

1 **Changes in the planktonic community structure related to trophic conditions:**
2 **the case study of the northern Adriatic Sea**

3 Dragica Fuks¹, Ingrid Ivančić^{1*}, Mirjana Najdek¹, Davor Lučić², Jakica Njire², Jelena
4 Godrijan¹, Daniela Marić¹, Maria Blažina¹, Sandi Orlić¹, Paolo Paliaga¹, Robert
5 Precali¹ and Tina Šilović¹

6 ¹Center for Marine Research, Ruđer Bošković Institute, 52210 Rovinj, Croatia

7 ²University of Dubrovnik, Institute for Marine and Coastal Research, 20000

8 Dubrovnik, Croatia

10 **Abstract**

11 During the 2003-2008 period differentiation in the plankton food web structure and
12 the carbon flow within it was studied in situation when the system of the northern
13 Adriatic Sea, one of the most productive area in the Mediterranean, switched from
14 low nutrient to higher nutrient regime. The biomasses distribution between
15 autotrophs, bacteria, protozoans and metazoans showed that within the upper part of
16 the water column the microbial food web was developed during the stratification
17 period (May-October) in oligotrophic conditions, with a larger
18 heterotrophic/autotrophic (H/A) ratio in the western (1.25-1.88) than in the eastern
19 part (1.23-1.45). The classical food web (H/A 0.36-0.63) was observed in the
20 stratification period during nutrient supply by freshwater or by mixing throughout the
21 water column (November-March). However, while the stratification period with
22 freshets was characterized by an enhanced biomass of autotrophs and heterotrophs,
23 during the mixing period reduction in biomass between consecutive trophic levels
24 indicated increased carbon export from the area. In the lower part of the water
25 column the multivorous food web was observed with greater export of the primary
26 produced organic matter in the western area irrespective of nutrient regime.

27 *Key words:* biomass pyramid, chl a, bacteria, heterotrophic flagellates, ciliated
28 protozoans, metazoans, northern Adriatic Sea

29

30 **1. Introduction**

31 One of the important aims of the community ecology is the understanding of
32 carbon flux through the trophic levels. There is evidence that the transfer of organic
33 carbon through the trophic levels differs between regions with diverse productivity.
34 The plankton from unproductive regions is characterized by high relative
35 heterotrophic biomasses resulting in an inverted biomass pyramid, whereas the
36 plankton from productive areas is characterized by a smaller contribution of
37 heterotrophs and a broad autotrophic base (Gasol et al., 1997). The Northern Adriatic
38 is one of the most productive regions of the Mediterranean Sea at several trophic
39 levels, from phytoplankton to fish (Vollenweider et al., 1992). Particularly, high but
40 variable plankton standing crop and production was quantified off the Po River delta

41 and related to the spreading of its plume (Gilmartin and Revelante, 1981). All
42 plankton biomasses show a decreasing gradient eastward from the Po River and
43 southward from the northern Adriatic (Fonda Umani 1996). The microzooplankton
44 composition is characterized by the dominance of ciliates other than tintinnids
45 (Revelante and Gilmartin, 1990), while the mesozooplankton by strictly neritic
46 copepod and cladoceran species (Fonda Umani 1996; Fonda Umani et al., 2005).
47 Copepod nauplii of the smallest size fractions were revealed as the major mediators
48 of material transfer between primary producers and higher trophic levels (Lučić et al.,
49 2003; Kršinić et al., 2007).

50 As in many coastal systems worldwide influenced by external nutrient load, the
51 productivity level in the northern Adriatic presents marked spatial and seasonal
52 changes related to the alternating influence of freshwater from the western coast and
53 advection of central Adriatic water (CAW) along the eastern coast. The prevalence of
54 cyclonic circulation during winter causes the inflow of oligotrophic CAW into the
55 region and the outflow of more eutrophic riverine waters along the western coast
56 ("open" circulation), while in the late spring and summer the formation of gyres
57 causes a lower exchange with the remainder of the Adriatic ("closed" circulation)
58 (Hopkins et al., 1999). During the winter enrichment of upper waters with nutrients
59 regenerated in deeper waters is superimposed to the CAW inflow. In contrast, in late
60 spring and summer months a stable pycnocline does not permit to nutrients from
61 deeper waters to reach upper waters where nutrient pools are consumed sustaining
62 phytoplankton growth, thus creating oligotrophic conditions. These conditions are
63 occasionally interrupted by the freshwater supply of new nutrients.

64 Since the relative biomass distribution between heterotrophs and autotrophs is
65 regulated by nutrient supply (Duarte et al., 2000), the objectives of the study were to
66 evaluate the functioning of the microbial food web in situation when the freshwater
67 supply of nutrients caused the production of new organic matter (mainly eutrophic
68 conditions) and the situation without a new input of nutrients (mainly oligotrophic
69 conditions) in which microbial growth was based on a constant recycling of the once
70 produced organic matter. Furthermore, the periods in which the produced organic
71 matter was retained in the system, and the periods in which it was mainly exported
72 from the system of the northern Adriatic were confronted. One of the objectives was
73 also to establish if there were differences in the transfer through the trophic pyramid

74 between the mainly oligotrophic eastern area and the more eutrophic western area.
75 Food web characterisation was allowed by an appropriate data set collected monthly
76 from 2003 to 2008 that included broad changes in trophic state.

77 **2. Material and methods**

78 Measurements were performed at three stations at the Po River delta - Rovinj
79 transect located in the northern Adriatic (Fig. 1) between 2003 and 2008 on a
80 monthly scale. Temperature and salinity were measured continuously throughout the
81 water column during the downcasts of a SEABIRD SBE 25 CTD probe, while
82 nutrients, chl *a*, heterotrophic bacteria (HB) and heterotrophic flagellates (HF) were
83 measured at five depths (surface, 5 m, 10 m, 20 m, and 1 m above the bottom: 30-35
84 m). Microzooplankton was taken at surface, 10 m, 20 m and near the bottom.

85 Water samples were collected with 5 l PVC Niskin samplers. Inorganic nutrients
86 were analyzed in unfiltered water immediately after collection (Parsons et al., 1984;
87 Ivančić and Degobbis, 1984). Dissolved inorganic nitrogen (DIN) was calculated as
88 the sum of nitrate, nitrite and ammonia. Total chlorophyll *a* concentrations (chl *a*)
89 were determined by filtration of 500 ml on Whatman GF/C filters. Filters were frozen
90 (−18 °C) and analyzed within a few days by the fluorometric procedure after Parsons
91 et al. (1984). Samples for heterotrophic bacteria (HB), heterotrophic pico- and
92 nanoflagellates (HF) abundances were preserved with formaldehyde (2% final
93 concentration) and stored at 4 °C. HB and HF were estimated by epifluorescence
94 microscopy (Leitz Laborlux D and Nikon Microphot-SA at a magnification of 1000x).
95 HB was determined after staining with 4', 6-diamidino-2-phenylindole (DAPI; 1 µg ml⁻¹,
96 final conc.) following a modification of the method of Porter and Feig (1980). HF
97 was obtained by the primuline staining technique (Caron, 1983). Microzooplankton
98 samples were preserved with formaldehyde (2.5% final concentration) neutralized
99 with CaCO₃. In the laboratory, samples were sedimented until the original volume of
100 5 L was reduced to 10 mL, which took 72 hours (Kršinić, 1980). The organisms were
101 counted with an Olympus inverted microscope at magnifications of 100x and 400x.
102 The microzooplankton was separated in two categories: ciliated protozoans
103 (nonloricate ciliates and tintinnids), and metazoans (copepods nauplii, copepodites
104 and small adult copepods).

