

1 Interrelation between environmental conditions, acanthocephalan infection and metal(loid)
2 accumulation in fish intestine: an in-depth study

3

4 Tatjana Mijošek¹, Sara Šariri¹, Zorana Kljaković-Gašpić², Željka Fiket¹, Vlatka Filipović
5 Marijić¹

6

7 ¹ Ruđer Bošković Institute, Bijenička cesta 54, Zagreb, Croatia

8 ² Institute for Medical Research and Occupational Health, Ksaverska 2, 10000 Zagreb,

9 Croatia

10

11

12 *Corresponding author:

13 Vlatka Filipović Marijić

14 Laboratory for Biological Effects of Metals

15 Division for Marine and Environmental Research

16 Ruđer Bošković Institute

17 Bijenička c. 54, 10000 Zagreb, Croatia

18 e-mail: vfilip@irb.hr

19

20

21 **Abstract**

22 Metal(loid) bioaccumulation in acanthocephalans (*Dentitruncus truttae*) and intestines of fish
23 (*Salmo trutta*) from the Krka River, influenced by industrial and municipal wastewaters, was
24 investigated in relation to exposure to meta(loid)s from fish gut content (GC), water, and
25 sediment to estimate potentially available metal(loid)s responsible for toxic effects and cellular
26 disturbances in biota. Sampling was performed in two seasons (spring and autumn) at the
27 reference site (river source, KRS), downstream of the wastewater outlets (Town of Knin,
28 KRK), and in the national park (KNP). Metal(loid) concentrations were measured by ICP-MS.
29 The highest accumulation of As, Ba, Ca, Cu, Fe, Pb, Se and Zn was observed mainly in
30 organisms from KRK, of Cd, Cs, Rb and Tl at KRS, and of Hg, Mn, Mo, Sr and V at KNP.
31 Acanthocephalans showed significantly higher bioaccumulation than fish intestine, especially
32 of toxic metals (Pb, Cd and Tl). Metal(loid) bioaccumulation in organisms partially coincided
33 to exposure from water, sediments and food, while in GC almost all elements were elevated at
34 KNP, reflecting the metal(loid) exposure from sediments. Seasonal differences in organisms
35 and GC indicated higher metal(loid) accumulation in spring, which follows enhanced fish
36 feeding rates. Higher number of acanthocephalans in the intestine influenced biodilution
37 process and lower concentrations of metal(loid)s in fish, indicating positive effects of parasites
38 to their host, as supported by high values of bioconcentration factors. Fish intestine and
39 acanthocephalan *D. truttae* were confirmed as sensitive indicators of available metal fraction
40 in conditions of generally low environmental exposure in karst ecosystem. Since metal(loid)
41 accumulation depended on ecological, chemical and biological conditions, but also on the
42 dietary habits, physiology of organisms and parasite infection, continuous monitoring is
43 recommended to distinguish between the effects of these factors and environmental exposure
44 when assessing dietary associated metal(loid) exposure in aquatic organisms.

46 **Keywords:** bioaccumulation, biodilution, brown trout, acanthocephalans, trace elements,

47 bioconcentration factors

48

49

50 **1. Introduction**

51 Metal(loid)s occur naturally in the environment, but their presence has rapidly increased
52 as a result of anthropogenic activities such as mining, traffic, smelting, agriculture, and the
53 release of industrial or municipal waste. This is of particular concern in freshwater karst
54 ecosystems, characterized by numerous caves, sinkholes, and open fractures in carbonate rocks
55 that, because of high porosity and permeability, allow rapid flow but also significant storage
56 of contaminants, including metal(loid)s, which can be released into the water column over time
57 (Padilla and Vesper, 2018).

58 The most commonly used bioindicator organisms in freshwater ecosystems are bivalves,
59 crustaceans and fish. In recent decades, parasites have also been recognized as potential
60 bioindicators of water quality, leading to the establishment of the field of Environmental
61 Parasitology (Sures et al., 2017). Among them, acanthocephalans have been proposed as
62 promising bioindicators of metal exposure (Filipović Marijić et al., 2013; Nachev and Sures,
63 2016; Mehana et al., 2020). Further, accumulated metals in acanthocephalans represent a
64 biologically available fraction, since contaminants have to cross the tegument and membranes
65 and might positively or negatively affect their hosts (Sures et al., 2017; Molbert et al., 2020).
66 Given their widespread distribution and high prevalence, parasites should not be neglected in
67 environmental research as being a crucial link for the correct interpretation of field studies.

68 Due to the complexity of dietary metal(loid) uptake, correlation to food sources depends
69 on metal(loid) bioavailability in the ingested food and selectivity in metal(loid) accumulation
70 to intestinal tissue in order to maintain homeostasis. Consequently, the uptake and potential
71 toxicity of dietary metal(loid)s have been studied predominantly under controlled laboratory
72 conditions (Ojo and Wood 2007; Creighton and Twining 2010). A drawback of these studies
73 is that they mostly employ higher concentrations of metals and shorter exposure times than in
74 natural conditions (Giguère et al. 2004). There have been limited studies on the metal levels in

75 the intestine of indigenous freshwater fish (Dallinger and Kautzky 1985; Sures et al., 1999;
76 Giguère et al., 2004; Filipović Marijić and Raspor, 2010, 2012; Mijošek et al., 2019, 2022),
77 leaving much of the research on metal uptake from diet under field conditions largely
78 unexplored.

79 Previous studies involving biota from the Krka River have already confirmed negative
80 impact of urban and industrial wastewaters at the KRK site near the Town of Knin in terms of
81 increased accumulation of metal(loid)s and disruption of biomarker responses in organisms
82 compared to the reference site represented by the Krka River source, which is not under known
83 anthropogenic impact and, according to previous studies, not characterized by stress responses
84 (Dragun et al., 2018; Filipović Marijić et al., 2018; Mijošek et al., 2019, 2022). However, there
85 have been no investigations that concurrently examined the concentrations of metalloids in
86 sediments, organisms, and the intestinal contents of fish within the boundaries of the national
87 park.

88 As a continuation of the research in this important area, we measured total concentrations
89 of 22 metal(loid)s in the intestine and gut content (GC) of brown trout *Salmo trutta* and in
90 acanthocephalan *Dentitruncus truttae* from three locations with varying levels of
91 contamination (reference site – river source upstream of the wastewater discharge; pollution
92 impacted site near the Town of Knin – downstream of the wastewater discharge; and the most
93 downstream location directly in the Krka National Park). Additionally, due to significance of
94 dietborne metal uptake, metal(loid)s were measured in fish gut contents (GC) to investigate
95 possible bioavailability and metal(loid) transfer from food to fish. Metal(loid) concentrations
96 were also measured in water and sediment samples, and the relation of environmental
97 conditions with accumulation in organisms was evaluated. This comprehensive investigation
98 was conducted with the following objectives: 1) comparison of bioaccumulated metal(loid)
99 concentrations between fish intestine and acanthocephalans; 2) determination of spatial and

100 seasonal variability of metal(loid) concentrations in fish, acanthocephalans and gut content and
101 their connection to environmental metal(loid) exposure (water and sediment); 3) analysis of
102 possible association of bioaccumulated metal(loid) concentrations with the fish size, sex,
103 condition, feeding habits and acanthocephalan infection rate; 4) evaluation of the intestine and
104 acanthocephalans as bioindicators of metal(loid) exposure and their bioavailable levels and 5)
105 assessment of the overall impact of industrial and municipal wastewaters on the Krka National
106 Park.

107

108 **2. Materials and methods**

109 2.1. Research area

110 Significant part of the Krka River watercourse is protected as the Krka National Park,
111 which is in last decades influenced by extensive tourism, industrial and municipal wastewaters,
112 agriculture and fertilizers, especially in the upper river basin only 2 km upstream of the border
113 of the Krka National Park (Cukrov et al., 2008; Filipović Marijić et al., 2018; Sertić Perić et
114 al., 2018). Based on the results on water quality from previous studies (Filipović Marijić et al.,
115 2018) and current sampling campaigns (Mijošek et al., 2023, Šariri et al., 2024, Table S1),
116 three sites were selected for organisms and sediment sampling – Krka River source (KRS) as
117 the reference site, location downstream of the wastewater outlets and Town of Knin (KRK) as
118 the presumed contaminated site, and the third site in the area of the national park near the Brljan
119 Lake (KNP) (Fig. 1). Detailed description, as well as results on the water analyses were given
120 by Mijošek et al. (2023) and Šariri et al. (2024).

121

122 2.2. Sampling procedures

123 *2.2.1. Organisms*

124 Samplings were conducted in April and October 2021 at the three above described sites.
125 Electrofishing, carried out according to Croatian standard HRN EN 14011, was chosen for fish
126 sampling. Fish were kept aerated in the plastic tanks filled with water.

127 2.2.2. *Water*

128 At each location, river water samples were collected in precleaned polyethylene bottles
129 in triplicates. Appropriate aliquots from each bottle were then filtered into pre-cleaned
130 polyethylene bottles using a cellulose acetate filter with a pore diameter of 0.45 μm (Sartorius,
131 Germany) to measure dissolved metal(loid)s, acidified with concentrated HNO_3 (Rotipuran
132 Supra 69%, Carl Roth, Germany) and stored at 4 °C before further analysis.

