

Mladena Glavaš, Agata Gitlin – Domagalska, Natalia Ptaszyńska, Dominika Starego, Krzysztof Rolka

Department of molecular biochemistry, University of Gdańsk, Poland

E-mail: mladena.glavas@ug.edu.pl; agata.domagalska@ug.edu.pl

INTRODUCTION

PEPTIDES

crucial role in all living organisms [1]

application

- hormones, antibodies, neurotransmitters [1]
- development of some diseases [2]
- functional materials [3]
- pharmaceuticals [4]

shortcomings

- inability to cross intestinal epithelial barrier
- limited stability toward proteolysis [4][5]

solution

lipophilic prodrug charge masking (LPCM) [6] → positive charged group is masked by lipophilic alkyl residues, the compound with increased lipophilicity and better membrane permeability

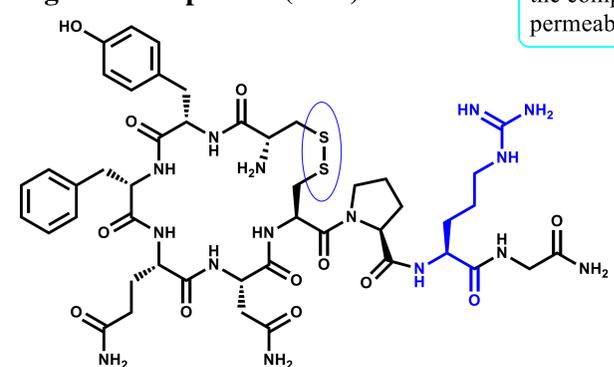
Function:

- antidiuretic [8]
- increases the blood pressure in septic shock
- maintains cardiovascular homeostasis [8a,8c]

Shortcomings:

- short biological half-life
- lack of specificity for receptors [9]
- may cause hyponatremia and decreasing of cardiac output
- increases bilirubin level in blood [10]

Arginine vasopressin (AVP)



Cys-Tyr-Phe-Gln-Asn-Cys-Pro-Arg-Gly-NH₂

SYNTHESIS

Synthesis of modified arginine started with optimization of reaction conditions (Table 1) for preparation of guanidynylating reagents **1** – **7** (Figure 1) [11]. All reactions were conducted using butyl chloroformate and the best results were obtained using triethylamine (TEA) as base in dimethylformamide (DMF) on 0 °C (entry 3, Table 1). Reaction was performed on gram-scale, and the product was isolated in 84 % (entry 8, Table 1). Reactions with other chloroformates (ethyl-, propyl-, hexyl-, octyl-, decyl- and dodecyl-) were performed under the same reaction conditions on the gram-scale and the products **1** – **7** were isolated in 35 – 94 %.