105 The autotrophic carbon content was obtained by converting chl *a* using a factor of
106 47 µg C/µg chl *a* (Latasa et al., 2005) and HB by a conversion factor of 20 fg C/cell
107 (Lee and Fuhrman, 1987). The carbon content in HF was obtained summing the
108 content in heterotrophic pico- and nanoflagellates. Heterotrophic picoflagellates'
109 abundance was converted to carbon content by a factor of 1500 fg C/cell (Zubkov et
110 al., 1998). Heterotrophic nanoflagellates cell volumes were estimated using the
111 spherical or cylindrical equation (Norland, 1993) and volumes were converted to
112 carbon content by the factor 0.22 pg C µm⁻³ (Børsheim and Bratbak, 1987). The
113 biovolume of nonloricate ciliates was calculated by comparing the shape of the
114 plasmatic body of each organism to one or more geometrical bodies (Edler, 1979).
115 The tintinnids biovolume was estimated for each species by measuring the linear
116 dimensions of the lorica. The geometrical method was also applied to determine the
117 biovolume of copepod nauplii and postnaupliar copepods. The biovolume of nauplii
118 was calculated according to the modified formula for the biovolume of rotifers
119 (Ruttner-Kolisko, 1977). The body of almost all copepods may be equated to two
120 geometrical forms: cephalothorax to the ellipsoid, and abdomen to the cylinder
121 (Shmeleva, 1965). The following conversion factors were used to transform
122 biovolumes into carbon biomass: for nonloricate ciliates 0.14 pgC µm⁻³ (Putt and
123 Stoecker, 1989), for tintinnids 444.5 pgC + (lorica volume in µm⁻³ x 0.053 pgC)
124 according to Verity and Langdon (1984), for copepod nauplii and postnaupliar
125 copepods 0.08 pgC µm⁻³ (Beers and Stewart, 1970; Monti and Fonda Umani, 1999).

126 The biomass pyramid was constructed taking the autotrophic biomass as 1. The
127 bacteria, protozoans (heterotrophic flagellates and ciliated protozoans) and
128 metazoans contribution in community biomass was obtained by dividing their
129 respective biomass by the autotrophic biomass.

130

131 **3. Results**

132 From May to September chl *a* concentrations were related to the freshwater input.
133 In absence of new freshwater input minimal chl *a* concentrations were observed in
134 the whole area (Fig. 2). In these months with "closed" circulation and stratified water
135 column, even low impulses of freshwater discharge can extend to the west (stations
136 SJ101 and SJ108) as indicated by lowering of the surface salinity. Concomitantly,

137 increased chl *a* values were observed, more frequently at the western stations, while
138 at the eastern station (SJ107) this was the case only after large impulses of
139 freshwater discharge, as observed in May 2004 and May-June 2008 (Fig. 2).

140 From November to March chl *a* concentrations were generally higher than during
141 the stratification without freshets and often not related to the freshwater input. For
142 instance during the winter of 2004 the freshwater input was markedly higher than
143 during the winter of 2007. Winter chl *a* concentrations of these two years did not differ
144 markedly (Fig. 2). Most often during the winter river waters did not influence the area,
145 as indicated by generally high surface salinity in these months.

146 Results for April and October are not included in the data elaboration, since in
147 these transient months progressive heating or cooling of the water surface starts the
148 stratification or mixing through the water column, but usually these processes are not
149 completed yet. The circulation in the area turns from “open” to “closed” type (April)
150 and vice versa (October). In these months freshwater could be retained in the area
151 and stimulate phytoplankton growth (case in November 2005 when impulses of 950-
152 1500 m³ s⁻¹ lowered surface salinity at SJ108 for 10 units; Fig. 2), or could be mainly
153 exported from the area not influencing phytoplankton biomass (case in November
154 2006 when similar impulses did not lower surface salinity). All values given in
155 brackets throughout the Results section are medians, if not reported otherwise.

156 **3.1. Nutrient status, autotrophic biomass and heterotrophic abundance**

157 During the May-September period DIN concentrations in the water column of the
158 eastern station were minimal (0.67-0.82 μmol l⁻¹) and PO₄ was practically exhausted
159 (Fig. 3). Comparably low nutrient concentrations were found in the water column of
160 the western stations in occasions when there was no alimentation with freshwater
161 (steady conditions; Fig. 3). Higher nutrient concentrations, at all stations were
162 observed only in bottom waters. Consequently, chl *a* values in the water column were
163 minimal and similar in the entire area (0.22-0.31 μg l⁻¹), and higher values were
164 observed only in bottom waters (Fig. 4). In conditions of similar chl *a* values, larger
165 abundance of heterotrophic organisms was observed at the western stations,
166 especially in the upper parts of the water column (Fig. 4, 5). The difference in
167 abundances increased with the trophic level: in the upper 5 m at the western stations
168 abundances of HB (61-63 ·10⁷ cell l⁻¹) were for about ¼, for HF (1.1-1.2·10⁶ cell l⁻¹)

169 twice, for ciliates (77 ind. l⁻¹) more than twice and for metazoans (47 ind. l⁻¹) several
170 times higher than at the eastern station.

171 While chl *a* minimum at the eastern station generally persisted during the whole
172 summer, at the western stations episodic inputs of freshwater nutrients, particularly at
173 the surface (DIN 3.7 μmol l⁻¹, PO₄ 0.08 μmol l⁻¹; Fig. 3), increased chl *a* values
174 interrupting the summer minimum (“freshets”; Fig. 4). During these episodes chl *a*
175 concentrations, HB, HF and ciliates abundances were generally maximal during the
176 year practically down to 20 m (Fig 4, 5). Maximal values were found at the surface
177 (chl *a* 2.12 μg l⁻¹, HB 124·10⁷ cell l⁻¹, HF 1.6·10⁶ cell l⁻¹, ciliates 308 ind. l⁻¹) and
178 gradually decreased through the water column. Only the abundances of metazoans
179 at the surface (34 ind. l⁻¹) were generally lower than in the absence of freshwater
180 (Fig. 5). On the contrary, in bottom waters chl *a* concentrations were lower and all
181 microbes were less abundant than in conditions without freshwater influence.