133 2.2.3. *Sediments*

134 Surface sediments (upper 10 cm) were sampled only once in 2021 from the same sites as
135 fish. We collected approximately 0.5 kg of surface and near surface sediment at each site using
136 plastic spatulas and bags for the storage. Samples were transported to the laboratory at 4 °C
137 and then stored at – 20 °C.

138

139 2.3. Procedure of biometric analysis, tissue dissection and storage

140 Fish were anesthetized immediately after the sampling using tricaine methane sulphonate
141 (MS 222, Sigma Aldrich) according to the Ordinance on the protection of animals used for
142 scientific purposes (NN 55, 2013). Firstly, basic biometric data (total length and body weight)
143 were taken for each fish. Further, organ (liver, gonads, viscera) dissection was performed on
144 ice to collect the data for calculation of biometric indices. Gonads also served for the gender
145 determination. The posterior part of the intestine was isolated, cleaned from the exterior fat and
146 gut content, and examined for the presence of acanthocephalans. The gut content of each fish
147 was separated and stored. After isolating and quantifying acanthocephalans from the intestines
148 of each fish, the prevalence and intensity of infection were calculated. All samples were

149 subsequently stored in liquid nitrogen until transportation to the laboratory, where they were
150 kept at $-80\text{ }^{\circ}\text{C}$ until analyses.

151

152 2.4. Acid digestion of biological samples and sediments

153 *2.4.1. Biological samples*

154 Prior to acid digestion, acanthocephalans from the same fish were pooled together to
155 enable reliable measurements (3-24 individuals). Fish intestine and acanthocephalans were
156 digested with HNO_3 (Rotipuran® Supra 69%, Carl Roth, Germany) and 30% H_2O_2 (Suprapur,
157 Merck, Germany) at $85\text{ }^{\circ}\text{C}$ for 3.5 h, as described by Mijošek et al. (2022). Gut contents of fish
158 were digested under the same conditions using the mixture of HNO_3 (Rotipuran® Supra 69%,
159 Carl Roth, Germany) and HF (Suprapur, Merck, Germany).

160 *2.4.2. Sediments*

161 Sediment samples were air-dried, homogenized in agate mill and sediment subsamples
162 were digested in a two-step digestion procedure performed using microwave oven (Multiwave
163 ECO, Anton Paar, Austria). Mixture of 4 mL HNO_3 (65%, p.a., Kemika, Croatia), 1 mL HCl
164 (TraceSELECT, Fluka, Germany), and 1 mL HF (TraceSELECT, Fluka, Germany) is used in
165 the first step, followed by the addition of 6 mL H_3BO_3 (Fluka, Switzerland), as described by
166 Fiket et al. (2017).

167

168 2.5. ICP-MS measurements

169 Concentrations of trace and macroelements in water, organisms and GC were measured
170 using an Agilent 8800 instrument (Agilent Technologies, USA) as presented in Table S2. To
171 reduce interference, cell (helium and hydrogen) and reaction (oxygen) gases were used.
172 Ultrapure Mili-Q water (Barnstead Smart2Pure Water Purification System, Thermo Scientific,
173 Langenselbold, Germany) was used for preparation of standards and samples, and washing the

174 dishes and disposable plastics. Acidified water samples underwent analysis without dilution,
175 whereas the biological samples were diluted 10-13 times with a solution containing 1% (v/v)
176 of purified nitric acid [HNO₃, 65%, p.a., Merck; purified by quartz sub-boiling distillation
177 using the Milestone SubPUR system (Milestone S.r.l., Sorisole, Italy)] and hydrochloric acid
178 (HCl, 30%, Suprapur®, for trace analysis, Supelco®). An internal standard solution containing
179 3 µg L⁻¹ of internal standards (Ge, Rh, Tb, Lu, and Ir) (SCP Science, Canada) was used to
180 correct instrumental drifts and plasma fluctuations. Standard solutions used for external
181 calibration were prepared from individual PlasmaCAL Single-Element Standard solutions
182 which were obtained from SCP Science, Quebec, Canada. Four standard certified reference
183 materials [IAEA-407 (fish tissue), ERM-BB422 (fish muscle), ERM-CE278 (mussel tissue),
184 NIST SRM 1577a (bovine liver)] were analyzed with the samples as part of quality control.
185 The accuracy of the method for the elements in the referent biological samples mostly fell
186 within a range of ±10%, with the recoveries varying from 90.4% for rubidium (Rb) to 110%
187 for cadmium (Cd). Detailed information on the limits of detection (LODs) and the recoveries
188 for each element and certified reference material in water and biological samples are presented
189 in Tables S3 and S4.

190 Total element concentrations in sediments were measured using triple quadrupole
191 inductively coupled plasma mass spectrometer (ICP-QQQ, Agilent 8900, USA). Analytical
192 quality control was performed by simultaneous analysis of procedural blanks and certified
193 reference materials of stream sediment NCS DC 73309 (GBW 07311) (China National
194 Analysis Center for Iron and Steel, China). Recoveries in the referent sample were in range 91-
195 104%, depending on the element.

196

197 2.6. Data processing and statistical methods

198 *2.6.1. Biological and chemical data*

199 Prevalence (number of infected fish) and mean intensity of infections (number of
200 parasites per fish individuals) were calculated to describe acanthocephalan epidemiology.
201 Bioconcentration factors (BCFs) were calculated as ratios between the element concentrations
202 in acanthocephalans and fish intestine. Ratios between metal(loid) concentrations in fish
203 intestine and gut contents, as well as in acanthocephalans and gut contents, were calculated as
204 percentages and considered as the indication of possible meta(loid) bioavailability from the
205 dietborne uptake.

206 Metal(loid) concentrations are presented as average values \pm standard deviation and
207 expressed in $\mu\text{g L}^{-1}$ for water, $\mu\text{g kg}^{-1}$ or mg kg^{-1} of wet weight (w.w.) for organisms and GC,
208 and mg kg^{-1} or g kg^{-1} d.w. for sediments.

209 *2.6.2. Statistical methods*

210 Statistical analyses were done in SigmaPlot 11.0 (Systat Software, USA) and Microsoft
211 Office Excel 2007. Metal(loid) concentrations in fish, acanthocephalans, GC, and water were
212 compared between three sample locations using Kruskal-Wallis one-way analysis of variance
213 on ranks, followed by Dunn's or Tukey's tests. Mann-Whitney U test was applied to test the
214 data variability between the two seasons. Due to normal distribution of data on sediment
215 samples, spatial differences were tested using one-way ANOVA using Holm-Sidak method.
216 Differences were considered significant when $p < 0.05$. Correlations between parameters were
217 calculated using Spearman correlation analysis and correlation coefficients and levels of
218 significance were indicated in the text and all are reported in tables in supplementary material.

219

220 **3. Results and discussion**

221 3.1. Main biometric characteristics of fish and acanthocephalans

222 No significant spatial differences were observed in total length (TL), total body mass
223 (TM), and biometric indices of fish. Nevertheless, there were notable seasonal differences in

224 TL and TM, with higher values observed in autumn, as the spawning period of this species,
225 than in spring at all locations (Table 1). GSI values were also higher in autumn at all locations.
226 However, these seasonal differences were not statistically significant at KNP, likely as a
227 consequence of sporadic sampling of a few small, reproductively immature individuals and
228 generally lower number of fish captured at this site in the spring. There were no significant
229 differences depending on the fish sex.

230 All fish were infected with acanthocephalan *D. truttae* and there were no significant
231 spatial differences in mean intensity of parasites in any season. The mean intensities and
232 number of parasites were higher in autumn than in spring at all locations (Table 1). Comparable
233 epidemiological characteristics have already been reported for the period 2005-2016 (Vardić
234 Smrzlić et al., 2013; Filipović Marijić et al., 2022; Mijošek et al., 2022), probably all related to
235 the life-cycle of gammarids, intermediate hosts of acanthocephalans. Gammarids reproduce
236 mainly in late summer and autumn, leading to a higher abundance of acanthocephalans due to
237 higher presence of gammarids in fish food.

238 To describe the relationship of this host-parasite system, we examined the correlations
239 of parasite abundance and fish size and condition (FCI). Given the strong correlation between
240 TL and TW ($r = 0.887-1.000$, $p < 0.001$, depending on the site and season), we opted to utilize
241 TM as the variable to describe fish size in order to prevent issues with multicollinearity. For
242 this purpose, all data were tested together. A significant positive correlation ($r = 0.478$, $p < 0.001$)
243 was established between the abundance of parasites and fish TM, while no correlation was
244 observed between abundance and FCI, confirming that acanthocephalans do not have negative
245 effect on the fish fitness. Similar to findings of Vardić Smrzlić et al. (2013) and Mijošek et al.
246 (2022) for *D. truttae*, we observed higher prevalence of infection in male than in female fish,
247 but not statistically significant.

