration were observed in December 1997 and June 1998 (Fig. 3). Temporal variation of digestive gland mass is under influence of food availability and reproductive cycle. Since gonadic tissue penetrates into the digestive gland (Regoli and Orlando, 1993), two tissues cannot be separated by dissection (Regoli and Orlando, 1994). Lower concentrations of trace metals in the gonadic tissues (George and Coombs, 1977; La Touche and Mix 1982; Lobel and Wright, 1982) cause "biological dilution" of trace metal concentrations in the digestive gland. As already stated by Bordin et al. (1997), Amiard-Triquet et al. (1998) and Mouneyrac et al. (1998; 2000), variation of soft tissue mass also influences MT

concentration in bivalves. The "biological dilution" is, thus, a valid explanation for the MT concentration in the digestive gland, too. Kruskal-Wallis test of MT and trace metal concentrations at 4 caging sites shows that differences between selected sites within Kaštela Bay are not statistically significant. Cytosolic concentrations of biochemical and chemical parameters determined in the digestive gland of caged mussels are comparable, leading to the conclusion that living conditions (biotic and abiotic) for mussels are comparable, too. The dependence of MT concentration on the combination of the following independent variables: trace metal concentrations (Cd, Cu, Zn, Mn, Fe), digestive gland mass and the shell mass, was analyzed by multiple linear regression. For that purpose, the results at 4 caging sites were treated as the same population of data. The obtained model explains ~69% of MT concentration variability (Table 2), where Cu concentration has positive statistically significant influence (p<0.001), while Fe concentration and the digestive gland mass have negative statistically significant influence on MTs (p<0.001, p<0.01, respectively). The results of multiple regression analysis, thus, confirm that an increase of average digestive gland mass (Fig. 2) causes MT dilution in that organ, as already observed in Fig. 3. High positive correlation with Cu concentration reflects MT role in the Cu metabolism, as stated by Langston et al. (1998).

MT and trace metal content of the digestive gland

To counteract the effect of "biological dilution", cytosolic MT and metal concentrations were expressed as mass partition (multiplying their cytosolic concentration with factor of 4, which corresponds to the homogenate dilution) and than corrected for the average digestive gland mass; in that manner MT and metal tissue content was calculated. The variation of MT and trace metal contents in the digestive gland of mussels, caged over 12 months at 4 sites within Kaštela Bay, is shown in Fig. 4, a to f. It should be noticed that the maximal values of MT and metal contents were recorded in the period from February to June 1998. Exceptions were station C with high MT content in December 1997, and station B with high Zn content in August 1998, which could be related to somewhat higher digestive gland mass recorded at these two caging sites in the observed period.

Two-way analysis of variance showed statistically significantly higher (p<0.0001) MT and trace metal contents in the period from February to June, compared to the remaining months. In that period, MT contents are nearly two times, and trace metal contents nearly three times higher than during the remaining months. It can be hypothesized that the period of higher MT and metal contents is overlapping with the gonad development and the increased food availability. In the Mediterranean Sea, seasonal hydrological regime varies from winter mixing (January-February) to thermal stratification in summer and fall, causing nutrient depletion in the surface layer during summer, and reinjection to the surface layer during winter mixing (Marty et al, 2002). The same process was observed in the southern Adriatic Sea, where the supply of inorganic nutrients to the upper layer is followed by increased primary production and downward fluxes of particulate matter in early spring (Boldrin et al., 2002). The abundance of food may affect feeding rates and rates of water transport across the gills of filter-feeding organisms, thereby affecting the uptake of metals from both solution and food (Janssen and Scholz, 1979). Free metal ion concentration is the key metal form for its uptake from the solution, while high concentration of metals in food suggests it is an important source of metals, too (Luoma, 1983). Among trace metals, Fe and Mn are predominantly associated with the particles, while significant proportion of Cd is

present in the dissolved state (Phillips and Rainbow, 1993). Exposure to soluble metals results primarily in their accumulation in the tissues like mantle and gills of the mollusks (Amiard, 1978), while metal intake via food results in metal compartmentalization in storage tissue of glandular type, like the digestive gland (Tenore et al., 1968; Viarengo and Nott, 1993). In digestive gland, phytoplankton ingestion is, thus, an important source of metals, because primary producers show some of the highest levels of metal accumulation in food chains (Sanders and Riedel, 1998). As a result of high food availability the contents of metals increase, but simultaneous development of gonads, possibly also stimulated by favourable environmental conditions, and their penetrations. If MTs and trace metals are expressed as tissue content, the simultaneous increase with the average digestive gland mass occurs (Fig. 4, a to f).