182 From November to March mixing processes homogenized the entire water
183 column. Nutrient concentrations were markedly higher than during the stratification,
184 except during freshets (Fig. 3). At the eastern station higher nutrient concentrations
185 (0.02-0.03 and 1.87-2.12 μmol l⁻¹ for PO₄ and DIN, respectively; Fig. 3) were related
186 to about twice higher chl *a* concentrations (0.46-0.54 μg l⁻¹) than during the
187 stratification (Fig. 4). Yet, abundances of HB (35-39·10⁷ cell l⁻¹; Fig 4b), and
188 particularly HF (0.29-0.36 ·10⁶ cell l⁻¹; Fig 4) in upper waters were lower than during
189 the stratification. In contrast, ciliates abundances (46-63 ind. l⁻¹) were generally about
190 twice higher, while metazoans abundances (12-20 ind. l⁻¹) were comparable to the
191 ones during the stratification (Fig. 5). At the western stations during the mixing period
192 nutrient (0.05-0.15 μmol l⁻¹ and 2.95-5.24 μmol l⁻¹; for PO₄ and DIN, respectively; Fig.
193 3) and chl *a* concentrations (0.54-0.96 μg l⁻¹; Fig. 4) were higher than at the eastern
194 station. Yet, heterotrophs abundances were similar as at the east and lower than
195 during the stratification, with and without freshets (Fig. 4,5). Only ciliates abundances
196 (48-74 ind. l⁻¹) were comparable to the one during the stratification without freshets.

197 During all periods down to 20 m nonloricate ciliates predominated (60-80% of
198 ciliated protozoans abundance) over tintinnids, with the lowest contribution during the
199 stratification with freshets (Table 1). In bottom waters nonloricate ciliates dominated
200 (about 62-65%) over tintinnids during the mixing period, while during the stratification
201 their contribution was nearly equal. Among the metazoan, copepods nauplii

202 dominated in the upper 20 m during the stratification and in the upper 10 m during
203 the mixing period (60-78% of metazoans abundance). In bottom waters copepods
204 nauplii accounted for about half of metazoans abundance (Table 1).

205 **3.2. Transfer of matter through the food web**

206 During the stratification the biomass distribution at the eastern station was similar
207 as at the western stations in steady conditions (Fig. 6). The biomass of heterotrophs
208 largely exceeded the autotrophic one in the water column down to 20 m
209 (heterotrophic/autotrophic biomass ratios -H/A-1.23-1.88; Table 1). H/A ratios were
210 higher at the western than at the eastern stations. At the surface and 10 m, the
211 biomass was about evenly distributed between heterotrophic bacteria and
212 autotrophs, and the transfer to the protozoans was maximal, compared to other
213 depths (Fig. 6). The transfer to the metazoans was higher at 10 and 20 m than at the
214 surface. As opposed to the water column, the bottom layer of both areas was
215 characterized by a smaller contribution of heterotrophs (H/A ratios 0.44-0.58) and a
216 broad autotrophic base, approaching a “normal” pyramidal distribution, otherwise the
217 metazoans contribution exceeded that of protozoans (Fig. 6). The upward transfer
218 was lower than in the water column. The upward transfer in the water column was
219 always higher at the western stations, while it was contrary at the bottom (Fig. 6).

220 During the stratification with freshets, the biomass pyramid was characterized with
221 a broad autotrophic base (H/A 0.38-0.63; Table 1) and a stepwise upward decrease
222 in biomass (Fig. 6). Generally, during freshets, the transfer from autotrophs to higher
223 trophic levels was markedly lower than in steady conditions (Fig 6).

224 During the mixing period the biomass pyramid at eastern and western stations
225 was similar (Fig. 7). In the whole water column the pyramid was characterized by a
226 broad autotrophic base (0.36-0.61, Table 1), and a generally stepwise decrease in
227 biomass upward, although metazoans biomass exceeded that of protozoans (Fig 7).
228 H/A ratios were lower at the western than at the eastern stations. At the western
229 stations the “strength” of upward transfer was lower to that during the stratification in
230 steady conditions (Fig. 6 and 7). At the eastern station the transfer to the
231 heterotrophic bacteria and metazoans was generally higher than at the western
232 stations while the transfer to the protozoans was somewhat lower (Fig. 7).

233

234 **4. Discussion**

235 The most prominent shift from oligotrophic to eutrophic conditions was observed
236 during the stratification period, when a clear change from microbial to classical food
237 web occurred. During the stratification without freshets, characterised with mainly
238 oligotrophic conditions, the shape of the biomass pyramid in upper waters of the
239 entire area was similar and characterized with the heterotrophic dominance of
240 biomass. Such biomass distribution implies a fast turnover of autotrophs which allows
241 them to support high relative biomasses of heterotrophs (Gasol et al., 1997). In the
242 conditions of recycled production microbial food webs play an important role in
243 nutrient recycling (Azam et al., 1983) and bacteria use a higher portion of primary
244 production exerting control over them. The whole study area was dominated by small
245 autotrophs (Socal et al., 2008; CMR, unpubl. data), which were not grazed by
246 metazoans but rather consumed by nano- and microheterotrophs, and most of the
247 fixed carbon flows through the microbial food web as observed in similar conditions
248 (Legendre and Rassoulzadegan, 1995). The small size of the phytoplankton was
249 most probably the reason of the low grazing pressure of herbivorous metazoans
250 during that period, since copepods feed better on larger sized ($>10\mu\text{m}$) particles
251 (Nielsen and Hansen, 1995).

252 The leading role of heterotrophs in the northern Adriatic during the stratification
253 without freshets can be compared to the regular situation in the Mediterranean Sea,
254 but with an opposite spatial trend. In the Mediterranean Sea, higher values of H/A
255 ratios (0.9 to 3.9) were more frequent in oligotrophic regions during stratification
256 (Pedrós-Alió et al., 1999, Siokou-Frangou et al., 2002) and in the most oligotrophic
257 oceanic ecosystems in general (Cho and Azam, 1990, Caron et al., 1995). On the
258 contrary, higher H/A ratios in the northern Adriatic were found at the western, more
259 eutrophic part. In the water column down to 20 m, the overall trend of H/A ratios >1
260 was observed during the stratification without freshets, indicating the high efficiency
261 in keeping resources, that is a pattern more typical for oligotrophic ecosystems
262 (Siokou-Frangou et al., 2010). Although the production of fresh organic matter was
263 low and similar at both areas, a quite higher abundance of bacteria at the west was
264 probably supported by the excess of organic matter retained after freshets or/and
265 related to higher PO_4 input in this area under direct riverine influence. Since PO_4 is

266 confirmed as the limiting element for microbial growth in the northern Adriatic (Ivančić
267 et al., 2009), it could be that P addition had been exploited only by bacteria which
268 can utilize dissolved organic nitrogen thus outcompeting autotrophs, or added P was
269 accumulated in bacteria and picophytoplankton, forming a P enriched diet for
270 grazers. The resulting heterotrophic biomass would then have been quickly
271 channelled toward larger consumers, as observed for other areas (Thingstad et al.,
272 2005; Siokou-Frangou et al., 2010).