248

249 3.2. Main environmental conditions

250 *3.2.1. Concentrations of dissolved metal(loid)s in water*

251 The recent comprehensive study on the dissolved metal(loid)s in water samples from
252 eight locations along the upper stretch of the Krka River (Mijošek et al., 2023), revealed that
253 industrial wastewaters from the screw factory (IWW) represent the most significant source of
254 metal(loid)s in the Krka River primarily through the Orašnica River, a tributary that is directly
255 influenced by the wastewaters due to potential spillover during heavy rains. Metal(loid)
256 concentrations were also increased near the municipal outlet of the Town of Knin (KRK) in
257 comparison to the river source (KRS) and location in national park (KNP).

258 At the three locations where fish were sampled, a few distinct patterns of metal(loid)s in
259 the dissolved fraction were observed: As, Fe, Mg, Mn, Na, and Rb were predominantly the
260 highest at KRK, while Ba, Ca, Mo, Sr, and V were the highest at KNP, and Tl was the highest
261 at KRS. (Table S1). Seasonal differences between spring and autumn were not clearly
262 distinguishable between the three sites, which is in line with results of our comprehensive data
263 covering metal(loid) concentrations across eight locations, with no significant differences
264 observed between spring and autumn (Table S1, Mijošek et al., 2023). Furthermore, spatial
265 distribution of metal(loid)s coincided with water physico-chemical parameters, which were
266 below good at KRK due to high concentrations of nutrients and COD in both seasons.
267 Regarding Krka River source (KRS) and location in national park (KNP), mostly very good
268 ecological status was indicated (Mijošek et al., 2023; Šariri et al., 2024). The same pattern was
269 observed for meta(loid) concentrations, wherein the lowest concentrations were found at KRS,
270 followed by the highest concentrations of many elements at KRK and a subsequent gradual
271 decrease at KNP, with the exception of Ba, Ca, Mo, Sr and V, which increased to their
272 maximum concentrations at KNP (Table S1). Cukrov et al. (2008) verified the existence of a
273 self-purification mechanism in the Krka River, starting in the Brljan Lake and progressing

274 through subsequent tufa barriers, resulting in decreased metal(loid) concentrations in water
275 downstream of the pollution sources. Moreover, groundwater input of clean water from the
276 Zrmanja River also contributes to the decrease in water trace metal concentrations in a
277 downstream way of contamination in the Krka River (Cukrov et al., 2008). Hence, elevated
278 concentrations of many elements at KNP in comparison to KRS suggested potential future
279 threats to this protected area, especially since measuring contaminants solely in water is
280 inconclusive due to water discharge fluctuations and short resident times, making it a less
281 reliable method of assessment (Salati and Moore, 2010; Algül and Beyhan, 2020).

282 *3.2.2. Metal(loid) concentrations in sediments*

283 Concentrations of majority of elements in sediment samples showed significant site-
284 specific differences (Table S5). Potassium, Na and all trace elements in sediments exhibited
285 the highest concentrations at the location in close proximity to Brljan Lake in KNP (Table S5).
286 The sole deviations from this pattern were noted for Mg and Ca, with the highest average
287 concentrations at KRS and KRK, respectively. River source is a reference site without the
288 known pollution sources, characterized by the strong flow and low sedimentation rate, which
289 explains the lowest concentrations of most elements at this site, both in sediments and water.
290 However, only few kilometers downstream from the KRS, municipal wastewaters of the Town
291 of Knin cause the increase in metal(loid) concentrations, which is the highest in downstream
292 lake systems which serve as sink for metals due to intensive sedimentation and low flow (Chon
293 et al., 2012). The water flow rate decreases in a downstream direction and lowers the
294 transmission strength and carries smaller sediments particles which sink and deposit over
295 longer time, and during such extensive sedimentation metal(loid)s bind to these particles and
296 precipitate in sediments, consequently reducing metal(loid) concentrations in water in the
297 process of self-purification (Cukrov et al., 2008).

298 It is obvious that negative influence of anthropogenic activities is observed in the area of
299 the national park according to the sediment analyses (Table S5), which act as a sink, but also
300 the source of many contaminants, including metal(loid)s (Chon et al., 2012). Metal(loid)s are
301 not permanently bound to sediments and can be released into the water at any time depending
302 on pH, temperature or flow which affect metal balance and remobilization between the
303 sediments and water and consequently, cause possible toxic effects in biota. Therefore, both
304 water and sediments, in addition to food can serve as sources of metal(loid)s for organisms,
305 which will be discussed further.

306

307 3.3. Metal(loid) concentrations in biota

308 *3.3.1. Metal(loid) accumulation in fish intestine*

309 Metal(loid) accumulation in fish intestine followed the order $K > Na > Mg \geq Ca$ for
310 macroelements and $Zn > Fe > Rb > Cu \geq Se > Mn > Sr > Cd \geq Co \geq As > Mo > Tl > Ba \geq Cr > V \geq Hg > Pb \geq Cs$
311 for trace elements (Figs. 2-5), similar to the brown trout sampled in 2015/2016 (Mijošek et al.,
312 2022). Our findings are consistent with prior research that reported the highest levels of
313 essential elements as Zn, Fe, and Cu in the intestines of various fish species, including perch
314 (Sures, 2002), European chub (Filipović Marijić and Raspor, 2010), or barbel (Nachev and
315 Sures, 2016). When comparing the current state of accumulation of elements in brown trout
316 intestines with that of previous research that focused on two locations in Krka River (KRS and
317 KRK), it is evident that Ca, Cs, K, Mg, Na, Rb and Tl remained relatively constant, whereas
318 bioaccumulation of Cd, Co, Cu, Fe, Mn, Se, Sr and Zn increased with time (Figs. 2-5, Mijošek
319 et al., 2022). Only Pb concentrations were found to be around 3-5 times lower in the present
320 research compared to previous study, owing to possibly lower concentrations of this element
321 in sediment or gut content samples which are related to uptake sources and were not analyzed
322 in the previous study. Given the widespread utilization of Cu, Fe, Mn, and Zn in industrial,

323 municipal, and agricultural processes, our findings suggest the intensifying impact of
324 wastewaters on biota. This is particularly worrisome in unexplored region of the national park,
325 where fish frequently exhibited maximum concentrations of certain elements compared to
326 locations upstream. These patterns are consistent with the analysis of metal(loid)s in water
327 samples, which confirmed that the majority of values in 2021, particularly for Cr, Cu, Fe, and
328 Mn, were higher than those recorded between 2004 to 2015, probably all as a consequence of
329 industrial and agricultural activities in the region (Mijošek et al., 2023).

330 We further investigated the potential influence of size (represented by TM) and sex on
331 metal(loid) accumulation in the intestine. The influence of FCI was not considered since the
332 differences in FCI between sites and seasons were negligible and not significant (Table 1).
333 Spearman's correlation coefficients for the associations TM and metal(loid) concentrations in
334 fish intestine are presented in Table S6. Only a few notable correlations were observed between
335 the concentrations of elements in the intestine and TM. A statistically significant ($p < 0.05$ in all
336 cases) decrease with increasing TM was found for V at KRK ($r = -0.886$) in spring, Cd and Cs
337 at KRS ($r = -0.767, -0.833$) in spring, and Cs at KNP in autumn ($r = -0.786$). Conversely,
338 significant increase was detected for Mg at KRS in spring ($r = 0.667$) and As at KRK in autumn
339 ($r = 0.609$). Fish often show variations in metal-size associations, usually as a result of
340 differences in their metabolic rates, detoxification systems or feeding habits (Nikolić et al.,
341 2021). The influence of fish sex on bioaccumulation of metal(loid)s in fish intestine was
342 evaluated using pooled data for each sex. Statistical analysis showed there were no significant
343 differences, although average values of majority of elements were slightly higher in females,
344 possibly due to factors such as food preferences or specificities of the reproductive cycle
345 (Rajkowska and Protasowicki, 2013). Overall, the impact of fish biometric parameters on
346 metal(loid)s accumulation in the intestine of brown trout was mainly negligible.

347 *3.3.2. Metal(loid) accumulation in acanthocephalans*

348 Macroelements followed the order $K > Na \geq Ca > Mg$, and trace elements:
349 $Zn > Fe > Cu > Mn > Sr \geq Rb > Cd \geq Tl \geq Se > Pb > Ba > As > Co \geq Hg \geq Cr \geq Mo \geq V > Cs$ (Figs. 2-5), as in
350 previous research from 2015/2016 (Mijošek et al., 2022). Unlike in fish, concentrations of the
351 majority of elements did not vary between the studies, indicating the absence of temporal
352 trends, with the exception of Cd, Cu, Mn, and Zn, which were mostly higher in the current
353 research, implying continuous contamination impact of the screw factory. Our two studies
354 present the only data on metal(loid) accumulation in this endemic acanthocephalan species and
355 both indicated the certain level of environmental deterioration over time. Additionally,
356 concentrations of As, Ba, Cr, Hg, Mo and V in *D. truttae* were now measured for the first time.