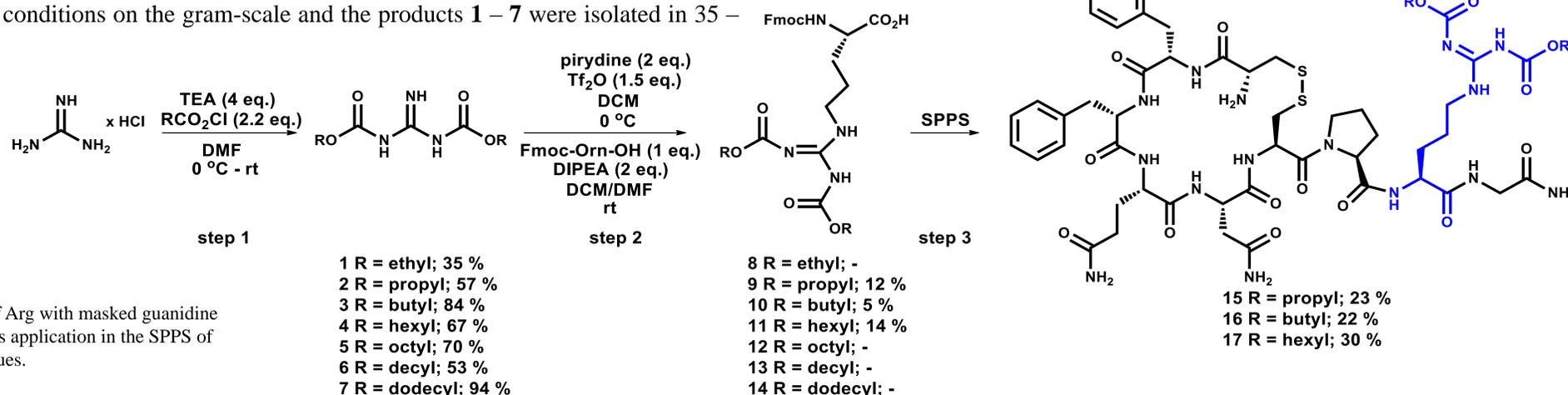


Figure 1. Synthesis of Arg with masked guanidine group and its application in the SPPS of AVP analogues.

AIM

- preparation of arginine (Arg) building blocks with masked guanidine group
- SPPS of AVP analogues with increased lipophilicity

Table 1. Optimization of reaction conditions for step 1.

Entry	m (guanidine hydrochloride)/g	base/eq.	solvent/mL	t/°C	chloroformate/2.2 eq.	η/%
1	0.25	TEA/4	ACN/H ₂ O/3/1	0→rt	butyl	13
2	0.10 ^a	TEA/4	ACN/H ₂ O/1.2/1.2	0→rt	butyl	-
3	0.10	TEA/4	DMF/2	0→rt	butyl	76
4	0.10	DIPEA/4	ACN/H ₂ O/1.2/1.2	0→rt	butyl	44
5	0.10	1M NaOH	ACN/H ₂ O/1.2/1.2	0→rt	butyl	-
6	0.10	TEA/4	ACN/H ₂ O/1.2/1.2	40	butyl	-
7	0.10	NaH/4	ACN/H ₂ O/1.2/1.2	0→rt	butyl	70
8	1.00	TEA/4	DMF/20	0→rt	butyl	84

^a DMAP was added in the reaction.

In the next step, compounds **1** – **7** were reacted with ornithine. The reaction was performed simultaneously in two flasks. In the first flask reacted compound **1** – **7**, respectively, with trifluoroacetic anhydride (Tf₂O) in dichloromethane (DCM) with addition of pyridine on 0 °C. In the second flask, reacted Fmoc protected ornithine with diisopropylamide (DIPEA) in DCM/DMF on rt. After 20 minutes of stirring, two parts were connected. Arginine derivatives with masked guanidine group **9** – **11** were isolated in modest yield of 5 – 14 %.

Last part was synthesis of AVP analogues, applying SPPS. Peptides **15** – **17** were obtained in moderate yield of 22 – 30 %. The main difference between peptides is in the group (ethyl-, propyl-, hexyl-) with which the Arg guanidine moiety is masked. The structure of all compounds were confirmed by NMR (1D and 2D), HRMS and HPLC.

CONCLUSION

- guanidynylating reagents **1** – **7** were isolated in moderate to excellent yield of 35 – 94 %, respectively
- only three compounds **9** – **11**, with masked Arg guanidine group were isolated in modest yield of 5 – 14 %
- three AVP analogues **15** – **17** were prepared and isolated in moderate yield of 22 – 30 %
- further research including computational studies is planned as well as the biological testing of prepared peptides