Principal component analysis (PCA)

According to PC analysis, 2 components control overall 86% of parameters variability (Table 3), the first component 63% and the second one 23%. The first component associates the following parameters, displaying correlation coefficients higher than 0.80: trace metal contents (Zn, Cu, Fe, Mn), MT content and the digestive gland mass. In the period from February to June, as seen on Fig. 4, simultaneous increase of the content of MTs and analyzed trace metals in the digestive gland of mussels has been observed, as well as the increase of digestive gland mass (Fig. 2), which is in accordance with the role of mussel digestive gland as a tissue for intracellular and extracellular digestion and storage of nutrients (Bayne et al., 1976). The abundance of food during that time of year enables the accumulation of the substances needed for

gonad development (like glycogen, lipids and proteins), thereby stimulating the process of gametogenesis. Glycogen constitutes the dominant energy reserve fueling gametogenesis. When the stored glycogen, lipids and proteins are exhausted, gametogenesis may be fueled directly by circulating metabolites originating from the transformation of ingested food (De Zwaan and Mathieu, 1992). Due to intensified food supply, increased uptake of metals, some of which are known as MT inducers, also occurs. However, increased levels of metals that are observed in the period from February to June do not have to result in additional induction of MT synthesis, because, compared to the other tissues, MT level in the digestive gland of mussels is already high, especially during the periods of intense food uptake. As a result of high basal physiological pool of MTs, presence of free binding sites on MT molecules can be presumed. After saturating free SH-sites, metals, like Cd and Cu(I) ions, which have higher stability constants of thiolate clusters, are able to displace Zn ions from a basal MT pool (Viarengo et al., 1985). The first component obtained by PCA, thereby, associates simultaneous abundance of food and gonad development with the higher digestive gland mass, MT and metal contents of mussels caged within Kaštela Bay. Cytosolic metal content of the digestive gland is therefore related to "trophically available metal" (Wallace and Luoma, 2003). Cadmium has the lowest correlation coefficient (0.62) with the first component. It is not surprising, since previous findings indicate that Cd accumulation is highly related to its concentration in seawater (Zaroogian, 1980; Cossa, 1988). Food is less important source of Cd to marine invertebrates (Kerfoot and Jacobs, 1976).

The second component associates the following parameters: shell mass, digestive gland mass and the contents of Cd, Zn and MTs. Due to the strong correlation with the shell

mass (0.94), the second component is associated with aging process of caged mussels. Our results confirm the statement by Fischer (1983), that calcareous shell continues to be formed even when the length increments are not observed. During 12 months of caging, calcareous shell mass increased on average from 4.5 to 7.0 g (Fig. 5), while the average length increased only 0.7 cm. Important is to point out the relationship between the aging process and the increased Cd content (correlation coefficient 0.62), indication of Cd accumulation in the digestive gland of mussels caged in the coastal zone which is under the urban pressure. Within "aging component" somewhat lower correlation coefficients were obtained for Zn and MT contents (0.47 and 0.28, respectively). The digestive gland contents of Cu, Fe and Mn are not associated with the mussels' age, but are "trophically available metals". The increase of both Cd and Zn contents with mussels' age in the cytosol of digestive gland is consistent with the known affinity of these metals for MTs. Since the caged mussels were transplanted from the same aquaculture site to 4 different sites within Kaštela Bay, Cd levels measured in October 1997 were practically identical at all 4 sites (Fig. 4b). With the time elapsed, the differences in the Cd content in the digestive gland of mussels caged at 4 sites became evident (Fig. 4b), probably due to different bioavailability of Cd at different sites. The differences are specifically well observed in June 1998, when the Cd content is at the maximum, and decreases along the caging sites as follows: A>C>B=D. The highest Cd content at site A can be explained by the fact that site A is flashed with the incoming water masses from the Split channel, where the urban run-off water is discharged via mixed sewer system into the Split harbour (Bogner et al., 1998). Although time-related accumulation of Cd, as toxic metal, occurs differently at 4 caging sites, these differences are not reflected in the level of MTs (Fig. 4a). The conclusion is that Cd in the digestive gland of mussels has not reached the level at which it could induce additional MT