357 Contrary to positive association between parasite abundance and fish size, we observed
358 a negative correlation between metal(loid)s accumulation in acanthocephalans and fish TM for
359 many metal(loid)s, especially in autumn (Table S6). Hence, significant negative correlations
360 were observed for Mn at KRS ($r = -0.800$; $p < 0.01$) in the spring, as well as for Tl at KNP ($r = -$
361 0.750 ; $p < 0.05$), Cd ($r = -0.700$; $p < 0.05$), Pb ($r = -0.624$; $p < 0.05$), Rb ($r = -0.886$; $p < 0.05$) and Tl
362 ($r = -0.627$; $p < 0.05$) at KRK, and Ba ($r = -0.661$; $p < 0.05$), Co ($r = -0.745$; $p < 0.05$), Fe ($r = -0.709$;
363 $p < 0.05$), Pb ($r = -0.767$; $p < 0.05$), Cd ($r = -0.758$; $p < 0.01$), Tl ($r = -0.806$; $p < 0.01$), Cu ($r = -0.830$;
364 $p < 0.001$) and Se ($r = -0.879$; $p < 0.001$) at KRS in the autumn. Higher concentrations of
365 metal(loid)s in acanthocephalans from smaller fish may be associated with higher metabolic
366 and feeding rates in smaller specimens, both of which decline with fish aging (Farkas et al.,
367 2003).

368 Analyses of associations between metal(loid)s accumulation in the intestines of brown
369 trout and in acanthocephalans failed to uncover many consistent correlations when data were
370 stratified by season and location. However, upon analyzing the pooled data, many positive
371 correlations were identified [$p < 0.05$ for As ($r = 0.369$), and Se ($r = 0.360$), $p < 0.01$ for Cd
372 ($r = 0.441$), Hg ($r = 0.400$), Mn ($r = 0.468$), Mo ($r = 0.372$), Pb ($r = 0.331$) and Zn ($r = 0.409$), $p < 0.001$

373 for Ba ($r=0.615$), Ca ($r=0.600$), Cs ($r=0.794$), Cu ($r=0.478$), Rb ($r=0.660$), Sr ($r=0.819$; p), Tl
374 ($r=0.511$), and V ($r=0.519$), suggesting that fish and parasites exhibit similar biological
375 responses. All results on this association are presented in Table S7.

376 Although various studies (Sures, 2002; Filipović Marijić et al., 2013; Brázová et al.,
377 2015) demonstrated that acanthocephalan-infected fish have lower metal concentrations than
378 the uninfected ones, the 100% prevalence of acanthocephalans in our research prevented us
379 from testing this hypothesis (Table 1). However, we observed notable differences in the
380 accumulation of metal(loid)s in the intestine based on the number of acanthocephalans in fish.
381 In particular, statistically significant inverse relationships between the abundance of
382 acanthocephalans in fish and the concentrations of As ($r= -0.545$; $p<0.001$), Cr ($r= -0.571$;
383 $p<0.001$), Mn ($r= -0.446$; $p<0.001$), Se ($r= -0.523$; $p<0.001$), V ($r= -0.507$; $p<0.001$), Mo ($r= -$
384 0.392 ; $p<0.01$), Ba ($r= -0.326$, $p<0.05$), Cu ($r= -0.351$, $p<0.05$), and Fe ($r= -0.339$, $p<0.05$) in
385 the intestine of fish was established. Correlation analysis is presented in Table S8. This
386 suggests that fish with greater infection levels absorbed lower concentrations of these elements
387 via the intestinal wall, as consistent with the possible protective and beneficial role for their
388 fish hosts (Sures, 2002; Nachev and Sures, 2016; Mehana et al., 2020).

389 Continuing to analyze the potential impact of acanthocephalan infrapopulation sizes on
390 metal(loid) accumulation mechanisms within parasites (Table S8), we observed that larger
391 infrapopulations had lower concentrations of Co ($r= -0.545$; $p<0.001$), Cr ($r= -0.500$; $p<0.001$),
392 V ($r= -0.646$; $p<0.001$), Zn ($r= -0.475$; $p<0.001$), As ($r= -0.414$; $p<0.01$), Fe ($r= -0.453$;
393 $p<0.01$), Mg ($r= -0.406$; $p<0.01$), Mo ($r= -0.466$, $p<0.01$), Ba ($r= -0.333$, $p<0.05$) and Se ($r= -$
394 0.361 , $p<0.05$) compared to smaller infrapopulations. Sures et al. (1999) previously reported
395 that concentrations of several elements within the parasite *Acanthocephalus lucii* decreased as
396 the infrapopulation of *A. lucii* in fish increased. Similar findings were reported by Brázová et
397 al. (2015), who recorded that a high intensity of infection was associated with lower heavy

398 metal concentration in *A. lucii* and *Proteocephalus percae*, and by Hassanine et al. (2018)
399 regarding the inverse relationship between metal concentrations in the acanthocephalan
400 *Sclerocollum saudii* and the size of its infrapopulation in fish *Siganus rivulatus*. It is still
401 unclear whether our findings support the traditionally accepted concept of intraspecific
402 competition among acanthocephalans for the absorption of metals from fish intestine (Sures,
403 2002; Hassanine and Al-Hasawi, 2021) or the more recent hypothesis of biodilution of
404 elements in the fish host proposed by Duarte et al. (2020). The latter concept proposes that in
405 larger parasite populations, elements may be dispersed among all parasite individuals, leading
406 to reduced metal(loid) concentrations in both parasites and fish. In the investigation of the
407 accumulation of 12 trace metals in the muscle, intestine, and liver of the white mullet and its
408 intestinal parasites, Leite et al. (2023) also noted that the size of the parasite infrapopulation
409 had an effect on the concentrations of trace metals in the parasites, with smaller
410 infrapopulations accumulating a greater quantity of metals. Regarding the host, fish with a
411 more severe parasitic infection had reduced concentrations of Cd exclusively in the muscle
412 tissue, while no significant differences were detected with regard to other tissues or metals.
413 They also explained this phenomenon by biodilution, resulting from the increased biomass of
414 the host-parasite system and the fact that after metal is excreted in the intestine, reabsorption
415 by fish does not occur because metal is captured by the parasite at the infection site.

416 Therefore, as parasites can be found in every free-living organism in all ecosystems there
417 is a need of investigating their impact on environmental health and biological responses of
418 organisms (Timi et al., 2020). Our results confirmed that accumulation in acanthocephalans
419 may have implications for accumulation levels in fish, that they have high accumulation
420 potential and response fast, as well as possible negative and positive effects on intermediate
421 and final hosts (Sures, 2002; Sures et al., 2017). However, it is challenging and almost
422 impossible to distinguish in field conditions acanthocephalan from environmental impact

423 because parameters as season, abundance of intermediate hosts, sex of the hosts, as well as
424 their biometric characteristics may sometimes impact epidemiological characteristics of
425 acanthocephalans and cannot always be excluded. That highlights the complexity of the host-
426 parasite relationship, as well as of the assessment of contamination exposure. Still, good
427 freshwater ecosystem management plans and monitoring programs should always contain
428 physico-chemical factors, water and sediment analyses, fish, but also parasites to interpret the
429 data and results from the field studies as accurate as possible.

430

431 *3.3.3. Spatial differences in metal(loid) accumulation*

432 Fish intestine and acanthocephalans showed variable spatial differences which were not
433 always statistically significant but could be mostly grouped in four patterns: 1) elements with
434 higher concentrations at the contaminated site, KRK; 2) elements with higher concentrations
435 at the reference site, KRS; 3) elements with higher concentrations at the site in national park,
436 KNP; 4) elements with unclear trends (Figs. 2-5).

437 Statistically significant differences in metal(loid) accumulation between sites in the fish
438 were observed for Cs, Mn, Rb and Se during both seasons. For As, Co, Fe, Tl, and Zn, these
439 differences were evident only in spring, whereas for Cd and Hg only in autumn (Figs. 2-5).
440 The highest levels of certain elements were recorded at different locations: 1) As, Mn, Se, Zn
441 at KRK; 2) Cd, Cs, Rb, Tl at KRS; and 3) Co, Hg, Mn at KNP. Similar patterns were previously
442 documented for brown trouts from KRS and KRK collected in 2015/2016, with no sampling
443 conducted at KNP during that period (Mijošek et al., 2022). These patterns could not be fully
444 explained by uptake from water or sediments alone, suggesting the importance of long-term
445 dietborne metal(loid) uptake and feeding preferences. Namely, As, Se and Zn were found in
446 higher concentrations in fish from KRK (Fig. 2), but not in the water or sediment samples from
447 that site (Tables S1 and S5). Similarly, Cs and Rb concentrations in fish (Fig. 3b and 3c) did

448 not correspond to their levels in water or sediments; in water, the levels were either similar
449 across sites or the highest at KRK (Table S1) whereas for sediments, they were highest at KNP
450 (Table S5). Bioaccumulation of Co, Fe, Hg and Mn in fish could mostly be explained by their
451 uptake and remobilization from sediments, while concentrations of Tl and Cd in fish from the
452 reference site seemed to be more closely related to their concentrations in water than sediment
453 samples (Tables S1 and S5). Concentrations of Cd and Tl tended to be higher in fish from the
454 KRS (Fig. 3), possibly due the prevalence of Cd-enriched limestones and dolomites in Dinaric
455 karst (Cukrov et al., 2008). In addition to natural and anthropogenic causes, food preferences
456 can significantly affect metal accumulation in the intestine, since dietborne metal uptake may
457 be of equal or greater importance than waterborne uptake for some elements, including Tl
458 (Lapointe and Couture, 2009).