273 In contrast, a clear west to east gradient in terms of abundances and biomass
274 transfer was evidenced during the freshets. H/A ratios were lower (0.4-0.6)
275 approaching the values regularly found in eutrophic and coastal zones (~0.2;
276 Ducklow and Carlson, 1992). During intermittent periods of the freshwater nutrient
277 supply, the clear change from microbial to “classical food web” occurred. Increased
278 nutrients led to an enhanced biomass of autotrophs, both nano- and
279 microphytoplankton. The biomass of bacteria, protozoans (except heterotrophic
280 flagellates) and metazoans also increased, matching the changes in autotrophic
281 biomass. Besides the considerably higher ciliated protozoan densities, the most
282 abundant component of the northern Adriatic microzooplankton (Kršinić, 1995), the
283 contribution of tintinnids to the total protozoan biomass also increased. This change
284 in the community structure occurred throughout the entire water column. Similar
285 changes in the structure of protozoan population were recorded in the eutrophicated
286 part of the northern and central Adriatic (Revelante and Gilmartin, 1983; Bojanić et
287 al., 2005). During freshets ciliated protozoans might act as important direct or indirect
288 factor of “top-down” control by cropping on bacteria or their potential consumers,
289 heterotrophic flagellates, respectively. The biomass of metazoans, in which copepod
290 nauplii prevailed over copepods, also increased and exceeded that of protozoans.
291 Since the food availability rapidly increased, the metazoans grazing became very
292 intensive. Although the protozoans are always present in the diet of metazoans, large
293 nauplii and copepods were recognized to be important grazers of microphytoplankton
294 in the northern Adriatic when ephemeral increases of autotrophic biomass occur
295 (Lučić et al., 2003; Fonda Umani et al., 2005; Kršinić et al., 2007). Thus, during the
296 freshets, a fraction of microalgal production would be channelled by herbivorous
297 mesograzers to higher consumers. A shift in biomass distribution, from a dominance
298 of heterotrophs at low to that of autotrophs at high nutrient inputs was experimentally

299 confirmed (Duarte et al., 2000). From these connections between the trophic levels in
300 the northern Adriatic, it derives that during the freshets heterotrophic nutrition (i.e.
301 predation) prevailed, while in steady conditions “bottom-up” control dominated.

302 During the stratification in steady conditions autotrophic biomass notably
303 increased below 20 m, due to the pool of regenerated nutrients. The excess biomass
304 of autotrophs under conditions of nutrient availability fairly exceeded the capacity of
305 heterotrophs to use their production. While in the upper layers bacterial biomass
306 accumulated probably due to closed circulation, in the lower layers it could partly sink
307 to the sediments or be exported southerly from the northern Adriatic. Although the
308 microbial food web pathways were also important, especially on the eastern part, the
309 grazing activity of metazoans increased, removing a considerable portion of microbial
310 biomass. The occurrence of herbivorous and microbial grazing modes (multivorous
311 food web; Siokou-Frangou et al., 2002) suggested more efficient energy transfer to
312 higher trophic levels in the lower part of the water column during the stratification.

313 During the water column mixing (November to March) chl a concentrations were
314 higher than during the stratification without freshets. Phytoplankton growth was
315 sustained mostly by the nutrients regenerated during autumn and redistributed in the
316 water column after the mixing (Gilmartin et al., 1990), while freshwater nutrients were
317 a less important source. Most often the river waters were directed southward in a
318 narrow coastal belt and did not influence the area. During this period the biomass
319 distribution was represented by broad autotrophic base with reduction in biomass
320 between consecutive trophic levels indicating increased carbon export, usual in these
321 conditions (Gasol et al., 1997), being less available to planktonic heterotrophs. In the
322 eastern part a much greater proportion of autotrophs flowed to bacteria than at the
323 west. Consequently at the east lower autotrophic biomass caused significantly lower
324 grazing pressure of metazoans on primary production. Thus copepod predation was
325 probably channelled to ciliates. In contrast, on the west higher biomass of ciliates
326 could be attributed to a weaker grazing control by metazoans. Their grazing pressure
327 was more intensive on autotrophs due to their higher biomass. The greater
328 availability of autotrophs on the west and similar abundance of heterotrophs at both
329 parts, suggests that the export of primary production was larger on the western part.

330 The results showed a seasonal differentiation regarding the food web structure
331 and the carbon flow within it in the northern Adriatic. Although conversion factors for

332 the calculation of autotrophic and heterotrophic biomass could be a source of errors,
333 the obtained results are robust enough to support observed differences. The
334 observation that the microbial food web has been developed in the upper part of the
335 water column during the stratification without new input of nutrients (oligotrophic or
336 steady conditions), whereas the classical food web was found during increased
337 nutrients, provided by freshets and water column mixing, could support a wide range
338 of conversion factors. During the period of microbial food web domination the carbon
339 transfer was higher at the western part, while on the contrary, in the period of
340 classical food web domination the transfer was higher at the eastern part. The
341 multivorous food web was observed in the lower part of the water column with greater
342 export of the primary produced organic matter at the west, irrespective of the season.

343 **Acknowledgements**

344 The authors thank S. Dujmović and M. Buterer for the determination of chl *a* and
345 nutrients. Dr. T. Đakovac, P. Krelja and the crew of RV “Vila Velebita” are thanked for
346 the measurements of hydrographic parameters and the help during sampling. Dr. D.
347 M. Lyons is thanked for English corrections. This work is part of the scientific projects
348 (098-0982705-2729, 098-0982705-2731 and Project “Jadran”) funded by the Ministry
349 of Science, Education and Sports of the Republic of Croatia.