synthesis, because the basal physiological pool of MTs in the digestive gland is already high and is related to high tissue content of essential metals, Zn and Cu (Geffard et al., 2001). The existing level of MTs is, thus, sufficient to detoxify the levels of cytosolic metals (Cd, Cu, Zn) reported in this study, without the need for additional induction. Similar observation was previously reported for digestive gland of <u>Littorina littorea</u> and <u>Ruditapes decussata</u>, where only small overall increase in MT level over the year could be detected, in spite of relatively high Cd influx, because Cd was sequestered by high levels of MTs inherent to that tissue, probably in relation to storage and turnover of essential metals (Bebianno et al., 1992; Bebianno et al., 1993). Geffard et al. (2001) came to similar conclusion for <u>Crassostrea gigas</u>, emphasizing that the digestive gland of this oyster has limited value as a tissue for biomonitoring through the measurement of MTs as a biomarker. The low level of significant <u>de novo</u> MT synthesis, as was suggested by Bebianno et al. (1993) for <u>R. decussata</u>, renders digestive gland unsuitable for detecting responses to sublethal level of Cd in the case of <u>Mytilus galloprovincialis</u>, too.

Concluding remarks

Digestive gland of mussels <u>Mytilus galloprovincialis</u> is often selected as a target tissue for the analysis of MTs as a biomarker of metal exposure. For environmental survey, of special interest is to resolve the effect of toxic metal Cd by means of MT level. Digestive gland, as a storage organ for nutrients, has the highest total protein content, compared to other mussel tissues, and consequently the highest content of a specific metal-binding proteins (MTs). Although, time-related accumulation of toxic metal Cd occurs differently at 4 caging sites within the survey area, these differences are not reflected in the MT level. Since basal MT pool in the digestive gland of *M*. *galloprovincialis* is high, related to the high cytosolic contents of Cu and Zn, cadmium content has not reached the level which might induce additional MT synthesis. Inspite of high metal accumulation in the digestive gland, this organ does not meet the criteria as a tissue of choice for estimating the exposure of *M*. *galloprovincialis* to sublethal levels of Cd. The physiological changes caused by gonadal development and food abundance contribute more to the changes in MT level than the bioavailable Cd concentrations.

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- Amiard JC. ^{110m}Silver contamination mechanisms in a marine benthic food chain. 4.
 Effects of contamination mechanisms on radionuclide elimination. J. Exp. Mar.
 Biol. Ecol. 1978; 34: 215-225.
- Amiard-Triquet C, Rainglet F, Larroux C, Regoli F, Hummel H. Metallothioneins in Arctic bivalves. Ecotoxicol. Environ. Saf. 1998; 41: 96-102.
- Barić A, Marasović I, Gačić M. Eutrophication phenomenon with special reference to the Kaštela Bay. Chem. Ecol. 1992; 6: 51-68.
- Bayne BL, Thompson RJ, Widdows J. Physiology: I. In: Bayne BL, editor, Marine mussels: their ecology and physiology, Cambridge University Press, Cambridge, 1976, pp. 121-206.
- Bebianno MJ, Langston WJ. Metallothionein induction in <u>Mytilus edulis</u> exposed to cadmium. Mar. Biol. 1991; 108: 91-96.
- Bebianno MJ, Langston WJ, Simkiss K. Metallothionein induction in <u>Littorina littorea</u> (Mollusca: Prosobranchia) on exposure to cadmium. J. Mar. Biol. Assoc. UK 1992; 72: 329-342.
- Bebianno MJ, Nott JA, Langston WJ. Cadmium metabolism in the clam <u>Ruditapes</u> <u>decussata</u>: the role of metallothioneins. Aquat. Toxicol. 1993; 27: 315-334.
- Beg-Paklar G, Gačić M. The wind effect on the Kaštela Bay current field. Acta Adriatica 1997; 38:31-43.
- Bogner D, Juračić M, Odžak N, Barić A. Trace metals in fine-grained sediments of the Kaštela Bay, Adriatic Sea. Wat. Sci. Technol. 1998; 38:169-175.