459 Acanthocephalans showed significant spatial differences for Cs, Fe, and Sr in both
460 seasons, while variations for Cd, Co, Hg, Mo, Na, and V were apparent during only one season
461 (Figs. 2-5), which partially corresponded to patterns in the intestines. Similar to previous
462 research of Mijošek et al. (2022), the highest levels of certain metals in fish parasites were
463 found at the following locations: 1) Co and Fe at KRK; 2) Cd and Cs at KRS; and 3) Mo, Na,
464 Sr, and V at KNP. In contrast to the accumulation of Cd, Co, Cs, and Hg in parasites, which,
465 similar to the intestinal accumulation, was largely unrelated to exposure from water and
466 sediments, the highest accumulation of Mo, Sr, and V at KNP was concurrent with their highest
467 levels in water, sediment and GC samples from that site (Tables S1, S5 and S9). Patterns of
468 other elements might be associated with long-term feeding preferences of their fish hosts or
469 natural geological background. Although there were notable differences in the average values
470 of other elements in acanthocephalans across different sites, these variations were not
471 statistically significant due to the high variability and high standard deviations in metal(loid)
472 concentrations among individual acanthocephalans, as already stated for different

473 acanthocephalan species (Filipović Marijić et al., 2013; Nachev and Sures, 2016; Mijošek et
474 al., 2022).

475 *3.3.4. Temporal differences in metal(loid) accumulation*

476 Seasonal variations of metal(loid) concentrations in the intestine of brown trout were
477 mostly not uniform and could not be extrapolated to all locations (Figs. 2-5). Nevertheless,
478 more elements showed higher levels in spring compared to autumn, with significant differences
479 observed for Ca, Hg, Mg, Mn, Se, and Tl at KRS, As, Co, Hg, Mn, and V at KRK, and As, Co,
480 Cu, Fe, Mn, Mo, and Se in KNP. Conversely, greater accumulation of Cs at KRK, Na at KNP,
481 and Zn at KRS was observed during the autumn (Figs. 2-5). Various biotic and abiotic factors,
482 in addition to exposure conditions, can influence seasonal variability in metal accumulation.
483 For example, differences in growth, metabolism rate or water parameters like temperature, pH,
484 or water hardness may influence bioaccumulation in organisms. Further, food type or feeding
485 rates can also significantly affect bioaccumulation (Farkas et al., 2003) and these differences
486 were clearly observed during fish dissection. Specifically, we observed high gut fullness index
487 in nearly all fish during spring. The feeding intensity was lower in autumn with the higher
488 proportion of half-empty to empty stomachs. Greater diversity of the gut contents was also
489 observed in spring, which contributed to the higher metal(loid) concentrations in fish GC in
490 that season as will be further discussed. This observation is in accordance with research
491 reporting maximum feeding rates of trouts in spring (Debeljak, 1986; Kara and Alp, 2005).
492 Fullness indices of the brown trouts were also already found to be the lowest in autumn,
493 increasing from winter to summer (Kara and Alp, 2005).

494 Higher accumulation in spring than autumn was also observed in acanthocephalans,
495 although less pronounced and engaged different elements than in fish intestines. Namely,
496 concentrations of Co, Mo, Pb, and V at KRS, Mo and V at KRK, and Co at KNP were
497 significantly higher in spring than in autumn (Figs. 2, 4 and 5). Potassium was significantly

498 higher in autumn compared to spring in acanthocephalans from KRS, while other elements did
499 not display significant seasonal variations (Figs. 2-5). According to Nachev and Sures (2016),
500 seasonality in metal accumulation in parasites can be explained by acanthocephalan
501 transmission, maturation in the intestine of the hosts, or by physiological or behavioral changes
502 of their final hosts, such as mobility or alterations in food preferences.

503 3.3.5. Bioconcentration factors (BCFs)

504 To define the relation of metal(loid) concentrations in acanthocephalans and fish
505 intestine, we calculated BCFs (Table 2). Due to the different life spans of acanthocephalans
506 and fish, high BCFs indicate recent metal exposures, while lower ratios represent continuous,
507 longer exposure (Sures et al., 1999).

508 Our research supported findings of superior efficiency of metal(loid) accumulation in
509 acanthocephalans compared to fish (Filpović Marijić et al., 2013; Nachev and Sures, 2016;
510 Sures et al., 2017; Mijošek et al., 2022). Acanthocephalans showed higher accumulation of As,
511 Ba, Cd, Cr, Cu, Hg, Mn, Pb, Sr, Tl, Ca, and Na in all sites and seasons, and of Co, Fe, and V
512 in majority of cases (Table 2). The only elements with lower accumulation in acanthocephalans
513 than in fish intestine ($BCF < 1$) were Cs, Mo, Rb, Zn and K, with the occasional inclusion of
514 Co, Se, V and Mg in a few instances (Table 2). This is in accordance with a recent research
515 conducted on the Krka River, which found that the same few elements accumulated more in
516 fish intestines than in acanthocephalans (Mijošek et al., 2022). Both investigations confirmed
517 remarkably efficient accumulation of potentially highly toxic elements (Pb, Cd and Tl), which
518 goes in favor of possible protection of the infected fish (Sures, 2002; Filipović Marijić et al.,
519 2013; Mehana et al., 2020; Molbert et al., 2020). Moreover, high BCF values observed for Pb,
520 Cd, Tl, Cu, Ba, Mn, Sr, Ca, and Hg (Table 2) suggest the likelihood of recent high exposure to
521 these elements in the Krka River.

522 Although we expected to observe some significant spatial or temporal trends regarding
523 BCF values, the number of distinctive patterns was limited (Table 2). Still, the maximum BCFs
524 in both seasons for As, Hg, Mn, and Se were observed at KRS, for Ba and Ca at KRK, and for
525 Sr at KNP, showing that used acanthocephalan species provides a valuable biological response
526 even at the site of the minimal metal(loid) exposure, KRS. Regarding seasonal changes, only
527 the BCFs for Fe, Na, and Sr were higher in spring compared to autumn at all sites, whereas the
528 opposite trend was seen for As, Mn and Pb (Table 2). These trends could not be associated with
529 the metal(loid) exposure from water or sediments, but rather to factors such as age, physiology,
530 reproductive stage or size of the acanthocephalans or fish host (Filipović Marijić et al., 2013;
531 Sures et al., 2017).

532 The BCF ranges for most elements in the current research were comparable to the
533 2015/2016 study on the same host-parasite system (Table 2, Mijošek et al., 2022). However,
534 the BCF values for Pb were 2 to 8 times greater in the current study, which can be attributed to
535 the aforementioned decrease in Pb concentrations in fish intestines nowadays. In summary,
536 while there is a lack of additional research pertaining to this particular ecosystem and species,
537 our data suggests that *D. truttae* is a promising indicator, even in ecosystems characterized by
538 low to moderate metal(loid) exposure, such as the karst Krka River.

539

540 3.4. Metal(loid) accumulation in gut contents (GC)

541 Dietborne metal(loid) uptake was considered through measuring metal(loid)
542 concentrations in GC of fish. The sequences of macroelements (Ca>Na>K>Mg) and trace
543 elements (Fe>Zn>Sr>Mn>Cu>Ba>V>Cr>Co>Rb>Pb>Cd>As>Se>Mo>Cs>Tl>Hg) (Table
544 S9) were notably different in comparison to fish intestine (Figs. 2-5), indicating that metal(loid)
545 accumulation depends on the bioavailability of metal(loid)s in fish diet. Further, fish have the
546 ability to regulate metal uptake from food into the tissue depending on the specific

547 requirements of organisms (Clearwater et al., 2000), which is reflected in different efficiency
548 of metal absorption. Similar patterns and differences in metal accumulation (Cd, Cu, Fe, Mn,
549 and Zn) between fish intestine and GC have been reported for the European chub from the Sava
550 River (Filipović Marijić and Raspor, 2012, 2013). As brown trout is an omnivorous species,
551 the gut contents included a variety of species including insect larvae, detritus, plants to
552 gammarids or small mollusks. This led to high variations in metal(loid) concentrations among
553 individual fish (Table S9). For example, the higher average contents of Ca and Sr in GC
554 compared to fish intestine is associated to the prevalence of gammarids as a favored food source
555 for many fish, which exoskeletons are particularly rich in Ca and Sr (Greenway, 2008), but
556 only a small portion of these elements is bioavailable to organisms. To our knowledge, this is
557 the first report on metal(loid)s in GC of brown trouts, so comparison with previous data was
558 not possible.

559 Spearman correlation analyses revealed sporadic associations between concentrations of
560 metal(loid)s in GC and fish intestine or acanthocephalans. More frequently, we observed
561 negative correlation between metal(loid)s in GC and fish size (TM), which corresponds to
562 higher metabolic needs and feeding rates during the development in early stages (Farkas et al,
563 2003).