350 **References**

- 351 Beers, J.R., Stewart, G.L. 1970. Numerical abundance and estimated biomass of
352 microzooplankton. Bull. Scripps. Inst. Oceanogr. 17, 67–87.
- 353 Bojanić, N., Šolić, M., Krstulović, N., Šestanović, S., Marasović, I., Ninčević, N. 2005.
354 Temporal variability in abundance and biomass of ciliates and copepods in the
355 eutrophicated part of Kaštela Bay (Middle Adriatic Sea). Helgol. Mar. Res. 59,107-
356 120.
- 357 Børsheim, K.Y., Bratbak, G. 1987. Cell volume to cell carbon conversion factors for
358 bacterivorous *Monas* sp. enriched from seawater. Mar. Ecol. Prog. Ser. 36,171-
359 175.
- 360 Caron, D.A. 1983. A technique for the enumeration of photosynthetic and
361 heterotrophic nanoplankton using epifluorescence microscopy, and a comparison
362 with other procedures. Appl. Environ. Microbiol. 46, 491-498.
- 363 Caron, D.A., Dam, H.G., Kremer, P., Lessard, E.J., Madin, L.P., Malone, T.C., Napp,
364 J.M., Peele, E.R., Roman, M.R., Youngbluth, M.J. 1995. The contribution of
365 microorganisms to particulate carbon and nitrogen in surface waters of the
366 Sargasso Sea near Bermuda. Deep-Sea Res. I 42, 943-972.
- 367 Cho, B.C., Azam, F. 1990. Biogeochemical significance of bacterial biomass in the
368 ocean's euphotic zone. Mar. Ecol. Prog. Ser. 63, 253-259.
- 369 Duarte, C.M., Agusti, S., Gasol, J.M., Vaque, D., Vazquez-Dominguez, E. 2000.
370 Effect of nutrient supply on biomass structure of planktonic communities: an
371 experimental test on a Mediterranean coastal community. Mar. Ecol. Prog. Ser.
372 206, 87-95.
- 373 Ducklow, H.W., Carlson, C.A. 1992. Oceanic bacterial production. Advances in
374 Microb. Ecol. 12, 113-181.
- 375 Edler, L. 1979. Recommendations on methods for marine biological studies in the
376 Baltic Sea. Phytoplankton and chlorophyll. The Baltic marine biologists working
377 group 5, 5–38.
- 378 Fonda Umani, S. 1996. Pelagic biomass and production in the Adriatic Sea. In:
379 Palomera, J, Rubies, P. (Eds) The European anchovy and its environment: Sci.
380 Mar. 60, 65-77.
- 381 Fonda Umani, S., Tirelli, V., Beran, A., Guardiani, B. 2005. Relationships between
382 microzooplankton and mesozooplankton: competition versus predation on natural

383 assemblages of the Gulf of Trieste (northern Adriatic Sea). *J. Plankt. Res.* 27,
384 973-986.

385 Gasol, J.M., delGiorgio, P.A., Duarte, C.M. 1997. Biomass distribution in marine
386 planktonic communities. *Limnol. Oceanogr.* 42, 1353-1363.

387 Gilmartin, M., Revelante, N. 1981. Regional variations in phytoplankton standing
388 crops in the northern Adriatic Sea. *Rapp. Comm. Int. Mer Mediterr.* 27, 47-74.

389 Gilmartin, M., Degobbis, D., Revelante, N., Smodlaka, N. 1990. The mechanism
390 controlling plant nutrient concentrations in the northern Adriatic Sea. *Int. Rev.*
391 *gesam. Hydrobiol. Hydrogr.* 75, 425-445.

392 Hopkins, T.S., Artegiani, A., Kinder, C., Pariente, R, 1999. A discussion of the
393 northern Adriatic circulation and flushing as determined from the ELNA
394 hydrography. In: Hopkins TS, Artegiani A, Cauwet G, Degobbis D, Malej A (Eds.)
395 The Adriatic Sea. Ecosystem Research Report No. 32, EUR 18834. European
396 Commission, Brussel.

397 Ivančić, I., Degobbis ,D. 1984. An optimal manual procedure for ammonia analysis in
398 natural waters by the indophenol blue method. *Wat. Res.* 18, 1143-1147.

399 Ivančić, I., Radić, T., Lyons, D.M., Fuks, D., Precali, R., Kraus, R. 2009. Alkaline
400 phosphatase activity in relation to nutrient status in the northern Adriatic Sea. *Mar.*
401 *Ecol. Prog. Ser.* 378, 27-35.

402 Kršinić, F. 1980. Comparison of methods used in micro-zooplankton research in
403 neritic waters of the Eastern Adriatic. *Nova Thalassia* 4, 91-106.

404 Kršinić, F. 1995. Changes in the microzooplankton assemblages in the northern
405 Adriatic Sea during 1989 to 1992. *J. Plankt. Res.* 17, 935-953.

406 Kršinić, F., Bojanić, D., Precali, R., Kraus, R. 2007. Quantitative variability of the
407 copepod assemblages in the northern Adriatic Sea from 1993 to 1997. *Estuar.*
408 *Coast. Shelf Sci.* 74, 528-538.

409 Latasa, M., Moran, X., Scharek, R., Estrada, M. 2005. Estimating the carbon flux
410 through main phytoplankton groups in the northwestern Mediterranean. *Limnol.*
411 *Oceanogr.* 50, 1447-1458.

412 Legendre, L., Rassoulzadegan, F. 1995. Plankton and nutrient dynamics in marine
413 waters. *Ophelia* 41, 153-172.

414 Lee, S., Fuhrman, J.A. 1987. Relationships between biovolume and biomass of
415 naturally derived marine bacterioplankton. *Appl. Environ. Microbiol.* 53, 1298-
416 1303.

417 Lučić, D., Njire, J., Morović, M., Precali, R., Fuks, D., Bolotin, J. 2003.
418 Microzooplankton in the open waters of the northern Adriatic Sea from 1990 to
419 1993: the importance of copepod nauplii densities. *Helgol. Mar. Res.* 57, 73-81.

420 Monti, M., Fonda Umani, S. 1999. Distribution of the main microzooplankton taxa in
421 the Ross Sea (Antarctica): austral summer 1994. In: Faranda F, Guglielmo L,
422 Ianora A (eds) *Ross Sea ecology—Itali-Antartide expeditions 1987–1995*,
423 Springer-Verlag, Heidelberg

424 Nielsen, T.G., Hansen, B.W. 1995. Plankton community structure and carbon cycling
425 on the western coast of Greenland during and after sedimentation of a diatom
426 bloom. *Mar. Ecol. Prog. Ser.* 125, 239-257.

427 Parsons, T.R., Maita, Y., Lalli, C.M. 1984. *A Manual for Chemical and Biological*
428 *Methods for Seawater Analysis*. Pergamon press, New York

429 Pedrós-Alió, C., Calderón-Paz, J-I., Guixa-Boixereu, N., Estrada, M., Gasol, J.M.
430 1999. Bacterioplankton and phytoplankton biomass and production during
431 summer stratification in the northwestern Mediterranean Sea. *Deep-Sea Res. I*
432 46, 985-1019.

433 Porter, K.G., Feig, Y.S. 1980. The use of DAPI for identifying and counting aquatic
434 microflora. *Limnol. Oceanogr.* 25, 943-948.

435 Putt, M., Stoecker, D.K. 1989. An experimentally determined carbon: volume ratio for
436 marine 'oligotrichous' ciliates from estuarine and coastal waters. *Limnol.*
437 *Oceanogr.* 34, 1097–1103.