- Boldrin A, Miserocchi S, Rabitti S, Turchetto MM, Balboni V, Socal G. Particulate matter in the southern Adriatic and Ionian Sea: characterisation and downward fluxes. J. Mar. Syst. 2002; 33: 389-410.
- Bordin G, McCourt J, Rodriguez AR. Trace metals in the marine bivalve <u>Macoma</u> <u>baltica</u> in the Westerschelde Estuary (The Netherlands). Part 1: Analysis of total copper, cadmium, zinc and iron concentrations-Locational and seasonal variations. Sci. Total Environ. 1992; 127: 255-280.
- Bordin G, McCourt J, Cordeiro-Raposo F, Rodriguez AR. Metallothionein-like metalloproteins in the baltic clam <u>Macoma balthica</u>: seasonal variations and induction upon metal exposure. Mar. Biol. 1997; 129: 453-463.
- Cossa D. Cadmium in Mytilus spp: worldwide survey and relationship between seawater and mussel content. Mar. Environ. Res. 1988; 26: 265-284.
- De Kock WC, Kramer KJM. Active biomonitoring (ABM) by translocation of bivalves molluscs. In: Kramer KJM, editor, Biomonitoring of coastal waters and estuaries, CRC Press, Boca Raton, 1994, pp. 51-84.
- De Zwaan A, Mathieu M. Cellular biochemistry and endocrinology. In: Gosling E, editor, The mussel Mytilus: Ecology, physiology, genetics and culture, Elsevier Science Publishers B.V., Amsterdam, 1992, pp. 223-307.
- Fischer H. Shell Weight as an independent variable in relation to cadmium content of molluscs. Mar. Ecol. Prog. Ser. 1983; 12: 59-75.
- Geffard A, Amiard-Triquet C, Amiard JC, Mouneyrac C. Temporal variations of metallothionein and metal concentrations in the digestive gland of oysters (<u>Crassostrea gigas</u>) from a clean and a metal-rich site. Biomarkers 2001; 6: 91-107.

- George SG, Coombs TL. The effects of chelating agents on the uptake and accumulation of cadmium by Mytilus edulis. Mar. Biol. 1977; 39: 261-268.
- Giusti L, Williamson AC, Mistry A. Biologically available trace metals in <u>Mytilus</u> <u>edulis</u> from the coast of Northeast England. Environ. Int. 1999; 25: 969-981.
- Janssen HH, Scholz N. Uptake and cellular distribution of cadmium in <u>Mytilus edulis</u>. Mar. Biol. 1979; 55: 133-141.
- Kerfoot WB, Jacobs SA. Cadmium accrual in combined wastewater treatmentaquaculture system. Environ. Sci. Technol. 1976; 10: 662-667.
- Kušpilić G, Marasović I, Vukadin I, Odžak N, Stojanoski L. Hydrographic conditions and nutrient requirements during red tide in the Kaštela Bay (middle Adriatic). Acta Adriatica 1991; 32:813-826.
- Langston WJ, Bebianno MJ, Burt GR. Metal handling strategies in molluscs. In: Langston WJ, Bebianno MJ, editors, Metal metabolism in aquatic environments, Chapman & Hall, London, 1998, pp. 219-283.
- La Touche YD, Mix MC. Seasonal variation of arsenic and other trace elements in bay mussels (<u>Mytilus edulis</u>). Bull. Environ. Contam. Toxicol. 1982; 29: 665-670.
- Lobel PB, Wright DA. Gonadal and nongonadal zinc concentrations in mussels. Mar. Pollut. Bull. 1982; 13: 320-323.
- Luoma SN. Bioavailability of trace metals to aquatic organisms a review. Sci. Total Environ. 1983; 28: 1-22.
- Marty JC, Chiaverini J, Pizay MD, Avril B. Seasonal and interannual dynamics of nutrients and phytoplankton pigments in the western Mediterranean Sea at the DYFAMED time-series station (1991-1999). Deep-Sea Res. (II Top. Stud. Oceanogr.) 2002; 49: 1965-1985.