564 *3.4.1. Spatial differences*

565 Statistically significant spatial differences were observed for most elements, except Ba,
566 Ca, Mg, Na, Se, Sr and Tl (Table S9). The levels of most of the remaining elements were
567 highest in GC of fish from KNP, similar to spatial distribution observed in sediment samples
568 (Tables S5 and S9). During dissection, particles of sediments could be observed in GC of many
569 fish, suggesting that sediments play a significant role in the absorption of metals through the
570 diet and may pose a threat to organisms in the area of the national park and sensitive lake
571 systems in case of the prolonged exposure. Exceptionally, Cd concentrations in GC were

572 highest at KRS, while Cu concentrations in GC were highest at KRK, which aligns with the
573 spatial distribution of Cu in fish intestine and acanthocephalans (Figs. 2d and 3a). Overall,
574 metal(loid) concentrations in GC of fish were typically the lowest at KRS, demonstrating the
575 lowest metal(loid) exposure at this location, which corresponded to water and sediment
576 samples.

577 *3.4.2. Temporal differences*

578 The majority of elements in GC had higher concentrations in spring than autumn, with
579 the highest number of statistically significant differences at KRS (Table S9). As already
580 mentioned, higher gut fullness indices were found in spring, a season characterized by
581 maximum feeding rates for this species, as well as greater variability of the prey, which led to
582 the increased metal(loid)s accumulation in spring. A similar pattern was also observed in
583 parasites and intestine, which further suggests that the uptake of metal(loid)s through the diet
584 influences the responses of organisms, prompting us to investigate their relationships further.

585 *3.4.3. Relationships between metal(loid) concentrations in fish intestine and acanthocephalans* 586 *to gut contents*

587 To compare levels of metal(loid)s in biological samples to those found in GC, as an
588 important uptake route, we calculated average ratios between the concentrations of metal(loid)s
589 in the intestine of fish and GC (INT/GC), as well as in acanthocephalans and GC (AC/GC)
590 (Table S10). With this approach, we were able to evaluate and estimate the potential
591 bioavailability of metal(loid)s from food to organisms. However, it is important to keep in mind
592 that this is only an estimate since the metal(loid)s found in the GC reflect the source of
593 metal(loid)s in the food at the time of sampling, whereas organisms accumulate metal(loid)s
594 over a longer period of time and exhibit a more chronic response.

595 Nevertheless, it can be presumed that the bioavailability of As, Ba, Ca, Cr, Cu, Fe, Mn,
596 Pb, Sr, and V from food to the fish intestine may be minimal, since the majority of the INT/GC

597 ratios for these elements were <15%, suggesting that their absorption is physiologically
598 regulated and not dependent on their quantity in the food source. Although concentrations of
599 these elements in GC were predominantly highest at KNP, the calculated ratios were either
600 similar across all locations or the lowest at KNP (Table S10). With the exception of Cd and Pb,
601 the lowest values of the AC/GC ratios were found once more for As, Ba, Ca, Cr, Fe, Mn, Sr,
602 and V, confirming low bioavailability of these elements from food. Low values mostly up to
603 50% were also observed for Co, Cs, and Mo for both fish intestine (INT/GC) and
604 acanthocephalans (AC/GC) (Table S10), suggesting that although concentrations of these
605 elements in the GC are elevated (Table S9), their burden is significantly lower in fish intestine
606 and parasites. This indicates that a large portion of these elements is excreted via the faeces
607 (Dallinger and Kautzky, 1985).

608 Thallium, Se, Hg, Rb, K and Zn exhibited remarkable luminal absorption efficiency, as
609 evidenced by their substantially higher concentrations in the intestine and acanthocephalans
610 compared to GC (Table S10). In addition, Cd, Cu, and Pb were also accumulated more
611 efficiently in acanthocephalans than GC and fish intestine showing they may selectively
612 accumulate certain elements over others, potentially influencing the overall element
613 distribution within the fish host and providing a protective role for the fish host when exposed
614 to potentially toxic elements (Sures, 2002). Understanding these differences in element
615 accumulation is important since it can provide valuable insights into the dynamics of host-
616 parasite interactions and their impact on element cycling in aquatic ecosystems. To our
617 knowledge, there is no available research that compares metal(loid) accumulation in
618 acanthocephalans and fish gut content. However, Dallinger and Kautzky (1985) published data
619 on Cd, Cr, Cu, Mn, Pb and Zn concentrations in the intestine and GC of species *Salmo gairdneri*
620 from Augraben River and Leiferer Graben River in Italy. The obtained ratios of Cd, Cu, Mn
621 and Zn were similar to those in our research, whereas Cr and Pb appeared to be less bioavailable

622 in our study. Further, Filipović Marijić and Raspor (2012), reported comparable ratios to those
623 found in the present research for European chub ($Zn > Cd \geq Cu > Fe \geq Mn$). Prior studies have
624 consistently shown that fish intestines contain very high levels of Zn, serving as an important
625 storage location for Zn in many fish species (Dallinger and Kautzky, 1985; Rajkowska and
626 Protasowicki, 2013). In addition, it is also important to consider the potential influence of thiol-
627 containing proteins, which can be found in even higher quantities in fish intestines compared
628 to liver and gills, as indicated in previous studies on various fish species (Roesijadi et al., 2009;
629 Filipović Marijić and Raspor, 2010), including brown trout from the Krka River (Mijošek et
630 al., 2019). Given the high affinities of Zn and Hg for thiol-containing proteins like
631 metallothioneins (MTs) (Roesijadi et al., 2009), it is possible that MTs may serve as potential
632 binders of Zn and Hg, leading to their substantially higher intestinal concentrations compared
633 to GC. Further, Hg, but also Rb and Se, are among rare elements for which biomagnification
634 in food webs is suggested (Campbell et al., 2005; Córdoba-Tovar et al., 2022). High
635 accumulation and absorption of K in the intestine in comparison with other macroelements has
636 also been previously described by Bucking and Wood (2006). Also, accumulation of K, Se,
637 and Zn over time is probably related to their important biological roles (e.g. as a part of different
638 enzymes, metallothioneins or K^+ channels).

639 However, it is consistently challenging to make a clear distinction between the uptake
640 through the alimentary tract (dietborne uptake) and absorption through the gills (waterborne
641 uptake) in the field studies. Moreover, the composition and metal(loid) content of fish diet are
642 extremely heterogenous, making it impossible to establish a direct link between a specific food
643 source and the fish.

644

645 **4. Conclusions**

646 This study provided information on total concentrations of 22 metal(loid)s in the intestine
647 of brown trout *S. trutta*, fish gut content, and acanthocephalan species *D. truttae*.
648 Concentrations of As, Ba, Cr, Hg, Mo, and V in parasites provide the first-ever information on
649 this endemic acanthocephalan species, as well as metal(loid)s in GC of brown trouts, serving
650 as an indication of metal intake through food sources. Although often neglected in fish ecology
651 and ecotoxicology, parasites were integrated in the study to achieve more complete
652 understanding of fish biology and to realize influence of parasitic infections on fish responses.
653 Acanthocephalans and GC were also applied to assess possible bioavailability of metal(loid)s
654 to fish intestinal tissue and intestinal parasites.

655 Great efficiency of metal(loid) accumulation in acanthocephalans was confirmed for
656 many elements, especially potentially toxic ones such as Pb, Cd, and Tl. With few exceptions,
657 concentrations of various elements were found to be the highest at different sites: As, Ba, Ca,
658 Cu, Fe, Pb, Se, and Zn at KRK; Cd, Cs, Rb, and Tl at KRS; and Hg, Mn, Mo, Sr, and V at
659 KNP. These patterns corresponded only partially to metal(loid) exposure through water and/or
660 sediments, and indicated that differences in metal(loid) bioaccumulation were strongly related
661 to the biology and ecology of host and parasite species, dietary habits, and the presence of
662 acanthocephalans in the fish intestine. Metal(loid) concentrations in GC were generally the
663 highest at KNP, reflecting the spatial distribution of metal(loid)s in river surface sediment.
664 Ratios between metal accumulation in the intestine and GC, and acanthocephalans and GC,
665 indicate potential low bioavailability of As, Ba, Ca, Cr, Cu, Fe, Mn, Pb, Sr, and V from the
666 dietary sources, while high absorption of Tl, Se, Hg, Rb, K and Zn is suggested. However, that
667 should be interpreted with caution since concentrations in GC represent the metal(loid) burden
668 in the moment of sampling, while concentrations in organisms reflect chronic exposure, which
669 might give different response.

670 We observed a biodilution effect associated with increased acanthocephalan mean
671 intensity. This requires further investigations of other species exposed to different levels to
672 evaluate the potential benefits to hosts and parasites. Comparison with previous research in this
673 area revealed an increase of Cd, Cu, Mn, and Zn concentrations, pointing to the still negative
674 influence of the industrial and municipal wastewater outlets on the Krka National Park.