438 Revelante, N., Gilmartin, M. 1990. Vertical water column resource partitioning by a
439 ciliated protozoan population under stratified conditions in the northern Adriatic. *J.*
440 *Plankt. Res.* 12, 89-107.

441 Ruttner-Kolisko, A. 1977. Suggestions for biomass calculations of plankton rotifers.
442 *Arch. Hydrobiol. Beih. Ergebn. Limnol.* 8, 71–76.

443 Siokou-Frangou, I., Bianchi, M., Christaki, U., Christou, E.D., Giannakourou, A.,
444 Gotsis, O., Ignatiades. L., Pagou, K., Pitta, P., Psarra, S., Souvermezoglou, E.,
445 Van Wambeke, F., Zervakis, V. 2002. Carbon flow in the planktonic food web
446 along the gradient of oligotrophy in the Aegean Sea (Mediterranean Sea). *J. Mar.*
447 *Syst.* 33-34, 335-353.

448 Siokou-Frangou, I., Christaki, U., Mazzochi, M.G., Montresor, M., Ribera d'Alcala, M.,
449 Zingone, A. 2010. Plankton in the open Mediterranean Sea: a Review.
450 *Biogeosciences* 7, 1543-1586.

- 451 Socal, G., Acri, F., Bastianini, M., Bernardi Aubry, F., Bianchi, F., Cassin, D.,
452 Coppola, J., DeLazzari, A., Bandelj, V., Cossarini, G., Solidoro, C. 2008.
453 Hydrological and biogeochemical features of the Northern Adriatic Sea in the
454 period 2003–2006. *Mar. Ecol.* 29, 449–468.
- 455 Vollenweider, R.A., Rinaldi, A., Montanari, G. 1992. Eutrophication, structure and
456 dynamics of a marine coastal system: results of ten years monitoring along the
457 Emilia Romagna coast (Northwest Adriatic Sea). *Sci. Total Environ. Suppl*, 63-
458 106.
- 459 Zubkov, M.V., Sleigh, M.A., Taran, G.A., Burkull, P.H., Leakey, R.J.G. 1998.
460 Picoplankton community structure on an Atlantic transect from 50°N to 50°S.
461 *Deep-Sea Res.* 45, 1339-1355.

462

463 **Figure caption**

464 Fig. 1. Research area and sampling stations in the northern Adriatic Sea.

465 Fig. 2. Daily mean of the Po River discharge rate (Q), salinity (S) and chlorophyll *a*
466 concentrations (chl *a*) at the surface of sampling stations during the period 2003-
467 2008.

468 Fig. 3. Box-whiskers plot of orthophosphate (PO₄) and dissolved inorganic nitrogen
469 (DIN) during the stratification and mixing period at the eastern (ES) and western
470 (WS) stations in conditions with (freshets) and without freshwater input (steady)
471 during 2003-2008. Surface values are denoted with □, 5 m ■, 10 m ■, 20 m ■,
472 and bottom ■. Vertical bars are referred to 95% of data.

473 Fig. 4. Box-whiskers plot of chlorophyll *a* (chl *a*), heterotrophic bacteria (HB) and
474 heterotrophic flagellates (HF) during the stratification and mixing period at the
475 eastern (ES) and western (WS) stations in conditions with (freshets) and without
476 freshwater input (steady) during 2003-2008. Surface values are denoted with □, 5
477 m ■, 10 m ■, 20 m ■, and bottom ■. Vertical bars are referred to 95% of data.

478 Fig. 5. Box-whiskers plot of ciliated protozoans and metazoans during the
479 stratification and mixing period at the eastern (ES) and western (WS) stations in
480 conditions with (freshets) and without freshwater input (steady) during 2003-
481 2008. Surface values are denoted with □, 10 m ■, 20 m ■, and bottom ■. Vertical
482 bars are referred to 95% of data.

483 Fig. 6. Mean (plus standard error) biomasses of metazoans, protozoans and bacteria
484 relative to that of autotrophic biomass for the stratification period at the eastern
485 and western stations in conditions with (freshets) and without freshwater input
486 (steady) during 2003-2008.

487 Fig. 7. Mean (plus standard error) biomasses of metazoans, protozoans and bacteria
488 relative to that of autotrophic biomass for the mixing period at the eastern and
489 western stations during 2003-2008.

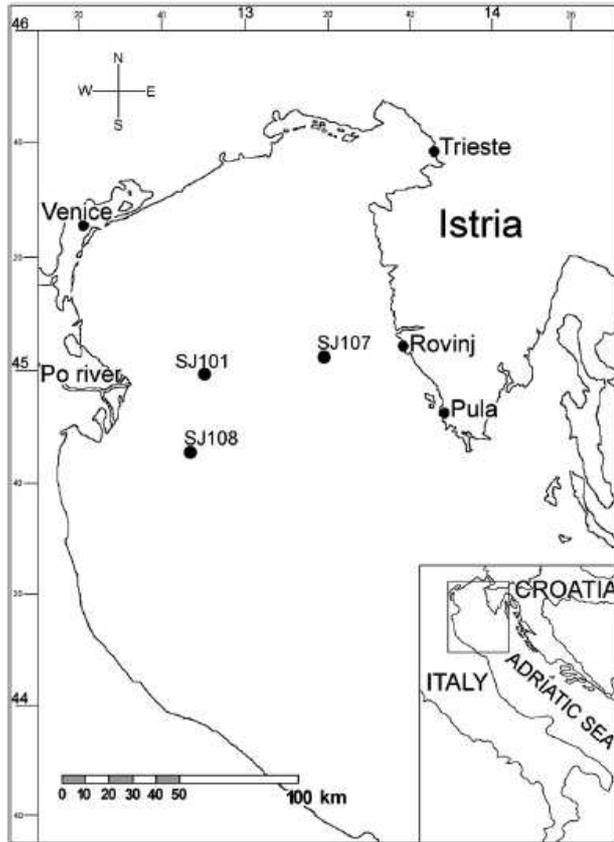


Fig. 1. Research area and sampling stations in the northern Adriatic Sea.