REFERENCES

- [1] G. Koopmanschap, E. Ruijter, R. V. A. Orru, *Beilstein J. Org. Chem.* **10** (2014) 544 – 598.; [2] A. Grauer, B. König, *J. Org. Chem.* **30** (2009) 5099 – 5111.; [3] M. Koshizuka, K. Makino, N. Shimada, *Org. Lett.* **22** (2020) 8658 – 8664.; [4] C. S. Brian Chia, *Int. J. Pept. Res. Ther.* **27** (2021) 1397 – 1418.; [5] E. Lenci, A. Trabocchi, *Chem. Soc. Rev.* **49** (2020) 3262 – 3277.; [6] A. Schumacher – Klinger, et al. *Molecular Pharmaceutics* **15** (2018) 3468 – 3477.; [7] a) C. M. Yea et al., *J. Med. Chem.* **51** (2008) 8124 – 8134.; b) Y. Shimada et al., *Bioorganic Med. Chem.* **14** (2006) 1827 – 1837.; c) Z. Dekan et al., *Chem Sci.* **12** (2021) 4057 – 4062.; [8] a) A. Krag, F. Bendtsen, E. B. Pedersen, N. H. Holstein-Rathlou, S. Møller, *Am. J. Physiol. - Ren. Physiol.* **295** (2008) 295 – 300.; b) V. PLiska, T. Chard, J. Rudinger, *Cat. Eur. J. Endocrinol.* **81** (1976) 474 – 481.; c) K. D. Döhler, M. Meyer, *Best Pract. Res. Clin. Anaesthesiol.* **22** (2008) 335–3502.; [9] R. De Franchis, *Dig. Liver Dis.* **36** (2004) 93 – 100, 2004.; [10] X. Xiao et al., *J. Surg. Res.* **195** (2015) 568 – 579.; [11] *Org. Synth.* **78** (2002) 91.

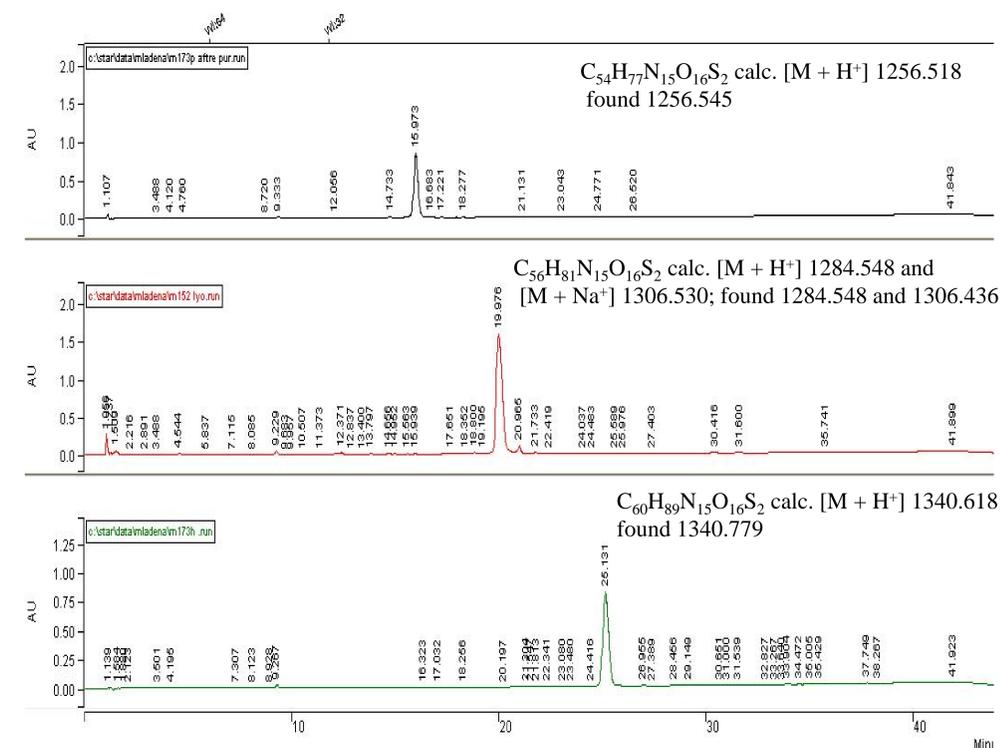


Figure 2. HPLC spectrums of peptides **15**, **16** and **17** and detected values in HRMS.