- Mouneyrac C, Amiard JC, Amiard-Triquet C. Effects of natural factors (salinity and body weight) on cadmium, copper, zinc and metallothionein-like protein levels in resident populations of oysters <u>Crassostrea gigas</u> from a polluted estuary. Mar. Ecol. Prog. Ser. 1998; 162: 125-135.
- Mouneyrac C, Geffard A, Amiard JC, Amiard-Triquet C. Metallothionein-like proteins in <u>Macoma balthica</u>: effects of metal exposure and natural factors. Can. J. Fish. Aquat. Sci. 2000; 57: 34-42.
- Odžak N, Zvonarić T, Kljaković-Gašpić Z, Barić A. Biomonitoring of copper, cadmium, lead, zinc and chromium in the Kaštela Bay using transplanted mussels. Fres. Environ. Bull. 2001;10: 37-41.
- Phillips DJH, Rainbow PS. Biomonitoring of trace aquatic contaminants, Elsevier Science Publishers Ltd, London, 1993.
- Raspor B, Pavičić J, Branica M. Possible biological reference material for environment control analyses - cadmium induced proteins from <u>Mytilus galloprovincialis</u>. Fresenius Z. Anal. Chem. 1987; 326: 719-722.
- Raspor B, Paić M, Erk M. Analysis of metallothioneins by the modified Brdička procedure. Talanta 2001; 55: 109-115.
- Regoli F, Orlando E. <u>Mytilus galloprovincialis</u> as a bioindicator of lead pollution: biological variables and cellular responses. Sci. Total Environ. 1993; Suppl (2): 1283-1292.
- Regoli F, Orlando E. Seasonal variation of trace metal concentrations in the digestive gland of the Mediterranean mussel <u>Mytilis galloprovincialis</u> comparison between a polluted and a non-polluted site. Arch. Environ. Contam. Toxicol. 1994; 27: 36-43.

- Sanders JG, Riedel GF. Metal accumulation and impacts in phytoplankton. In: Langston WJ, Bebianno MJ, editors, Metal metabolism in aquatic environments, Chapman & Hall, London, 1998, pp. 59-76.
- Tenore KR, Horton DE, Duke TW. Effects of bottom substrate on the brackish water bivalve <u>Rangia cuneata</u>. Chesapeake Sci. 1968; 9: 238-248.
- Tudor M, Distribution and residence time of mercury in seawater and sediments of the Kaštela Bay. Ph. D. Thesis 1993, Faculty of Natural Sciences, University of Zagreb, Zagreb (in Croatian),
- UNEP/MAP. MED POL Phase III, Programme for the assessment and control of pollution in the Mediterranean region. MAP Tech. Rep. Ser. No. 120, UNEP, Athens, 1999.
- Viarengo A, Nott JA. Mechanisms of heavy metal cation homeostasis in marine invertebrates. Comp. Biochem. Physiol., C 1993; 104: 355-372.
- Viarengo A, Palmero S, Zanicchi G, Capelli R, Vaissiere R, Orunesu M. Role of metallothioneins in Cu and Cd acumulation and elimination in the gill and digestive gland cells of <u>Mytilus galloprovincialis</u> Lam. Mar. Environ. Res. 1985; 16: 23-36.
- Wallace WG, Luoma SN. Subcellular compartmentalization of Cd and Zn in two bivalves. II. Significance of trophically available metal (TAM). Mar. Ecol. Prog. Ser. 2003; 257: 125-137.
- Yang MS, Chiu ST, Wong MH. Uptake, depuration and subcellular distribution of cadmium in various tissues of <u>Perna viridis</u>. Biomed. Environ. Sci. 1995; 8: 176-185.
- Zaroogian GE. <u>Crassostrea virginica</u> as an indicator of cadmium pollution. Mar. Biol. 1980; 58: 275-284.