675 According to collected data, wastewater discharges near the border of Krka National Park
676 currently have a moderate impact. However, concerning tendencies of frequently highest
677 metal(loid) concentrations in sediments, GC and occasionally even in organisms from the KNP
678 indicate the need for more thorough and frequent monitoring of this area. Besides water and
679 sediment as potential environmental sources of metal(loid)s, future monitoring programs
680 should always consider the dietary effects of metal(loid) exposure and the presence of
681 acanthocephalan parasites as possible important contribution factors to metal(loid) input,
682 which would enable more reliable interpretations of results in all ecosystems. Based on given
683 data, we recommend for future studies to inspect fish for the presence of other parasites like
684 trematodes as well, while for the analysis of dietary habits, it would be relevant to classify the
685 gut content in herbal and animal material to identify the most common groups of insects and
686 gammarids to possibly evaluate the contribution of each group to metal(loid) uptake in fish.
687 Further, estimation of bioavailable, and thus possibly toxic, metal(loid)s which binding to
688 sensitive biomolecules may actually change their molecular function, structure and/or
689 dynamics in the cell is a next crucial step in metal(loid) risk assessment.

690

691 **5. References**

692 Algül, F., Beyhan, M., 2020. Concentrations and sources of heavy metals in shallow sediments
693 in Lake Bafa, Turkey. *Sci. Rep.* 10, 11782.

694 Brázová, T., Hanzelová, V., Miklisová, D., Šalamún, P., Vidal-Martínez, V.M., 2015. Host-
695 parasite relationships as determinants of heavy metal concentrations in perch (*Perca fluviatilis*)
696 and its intestinal parasite infection. *Ecotoxicol. Environ. Saf.* 122, 551–556.

697 Bucking, C., Wood, C.M., 2006. Gastrointestinal processing of Na⁺, Cl⁻, and K⁺ during
698 digestion: implications for homeostatic balance in freshwater rainbow trout. *Am. J. Physiol.*
699 *Regul. Integr. Comp. Physiol.* 291(6), 1764-1772.

700 Campbell, L.M., Fisk, A.T., Wang, X., Köck, G., Muir, D.C.G., 2005. Evidence for
701 biomagnification of rubidium in freshwater and marine food webs. *Can. J. Fish. Aquat. Sci.*
702 62(5), 1161-1167.

703 Chon, H.-S., Ohandja, D.-G., Voulvoulis, N., 2012. The role of sediments as a source of metals
704 in river catchments. *Chemosphere* 88, 1250–1256.

705 Clearwater, S.J., Baskin, S.J., Wood, C.M., McDonald, D.G., 2000. Gastrointestinal uptake
706 and distribution of copper in rainbow trout. *J. Exp. Biol.* 203, 2455–2466.

707 Córdoba-Tovar, L., Marrugo-Negrete, J., Barón, P.R., Díez, S., 2022. Drivers of
708 biomagnification of Hg, As and Se in aquatic food webs: A review. *Environ. Res.* 204, 112226.

709 Creighton, N., Twining, J., 2010. Bioaccumulation from food and water of cadmium, selenium
710 and zinc in an estuarine fish, *Ambassis jacksoniensis*. *Mar. Pollut. Bull.* 60, 1815–1821.

711 Cukrov, N., Cmuk, P., Mlakar, M., Omanović, D., 2008. Spatial distribution of trace metals in
712 the Krka River, Croatia. An example of the self-purification. *Chemosphere* 72 (10), 1559–
713 1566.

714 Dallinger, R., Kautzky, H., 1985. The importance of contaminated food for the uptake of heavy
715 metals by rainbow trout (*Salmo gairdneri*): a field study. *Oecologia* 67, 82–89.

716 Debeljak, L., 1986. The nutrition of brown trout (*Salmo fario*) in Bager reservoir and Lepenica
717 Stream. J. Ichthos. 3, 1-7.

718 Dragun, Z., Filipović Marijić, V., Krasnići, N., Ivanković, D., Valić, D., Žunić, J., Kapetanović,
719 D., Vardić Smrzlić, I., Redžović, Z., Grgić, I., Erk, M., 2018. Total and cytosolic concentrations
720 of twenty metals/metalloids in the liver of brown trout *Salmo trutta* (Linnaeus, 1758) from the
721 karstic Croatian river Krka. Ecotoxicol. Environ. Saf. 147, 537–549.

722 Duarte, G.S.C., Lehun, A.L., Leite, L.A.R., Consolin-Filho, N., Bellay, S., Takemoto, R.M.,
723 2020. Acanthocephalans parasites of two Characiformes fishes as bioindicators of cadmium
724 contamination in two neotropical rivers in Brazil. Sci. Total Environ. 738, 140339.

725 Farkas, A., Salánki, J., Specziár, A., 2003. Age- and size-specific patterns of heavy metals in
726 the organs of freshwater fish *Abramis brama* L. populating a low-contaminated site. Water
727 Res. 37, 959-964.

728 Filipović Marijić, V., Raspor, B., 2010. The impact of the fish spawning on metal and protein
729 levels in gastrointestinal cytosol of indigenous European chub. Comp. Biochem. Physiol. C
730 708, 133–138.

731 Filipović Marijić V., Raspor, B. 2012. Site-specific gastrointestinal metal variability in relation
732 to the gut content and fish age of indigenous European chub from the Sava River. Water Air
733 Soil Pollut. 223, 4769–4783

734 Filipović Marijić, V., Vardić Smrzlić, I., Raspor, B., 2013. Effect of acanthocephalan infection
735 on metal, total protein and metallothionein concentrations in European chub from a Sava River
736 section with low metal contamination. Sci. Total Environ. 463–464,772–780.

737 Filipović Marijić, V., Kapetanović, D., Dragun, Z., Valić, D., Krasnići, N., Redžović, Z., Grgić,
738 I., Žunić, J., Kružlicová, D., Nemeček, P., Ivanković, D., Vardić Smrzlić, I., Erk, M., 2018.

739 Influence of technological and municipal wastewaters on vulnerable karst riverine system,
740 Krka River in Croatia. *Environ. Sci. Pollut. Res.* 25, 4715–4727.

741 Filipović Marijić, V., Mijošek, T., Dragun, Z., Retzmann, A., Zitek, A., Prohaska, T., Bačić,
742 N., Redžović, Z., Grgić, I., Krasnići, N., Valić, D., Kapetanović, D., Žunić, J., Ivanković, D.,
743 Vardić Smrzlić, I., Erk, M., 2022. Application of fish calcified structures as indicators of metal
744 exposure in the freshwater ecosystem, *Environments* 9, 14.

745 Fiket, Ž., Mikac, N., Kniewald, G., 2017. Mass fractions of forty-six major and trace elements,
746 including rare earth elements, in sediment and soil reference materials used in environmental
747 studies. *Geostand. Geoanal. Res.* 41, 123–135.

748 Giguère, A., Campbell, P. G. C., Hare, L., McDonald, D. G., Rasmussen, J. B., 2004. Influence
749 of lake chemistry and fish age on cadmium, copper, and zinc concentrations in various organs
750 of indigenous yellow perch (*Perca flavescens*). *Can. J. Fish. Aquat. Sci.* 61(9), 1702–1716.

751 Greenaway, P., 2008. Calcium balance and moulting in the Crustacea. *Biol. Rev. Cambridge*
752 *Philos. Soc* 60, 425-454.

753 Hassanine, R., Al -Hasawi, Z., Hariri, M., Touliabah, H., 2018. *Sclerocollum saudii*
754 (Acanthocephala: Cavisomidae) as a sentinel for heavy-metal pollution in the Red Sea. *J.*
755 *Helminthol.* 93, 177-186.

756 Hassanine, R., Al-Hasawi, Z., 2021. Acanthocephalan Worms Mitigate the Harmful Impacts
757 of Heavy Metal Pollution on Their Fish Hosts. *Fishes* 6, 49.

758 HRN EN 14011, 2005. Fish sampling by electric power (In Croatian). Croatian Standard
759 Institute, Zagreb.

760 Kara, C., Alp, A., 2005. Feeding habits and diet composition of brown trout (*Salmo trutta*) in
761 the upper streams of river Ceyhan and river Euphrates in Turkey, Turk J. Vet. Anim. Sci. 29,
762 417-428.

763 Lapointe, D., Couture, P., 2009. Influence of the route of exposure on the accumulation and
764 subcellular distribution of nickel and thallium in juvenile fathead minnows (*Pimephales*
765 *promelas*). Arch. Environ. Contam. Toxicol. 57, 571–580.

766 Leite, L.A.R., Agostinho, B.N., Oliveira, S.L.P., Pedreira Filho, W.D.R., de Azevedo, R.K.,
767 Abdallah, V.D., 2023. Trace metal accumulation is infrapopulation-dependent in
768 acanthocephalans parasites of the white mullet (*Mugil curema*) from an estuarine environment
769 of southeastern Brazil coast. Mar. Pollut. Bull. 194 (Pt. B), 115374.

770 Mehana, E.E., Khafaga, A.F., Elblehi, S.S., Abd El-Hack, M.E., Naiel, M.A.E., Bin-Jumah,
771 M., Othman, S.I., Allam, A.A., 2020. Biomonitoring of Heavy Metal Pollution Using
772 Acanthocephalans Parasite in Ecosystem: An Updated Overview. Animals (Basel) 10, 811.