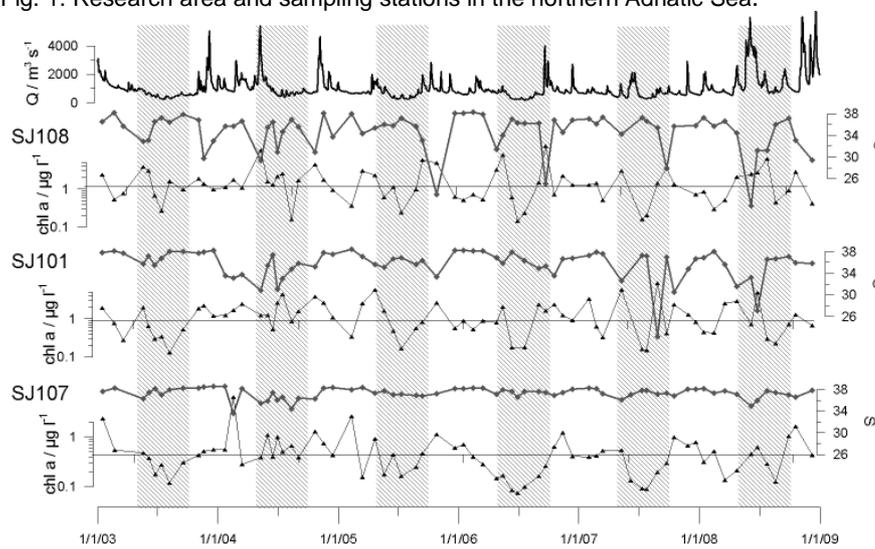


Fig. 2. Daily mean of the Po River discharge rate (Q), salinity (S) and chlorophyll a concentrations (chl a) at the surface of sampling stations during the period 2003–2008. Stratification periods (May–September) are shaded in gray.

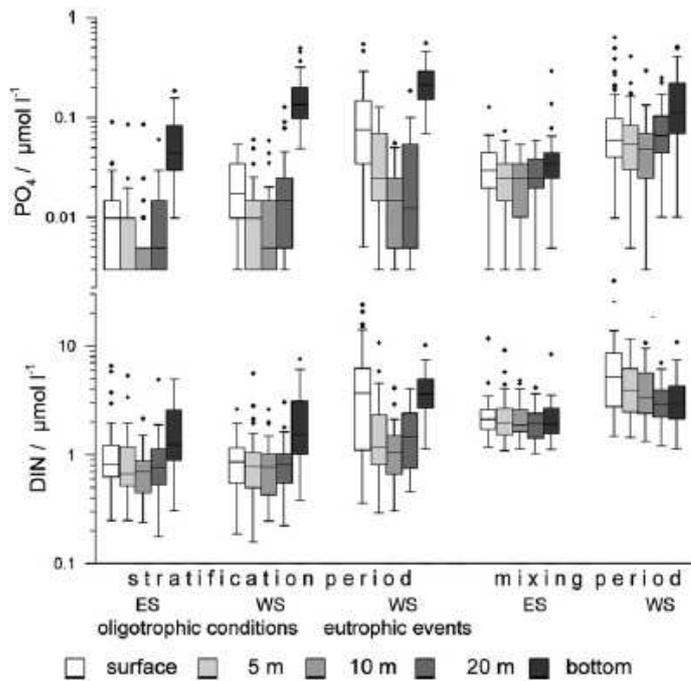


Fig. 3. Box-whisker plot of orthophosphate (PO₄) and dissolved inorganic nitrogen (DIN) during the stratification and mixing period at the eastern (ES) and western (WS) stations in oligotrophic conditions and during eutrophic events for the 2003–2008 period. Vertical bars are referred to 95% of data.

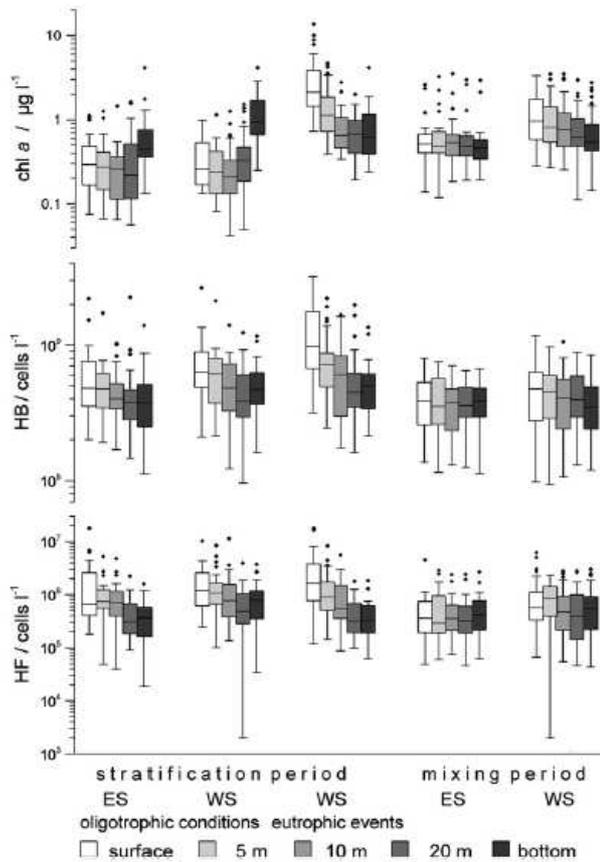


Fig. 4. Box-whisker plot of chlorophyll a (chl a), heterotrophic bacteria (HB) and heterotrophic flagellates (HF) during the stratification and mixing period at the eastern (ES) and western (WS) stations in oligotrophic conditions and during eutrophic events for the 2003–2008 period. Vertical bars are referred to 95% of data.

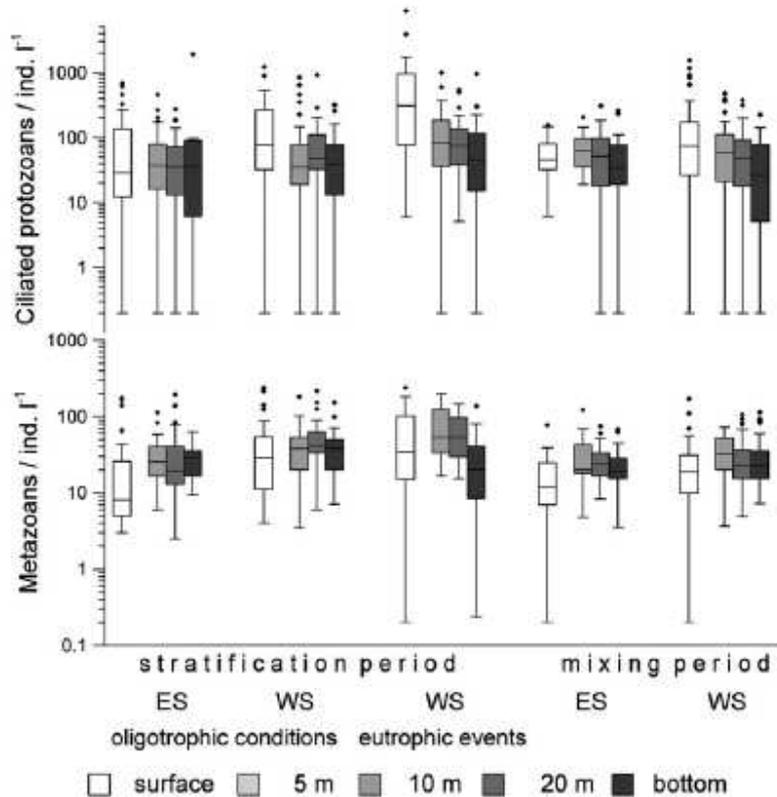


Fig. 5. Box-whisker plot of ciliated protozoans and metazoans during the stratification and mixing period at the eastern (ES) and western (WS) stations in oligotrophic conditions and during eutrophic events for the 2003–2008 period. Vertical bars are referred to 95% of data.