Table 1. The median and the concentration ranges of MTs, Cd, Zn, Cu, Mn and Fe in the heat-treated cytosol (S30) of the composite sample of digestive glands of <u>Mytilus</u> <u>galloprovincialis</u> Lmk., caged over 12 months at 4 sites (Fig. 1) within Kaštela Bay. Concentration ranges comprise the whole caging period of 12 months.

	MTs	<u>Cd</u>	Zn	<u>Cu</u>	<u>Mn</u>	Fe
	mg ml ⁻¹	µg ml ⁻¹	µg ml ⁻¹	µg ml⁻¹	µg ml ⁻¹	µg ml ⁻¹
Caging	Median	Median	Median	Median	Median	Median
site	Min-Max	Min-Max	Min-Max	Min-Max	Min-Max	Min-Max
Α	0.677	0.138	1.761	0.508	0.243	1.373
	0.525-0.865	0.084-0.205	1.355-2.366	0.374-0.621	0.171-0.365	1.235-1.853
В	0.656	0.100	1.879	0.516	0.268	1.381
	0.511-0.817	0.079-0.120	1.273-2.245	0.464-0.653	0.205-0.400	1.116-1.869
С	0.637	0.123	2.070	0.532	0.227	1.520
	0.547-0.812	0.093-0.165	1.245-2.232	0.457-0.766	0.208-0.397	1.148-2.161
D	0.645	0.122	2.154	0.587	0.218	1.377
	0.524-0.754	0.094-0.134	1.313-2.583	0.363-0.786	0.173-0.415	1.045-1.988

Table 2. Multiple linear regression analysis: Dependence of the heat-treated cytosolic MT concentration on the following independent variables: cytosolic trace metal concentrations, digestive gland mass and the shell mass

 $(R^2 = 0.69)$

Variable	Coefficient	р
Constant	0.89	< 0.0001
Cd	0.68	0.1464
Zn	-0.06	0.2164
Cu	0.84	0.0006^{*}
Mn	0.26	0.2068
Fe	-0.25	0.0008^{*}
Digestive gland mass	-0.59	0.0089*
Shell mass	-0.03	0.1089

* statistically significant dependence

Table 3. Parameters variability (trace metal and MT contents, digestive gland and shell mass) resolved in two principal components, by Varimax rotation method and Kaiser normalization. Cumulative % of variance amounts to 86%. First component (63% of variance) is associated to "trophically available metals" (TAM) and the second one (23% of variance) to the mussels age.

	Correlation coefficients		
	Component 1	Component 2	
Cd (µg)	0.62	0.62	
Zn (µg)	0.81	0.47	
Cu (µg)	0.96	0.17	
Fe (µg)	0.93	0.08	
Mn (µg)	0.84	-0.04	
MT (mg)	0.89	0.28	
Digestive gland mass (g)	0.85	0.46	
Shell mass (g)	0.01	0.94	
% of variance	63	23	
Cumulative % of variance	86		

- Fig. 1 Marine coastal area of the Kaštela Bay, Eastern Adriatic Sea, and the deployment sites (A to D) for caged mussels <u>Mytilus galloprovincialis</u> Lmk.
- Fig. 2 Temporal variation of average digestive gland mass (g) of mussels <u>M.galloprovincialis</u> caged over 12 months at 4 sites (A= $\int; B=(; C=|; D=\int)$).
- Fig. 3 Temporal variation of MT concentration (mg ml⁻¹) in heat-treated cytosol (S30) of digestive gland of <u>M. galloprovincialis</u> caged over 12 months at 4 sites (A= $\int; B=(; C=|; D=\int)$).
- Fig. 4 Temporal variation of MT (mg) and trace metal tissue content (μ g) in mussels <u>M. galloprovincialis</u> caged over 12 months at 4 sites: a) MTs, b) Cd, c) Mn, d) Cu, e) Zn and f) Fe. Site legend: A= $\int ; B= f : C= |; D= f$.
- Fig. 5 Temporal increase of average calcareous shell mass (g) of mussels <u>M.galloprovincialis</u> caged over 12 months at 4 sites (A to D, Fig. 1).