773 Mijošek, T., Filipović Marijić, V., Dragun, Z., Ivanković, D., Krasnići, N., Erk, M., Gottstein,
774 S., Lajtner, J., Sertić Perić, M., Matoničkin Kepčija, R., 2019. Comparison of electrochemically
775 determined metallothionein concentrations in wild freshwater salmon fish and gammarids and
776 their relation to total and cytosolic metal levels. Ecol. Ind. 105, 188–198.

777 Mijošek, T., Filipović Marijić, V., Dragun, Z., Ivanković, D., Krasnići, N., Erk, M., 2022.
778 Efficiency of metal bioaccumulation in acanthocephalans, gammarids and fish in relation to
779 metal exposure conditions in a karst freshwater ecosystem. J. Trace. Elem. Med. Biol. 73,
780 127037.

781 Mijošek, T., Kljaković-Gašpić, Z., Kralj, T., Valić, D., Redžović, Z., Šariri, S., Karamatić, I.,
782 Filipović Marijić, V., 2023. Spatial and temporal variability of dissolved metal(loid)s in water

783 of the karst ecosystem: consequences of long-term exposure to wastewaters. Environ. Technol.
784 Innov. 32, 103254.

785 Molbert, N., Alliot, F., Leroux-Coyau, M., Mèdoc, V., Biard, C., Meylan, C.S., Jacquin, L.,
786 Santos, R., Goutte, A., 2020. Potential benefits of acanthocephalan parasites for chub hosts in
787 polluted environments, Environ. Sci. Technol. 54, 5540–5549.

788 Nachev, M., Sures, B., 2016. Seasonal profile of metal accumulation in the acanthocephalan
789 *Pomphorhynchus laevis*: a valuable tool to study infection dynamics and implications for metal
790 monitoring, Parasit. Vectors 9, 300–308.

791 Nikolić, D., Skorić, S., Janković, S., Hegediš, A., Djikanović, V., 2021. Age-specific
792 accumulation of toxic metal(loid)s in northern pike (*Esox lucius*) juveniles. Environ. Monit.
793 Assess. 193, 229

794 NN 55, 2013. Ordinance on the Protection of Animals Used for the Scientific Purposes
795 [Pravilnik o zaštiti životinja koje se koriste u znanstvene svrhe].

796 Ojo, A. A., Wood, C. M., 2007. In vitro analysis of the bioavailability of six metals via the
797 gastro-intestinal tract of the rainbow trout (*Oncorhynchus mykiss*). Aquat. Toxicol. 83, 10–23.

798 Padilla, I.Y., Vesper, D.J., 2018. Fate, transport, and exposure of emerging and legacy
799 contaminants in karst systems: State of knowledge and uncertainty. In: White, W., Herman, J.,
800 Herman, E., Rutigliano, M. (Eds.), Karst Groundwater Contamination and Public Health. In:
801 Advances in Karst Science, Springer, Cham.

802 Rajkowska, M., Protasowicki, M., 2013. Distribution of metals (Fe, Mn, Zn, Cu) in fish tissues
803 in two lakes of different trophy in Northwestern Poland. Environ. Monit. Assess. 185, 3493-
804 3502.

805 Roesijadi, G., Rezvankhah, S., Perez-Matus, A., Mittelberg, A., Torruellas, K., Van Veld, P.A.,
806 2009. Dietary cadmium and benzo(a)pyrene increased intestinal metallothionein expression in
807 the fish *Fundulus heteroclitus*. Mar. Environ. Res. 67 (1), 25–30.

808 Salati, S., Moore, F., 2010. Assessment of heavy metal concentration in the Khoshk River water
809 and sediment, Shiraz, Southwest Iran. Environ. Monit. Assess. 164, 677–689.

810 Sertić Perić, M., Matoničkin Kepčija, R., Miliša, M., Gottstein, S., Lajtner, J., Dragun, Z.,
811 Filipović Marijić, V., Krasnići, N., Ivanković, D., Erk, M., 2018. Benthos-drift relationships as
812 proxies for the detection of the most suitable bioindicator taxa in flowing waters – a pilot-study
813 within a Mediterranean karst river. Ecotoxicol. Environ. Saf. 163, 125–135.

814 Sures, B., Steiner, W., Rydlo, M., Taraschewski, H., 1999. Concentrations of 17 elements in
815 the Zebra Mussel (*Dreissena polymorpha*), in different tissues of perch (*Perca fluviatilis*), and
816 in perch intestinal parasites (*Acanthocephalus lucii*) from the subalpine lake Mondsee, Austria.
817 Environ. Toxicol. Chem. 18, 2574-2579.

818 Sures, B., 2002. Competition for minerals between *Acanthocephalus lucii* and its definitive
819 host perch (*Perca fluviatilis*). Int. J. Parasitol 32, 1117–1122.

820 Sures, B., Nachev, M., Selbach, C., Marcogliese, D. J., 2017. Parasite responses to pollution:
821 what we know and where we go in ‘Environmental Parasitology’, Parasit. Vectors 10, 65–83.

822 Šariri, S., Valić, D., Kralj, T., Cvetković, Ž., Mijošek, T., Redžović, Z., Karamatić, I., Filipović
823 Marijić, V., 2024. Long-term and seasonal trends of water parameters in the karst riverine
824 catchment and general literature overview based on CiteSpace. Environ. Sci. Pollut. Res. 31,
825 3887-3901.

826 Timi, J.T., Poulin, R., 2020. Why ignoring parasites in fish ecology is a mistake, Int. J.
827 Parasitol. 50, 755–761.

828 Vardić Smrzlić, I., Valić, D., Kapetanović, D., Dragun, Z., Gjurčević, E., Četković, H.,
829 Teskeredžić E., 2013. Molecular characterisation and infection dynamics of *Dentitruncus*
830 *truttae* from trout (*Salmo trutta* and *Oncorhynchus mykiss*) in Krka River, Croatia, Vet.
831 Parasitol. 197, 604–613.

832

833

834

835

836

837

838

839

840

841

842

843

844

845

846

847

848 **Acknowledgements**

849 This research has been supported by the Croatian Science Foundation under the project
850 “Integrated evaluation of aquatic organism responses to metal exposure: gene expression,
851 bioavailability, toxicity and biomarker responses” (BIOTOXMET; IP-2020-02-8502). The
852 authors would like to acknowledge Dr. Zuzana Redžović, Dr. Damir Valić and Dr. Tomislav
853 Kralj for their valuable help in the field work and Dr. Tatjana Ocrt and Dr. Ankica Sekovanić
854 for their valuable help in the ICP-MS measurements.

855

856 **Figure captions:**

857 **Figure 1.** Study area with marked sampling sites on the Krka River (1 – Krka River source; 2
858 – Krka River near the Town of Knin; 3 – location in the national park near the Brijun Lake)

859

860 **Figure 2.** Concentrations (mg kg^{-1} or $\mu\text{g kg}^{-1}$ on wet mass basis) of six elements in the intestine
861 of brown trout *S. trutta* and acanthocephalans *D. truttae* from three sites of the Krka River in
862 two sampling campaigns (spring and autumn) that showed enhanced accumulation at the
863 contaminated site, KRK. Different letters refer to significant differences of metal(loid)
864 concentrations among three locations in spring (capital letters) and autumn (small letters),
865 while statistically significant differences between two seasons at each sampling site are marked
866 with asterisk (*).

867

868 **Figure 3.** Concentrations ($\mu\text{g kg}^{-1}$ on wet mass basis) of four elements in the intestine of brown
869 trout *S. trutta* and acanthocephalans *D. truttae* from three sites of the Krka River in two
870 sampling campaigns (spring and autumn) that showed enhanced accumulation at the reference
871 site, KRS. Different letters refer to significant differences of metal(loid) concentrations among
872 three locations in spring (capital letters) and autumn (small letters), while statistically
873 significant differences between two seasons at each sampling site are marked with asterisk (*).

874

875 **Figure 4.** Concentrations ($\mu\text{g kg}^{-1}$ on wet mass basis) of five elements in the intestine of brown
876 trout *S. trutta* and acanthocephalans *D. truttae* from three sites of the Krka River in two
877 sampling campaigns (spring and autumn) that showed enhanced accumulation at the site in the
878 national park, KNP. Different letters refer to significant differences of metal(loid)
879 concentrations among three locations in spring (capital letters) and autumn (small letters),

880 while statistically significant differences between two seasons at each sampling site are marked
881 with asterisk (*).

882

883 **Figure 5.** Concentrations (mg kg^{-1} or $\mu\text{g kg}^{-1}$ on wet mass basis) of five elements in the intestine
884 of brown trout *S. trutta* and acanthocephalans *D. truttae* from three sites of the Krka River in
885 two sampling campaigns (spring and autumn) that showed unclear trends of accumulation.
886 Different letters refer to significant differences of metal(loid) concentrations among three
887 locations in spring (capital letters) and autumn (small letters), while statistically significant
888 differences between two seasons at each sampling site are marked with asterisk (*).

889

890