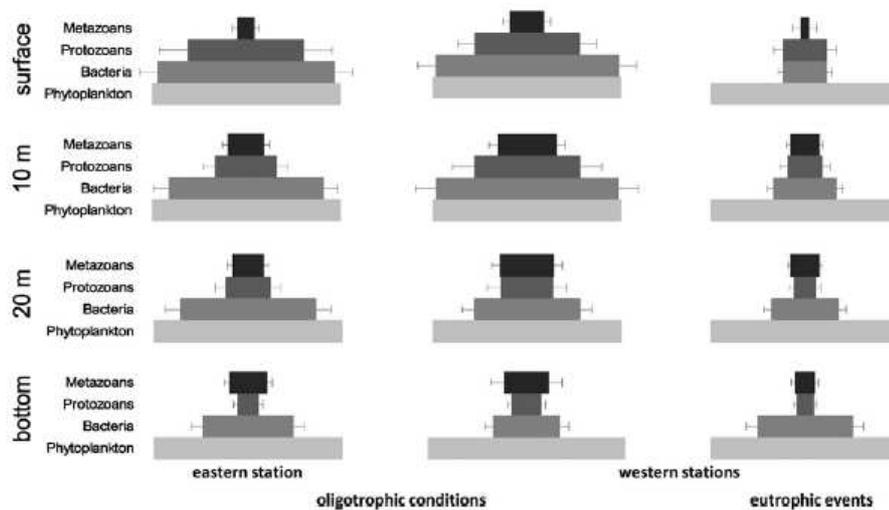


Fig. 6. Mean (plus standard error) biomasses of metazoans, protozoans and bacteria relative to that of autotrophic biomass for the stratification period at the eastern and western stations in oligotrophic conditions and during eutrophic events for the 2003–2008 period.

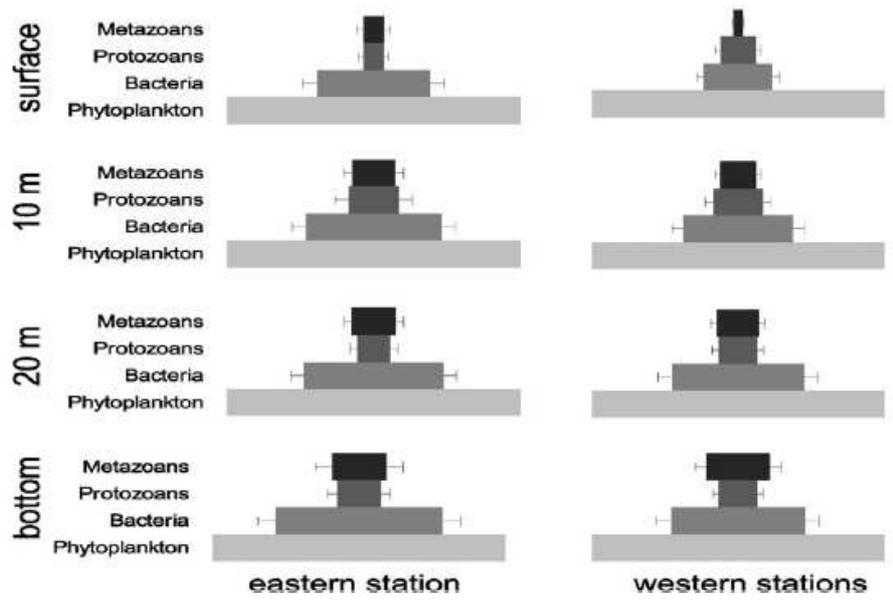


Fig. 7. Mean (plus standard error) biomasses of metazoans, protozoans and bacteria relative to that of autotrophic biomass for the mixing period at the eastern and western stations for the 2003–2008 period.

Table 1. Mean percentage (\pm SE) of nonloricate ciliates in ciliated protozoans and copepods nauplii in metazoans and heterotrophic/autotrophic ratio (H/A) during the stratification and mixing period at the eastern and western stations in conditions with (freshets) and without freshwater input (steady) during 2003-2008.

Parameter	depth	Stratification			Mixing	
		Eastern s.	Western s. steady	Western s. freshets	Eastern s.	Western s.
% Non- loricate ciliates	surf.	71.2 \pm 5.4(33)	70.6 \pm 5.4(34)	63.5 \pm 6.4(35)	71.7 \pm 6.5(26)	82.4 \pm 3.6(52)
	10	80.8 \pm 5.0(33)	74.8 \pm 4.8(37)	68.8 \pm 5.5(31)	82.6 \pm 5.3(26)	80.0 \pm 3.5(52)
	20	70.6 \pm 5.3(33)	71.0 \pm 5.1(37)	54.6 \pm 5.6(31)	79.9 \pm 5.8(26)	69.3 \pm 4.8(51)
	Bott.	55.9 \pm 6.9(31)	54.9 \pm 6.3(37)	44.4 \pm 6.3(31)	65.0 \pm 7.4(25)	61.9 \pm 4.9(51)
% Nauplii	surf.	78.1 \pm 3.6(33)	73.1 \pm 4.1(34)	70.0 \pm 3.1(34)	73.5 \pm 4.8(26)	77.8 \pm 2.8(52)
	10	69.5 \pm 3.8(33)	64.5 \pm 3.7(37)	60.3 \pm 3.3(31)	66.8 \pm 4.2(26)	66.4 \pm 2.6(52)
	20	61.7 \pm 4.2(33)	60.8 \pm 3.8(37)	56.0 \pm 3.3(31)	55.9 \pm 3.2(26)	53.9 \pm 2.9(51)
	Bott.	48.0 \pm 4.1(31)	42.5 \pm 3.6(37)	56.9 \pm 5.2(31)	54.8 \pm 4.3(25)	48.5 \pm 3.8(51)
H/A	surf.	1.45 \pm 0.13(33)	1.64 \pm 0.14(34)	0.38 \pm 0.03(35)	0.53 \pm 0.08(26)	0.36 \pm 0.03(52)
	10	1.44 \pm 0.11(33)	1.88 \pm 0.19(37)	0.62 \pm 0.05(31)	0.49 \pm 0.05(26)	0.43 \pm 0.04(52)
	20	1.23 \pm 0.12(33)	1.25 \pm 0.16(37)	0.63 \pm 0.06(31)	0.51 \pm 0.05(26)	0.50 \pm 0.05(51)
	bott.	0.58 \pm 0.05(31)	0.44 \pm 0.05(37)	0.50 \pm 0.05(31)	0.61 \pm 0.09(25)	0.56 \pm 0.06(51)